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The specification, claims, abstract, sequence listing, drawings and appendix I+II as filed with the application on the filing date indicated above.

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Patent- og Varemærkestyrelsen Erhvervsministeriet

TAASTRUP 27 October 1999

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AN IMPROVED METHOD for extracting quantitative information relating to an influence on a cellular response.

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### SUMMARY OF THE INVENTION

The present invention relates to an improved method and tools for extracting quantitative information relating to an influence on a cellular response, in particular an influence caused by contacting or incubating the cell with a substance influencing a cellular response, wherein the cellular response is manifested in redistribution of at least one component in the cell. In particular, the invention relates to an improved method for extracting the quantitative information relating to an influence on an intracellular pathway involving redistribution of at least one component associated with the pathway. The method of the invention may be used as a very efficient procedure for testing or discovering the influence of a substance on a physiological process, for example in connection with screening for new drugs, testing of substances for toxicity, identifying drug targets for known or novel drugs. In particular, the present invention relates to an improved method for parallelisation of the testing procedure so that a large number of substances can be tested simultaneously using commercially available instrumentation. The invention also describes several ways of contacting the cells with a substance influencing a cellular response and modifications made to the actual cells before, during or after contacting the cells with these substances as to improve the applicability and use of the method for extracting quantitative information relating to influence on an intracellular pathway in a highly parallel fashion. Other valuable uses of the method and technology of the invention will be apparent to the skilled person on the basis of the following disclosure. In a particular embodiment of the invention, the present invention relates to a method of detecting intracellular translocation or redistribution of biologically active polypeptides, preferably an enzyme, affecting intracellular processes, and a DNA construct and a cell for use in the method.

Two appendices are included herein, and are considered part of the application. Appendix I, "METHOD AND APPARATUS FOR HIGH DENSITY FORMAT SCREENING FOR BIOACTIVE MOLECULES", is a pending patent application. Appendix II, "CHANGES

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IN INTRACELLULAR cAMP VISUALIZED USING A CAMP-DEPENDENT PROTEIN KINASE-GREEN FLUORESCENT PROTEIN HYBRID", is a manuscript intended for publication.

### 5 BACKGROUND OF THE INVENTION

Intracellular pathways are tightly regulated by a cascade of components that undergo modulation in a temporally and spatially characteristic manner. Several disease states can be attributed to altered activity of individual signalling components (i.e. protein kinases, protein phosphatases, transcription factors). These components therefore render themselves as attractive targets for therapeutic intervention.

Protein kinases and phosphatases are well described components of several intracellular signalling pathways. The catalytic activity of protein kinases and phosphatases are assumed to play a role in virtually all regulatable cellular processes. Although the involvement of protein kinases in cellular signalling and regulation have been subjected to extensive studies, detailed knowledge on e.g. the exact timing and spatial characteristics of signalling events is often difficult to obtain due to lack of a convenient technology.

Novel ways of monitoring specific modulation of intracellular pathways in intact, living cells is assumed to provide new opportunities in drug discovery, functional genomics, toxicology, patient monitoring etc.

- The spatial orchestration of protein kinase activity is likely to be essential for the high degree of specificity of individual protein kinases. The phosphorylation mediated by protein kinases is balanced by phosphatase activity. Also within the family of phosphatases translocation has been observed, e.g. translocation of PTP2C to membrane ruffles [(Cossette et al. 1996)], and likewise is likely to be indicative of phosphatase activity.
- 25 Protein kinases often show a specific intracellular distribution before, during and after activation. Monitoring the translocation processes and/or redistribution of individual protein kinases or subunits thereof is thus likely to be indicative of their functional activity. A connection between translocation and catalytic activation has been shown for

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protein kinases like the diacyl glycerol (DAG)-dependent protein kinase C (PKC), the cAMP-dependent protein kinase (PKA) [(DeBernardi *et al.* 1996)] and the mitogenactivated-protein kinase Erk-1 [(Sano *et al.* 1995)].

Commonly used methods of detection of intracellular localisation/activity of protein kinases and phosphatases are immunoprecipitation, Western blotting and immunocytochemical detection.

Taking the family of diacyl glycerol (DAG)-dependent protein kinase Cs (PKCs) as an example, it has been shown that individual PKC isoforms that are distributed among different tissues and cells have different activator requirements and undergo differential translocation in response to activation. Catalytically inactive DAG-dependent PKCs are generally distributed throughout the cytoplasm, whereas they upon activation translocate to become associated with different cellular components, e.g. plasma membrane [(Farese, 1992),(Fulop Jr. et al. 1995)] nucleus [(Khalil et al. 1992)], cytoskeleton [(Blobe et al. 1996)]. The translocation phenomenon being indicative of PKC activation has been monitored using different approaches: a) immunocytochemistry where the localisation of individual isoforms can be detected after permeabilisation and fixation of the cells [(Khalil et al. 1992)]; and b) tagging all DAG-dependent PKC isoforms with a fluorescently labelled phorbol myristate acetate (PMA) [(Godson et al. 1996)]; and c) chemical tagging of PKC \(\beta\)1 with the fluorophore Cy3 [(Bastiaens & Jovin 1996)] and d) genetic tagging of PKC  $\alpha([Schmidt\ et\ al.\ 1997])$  and of PKC  $\gamma$  and PKC  $\delta([Sakai\ et\ al.\ 1996)]$ . The first method does not provide dynamic information whereas the latter methods will. Tagging PKC with fluorescently labelled phorbol myristate acetate cannot distinguish between different DAG-dependent isoforms of PKC but will label and show movement of all isoforms. Chemical and genetic labelling of specific DAG-dependent PKCs confirmed that they in an isoform specific manner upon activation move to cell periphery or nucleus.

In an alternative method, protein kinase A activity has been measured in living cells by chemical labelling one of the kinase's subunit [(Adams et al. 1991)]. The basis of the methodology is that the regulatory and catalytic subunit of purified protein kinase A is labelled with fluorescein and rhodamine, respectively. At low cAMP levels protein kinase A is assembled in a heterotetrameric form which enables fluorescence resonance energy

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transfer between the two fluorescent dyes. Activation of protein kinase A leads to dissociation of the complex, thereby eliminating the energy transfer. A disadvantage of this technology is that the labelled protein kinase A has to be microinjected into the cells of interest. This highly invasive technique is cumbersome and not applicable to large scale screening of biologically active substances. A further disadvantage of this technique as compared to the presented invention is that the labelled protein kinase A cannot be inserted into organisms/animals as a transgene.

Recently it was discovered that Green Fluorescent Protein (GFP) expressed in many different cell types, including mammalian cells, became highly fluorescent [(Chalfie et al. 1994)]. WO95/07463 describes a cell capable of expressing GFP and a method for detecting a protein of interest in a cell based on introducing into a cell a DNA molecule having DNA sequence encoding the protein of interest linked to DNA sequence encoding a GFP such that the protein produced by the DNA molecule will have the protein of interest fused to the GFP, then culturing the cells in conditions permitting expression of the fused protein and detecting the location of the fluorescence in the cell, thereby localizing the protein of interest in the cell. However, examples of such fused proteins are not provided, and the use of fusion proteins with GFP for detection or quantitation of translocation or redistribution of biologically active polypeptides affecting intracellular processes upon activation, such as proteins involved in signalling pathways, e.g. protein kinases or phosphatases, has not been suggested. WO 95/07463 further describes cells useful for the detection of molecules, such as hormones or heavy metals, in a biological sample, by operatively linking a regulatory element of the gene which is affected by the molecule of interest to a GFP, the presence of the molecules will affect the regulatory element which in turn will affect the expression of the GFP. In this way the gene encoding GFP is used as a reporter gene in a cell which is constructed for monitoring the presence of a specific molecular identity.

Green Fluorescent Protein has been used in an assay for the detection of translocation of the glucocorticoid receptor (GR) [(Carey, KL et al. 1996)]. A GR-S65TGFP fusion has been used to study the mechanisms involved in translocation of the glucocorticoid receptor (GR) in response to the agonist dexamethasone from the cytosol, where it is present in the absence of a ligand, through the nuclear pore to the nucleus where it remains after ligand

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binding. The use of a GR-GFP fusion enables real-time imaging and quantitation of nuclear/cytoplasmic ratios of the fluorescence signal. A similar genetic construct has been used to follow and quantify dexamethasone induced translocation of GR to the nucleus in HeLa cells [(Guiliano, K.A et al. 1997)] in a system called Array Scan<sup>™</sup> (WO 97/45730) designed for automated drug screening. Recently, several other investigators have demonstrated that tagging a specific protein (or part of a protein) involved in an intracellular signalling pathway with GFP provides a new means to measure and quantify the influence of substances on this pathway. The concept has been shown to work both for cytoplasmic to nuclear translocation of the androgen receptor [(Georget V et al. 1997)] and transcription factors such as NF-ATc [(Beals CR et al. 1997)] in analogy with what has already been described for GR above. Another relevant example is a β-arrestin – GFP construct that was shown to report on activation of G-protein coupled receptors by translocating from the cytosol to the plasma membrane [(Barak LS et al. 1997)]. Finally, it has also been demonstrated that attaching GFP to a smaller part of a protein like the pleckstrin homology domain of phospholipase C δ 1 [(Stauffer TP et al. 1998)] and a cysteine-rich domain of PKC y [(Oancea E et al. 1998)] can be used to report on an influence from a substance by quantifying their redistribution within the cells during activation of the specific signalling pathway to which they belong.

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Many currently used screening programmes designed to find compounds that affect protein kinase activity are based on measurements of kinase phosphorylation of artificial or natural substrates, receptor binding and/or reporter gene expression. The interest in fluorescence measurements as the basis for future high-throughput drug screening has however increased dramatically over the last few years [(Silverman L *et al.* 1998)]. Of particular interest to the present invention is a scanning laser imager for rapid screening of fluorescence changes in living cells [(Schroeder K & Neagle B 1996)] currently offered commercially by Molecular Devices, Inc. as the FLIPR<sup>TM</sup>.

#### **DETAILED DESCRIPTION OF THE INVENTION**

The present invention provides an important new dimension in the investigation of cellular systems involving redistribution in that the invention provides quantification of the

redistribution responses or events caused by an influence, typically contact with a chemical substance or mixture of chemical substances, but also changes in the physical environment. The quantification makes it possible to set up meaningful relationships, expressed numerically, or as curves or graphs, between the influences (or the degree of influences) on cellular systems and the redistribution response. This is highly advantageous because, as has been found, the quantification can be achieved in both a fast and reproducible manner, and - what is perhaps even more important - the systems which become quantifiable utilising the method of the invention are systems from which enormous amounts of new information and insight can be derived.

The present screening assays have the distinct advantage over other screening assays, e.g., receptor binding assays, enzymatic assays, and reporter gene assays, in providing a system in which biologically active substances with completely novel modes of action, e.g. inhibition or promotion of redistribution/translocation of a biologically active polypeptide as a way of regulating its action rather than inhibition/activation of enzymatic activity, can be identified in a way that insures very high selectivity to the particular isoform of the biologically active polypeptide and further development of compound selectivity versus other isoforms of the same biologically active polypeptide or other components of the same signalling pathway.

In its broadest aspect, the invention relates to an improved method, with higher throughput compared to previous methods, for extracting quantitative information relating to an influence on a cellular response, the method comprising recording variation, caused by the influence on mechanically intact living cells, in spatially distributed light emitted from a luminophore, the luminophore being present in the cells and being capable of being redistributed in a manner which is related with the degree of the influence, and/or of being modulated by a component which is capable of being redistributed in a manner which is related to the degree of the influence, the association resulting in a modulation of the luminescence characteristics of the luminophore, detecting and recording the spatially distributed light from the luminophore, and processing the recorded variation in the spatially distributed light to provide quantitative information correlating the spatial distribution or change in the spatial distribution to the degree of the influence. In one aspect of the present invention the mechanically intact living cell is permeabilised at some

time after the influence has begun but during or before the actual experimental recording. In another aspect, the present invention relates to an improved method for extracting quantitative information relating to an influence on a cellular response, the method comprising recording variation, caused by the influence on permeabilised living cells, in spatially distributed light emitted from a luminophore, the luminophore being present in the cells and being capable of being redistributed in a manner which is related with the degree of the influence, and/or of being modulated by a component which is capable of being redistributed in a manner which is related to the degree of the influence, the association resulting in a modulation of the luminescence characteristics of the luminophore, detecting and recording the spatially distributed light from the luminophore, and processing the recorded variation in the spatially distributed light to provide quantitative information correlating the spatial distribution or change in the spatial distribution to the degree of the influence. In a preferred embodiment of the invention the luminophore, which is present in the cells, is capable of being redistributed by modulation of an intracellular pathway, in a manner which is related to the redistribution of at least one component of the intracellular pathway. In another preferred embodiment of the invention, the luminophore is a fluorophore.

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In the invention the cell and/or cells are mechanically intact and alive throughout the experiment. In another embodiment of the invention, the cells are fixed at a point in time after the application of the influence at which the response has been predetermined to be significant, and the recording is made at an arbitrary later time. In another embodiment the cell and/or cells are mechanically intact and alive throughout the experiment but are mechanically or chemically disrupted or permeabilised as the initial step of experimental analysis. In another aspect of the invention the cells have their plasma membrane permanently and stably permeabilised before the initiation of the experiment in such a way that the plasma membrane stays permeable during the experiment. This allows the components of intracellular pathways to be contacted by substances that are not normally permeating the cell plasma membrane such as peptides, proteins and hydrophilic organic compounds.

The mechanically intact or permeabilised living cells could be selected from the group consisting of fungal cells, such as yeast cells; invertebrate cells including insect cells; and

vertebrate cells, such as mammalian cells. These cells are incubated at a temperature of 30°C or above, preferably at a temperature of from 32°C to 39°C, more preferably at a temperature of from 35°C to 38°C, and most preferably at a temperature of about 37°C during the time period over which the influence is observed. In one aspect of the invention the mechanically intact or permeabilised living cell is part of a matrix of identical or non-identical cells. In one embodiment of the invention the cells comprise a group or groups of cells contained within a spatial limitation or spatial limitations. In one embodiment, the cells comprise multiple groups of cells that are qualitatively the same but subjected to different influences. In another embodiment, the cells comprise multiple groups of cells that are qualitatively different but subjected to the same influence.

In one embodiment of the invention the spatial limitations are domains defined on a substrate on which the cells are present. The spatial limitations may be arranged in one or more arrays on a common carrier. The spatial limitations may be wells in a plate of microtiter type, such that 96, 384, 864 and 1536 wells are situated on the common carrier. In another embodiment the spatial limitations are wells in a plate of a format different from the microtiter type. In one embodiment of the invention the domains are established by the presence of the cells on the substrate in a pattern that defines the domains. In another aspect of the invention, the domains are instead established by the spatial pattern or array of the influence or influences as it/they are applied to or contacted by the cells. This aspect is thoroughly described in Appendix I. Briefly, in this aspect of the invention the mechanically intact or permeabilised living cells are part of a continuous or discontinuous sheet of cells cultured on an optically clear flat surface optimised or not for cell culture. The optically clear and flat surface may be a porous membrane that may allow cellular processes to grow through the membrane pores and may allow directed capillary flow of fluid through the pores.

A cell used in the present invention should contain a nucleic acid construct encoding a fusion polypeptide as defined herein and be capable of expressing the sequence encoded by the construct. The cell is a eukaryotic cell selected from the group consisting of fungal cells, such as yeast cells; invertebrate cells including insect cells; vertebrate cells such as mammalian cells. The preferred cells are mammalian cells.

In another aspect of the invention the cells could be from an organism carrying in at least one of its component cells a nucleic acid sequence encoding a fusion polypeptide as defined herein and be capable of expressing said nucleic acid sequence. The organism is selected from the group consisting of unicellular and multicellular organisms, such as a mammal.

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The luminophore is the component that allows the redistribution to be visualised and/or recorded by emitting light in a spatial distribution related to the degree of influence. The term redistribution is intended to cover all aspects of a change in spatial location, such as a translocation of the luminophore or other components. In one embodiment of the invention, the luminophore is capable of being redistributed in a manner that is physiologically relevant to the degree of the influence. It should be understood that redistribution. In another embodiment, the luminophore is capable of associating with a component that is capable of being redistributed in a manner that is physiologically relevant to the degree of the influence. In another embodiment, a correlation between the redistribution of the luminophore and the degree of the influence could be determined experimentally. In a preferred aspect of the invention, the luminophore is capable of being redistributed in substantially the same manner as the at least one component of an intracellular pathway. In another embodiment of the invention, the luminophore is capable of being quenched upon spatial association with a component that is redistributed by modulation of the pathway, the quenching being measured as a change in the intensity of the luminescence. In another embodiment of the invention, the luminophore is stationary but may have a certain spatial distribution, and interacts with at least one component that is capable of being redistributed in a manner which is physiologically relevant to the degree of the influence, in such a way that one or more luminescence characteristics of the luminophore is/are modulated as the component moves closer to, or farther from, the luminophore.

The luminophore could be a fluorophore. In a preferred embodiment of the invention, the luminophore is a polypeptide encoded by and expressed from a nucleotide sequence harboured in the cells. The luminophore could be a hybrid polypeptide comprising a fusion of at least a portion of each of two polypeptides one of which comprises a luminescent polypeptide and the other one of which comprises a biologically active polypeptide, as

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defined herein.

The luminescent polypeptide could be a GFP as defined herein or could be selected from the group consisting of green fluorescent proteins having the F64L mutation as defined herein such as F64L-GFP, F64L-Y66H-GFP, F64L-S65T-GFP, and EGFP. The GFP could be N- or C-terminally tagged, optionally via a peptide linker, to the biologically active polypeptide or a part or a subunit thereof. The fluorescent probe could be a component of an intracellular signalling pathway. The probe is coded for by a nucleic acid construct.

The pathway of investigation in the present invention could be an intracellular signalling pathway.

In a preferred embodiment of the invention, the influence could be contact between the group or groups of mechanically intact or permeabilised living cells and a chemical substance, and/or incubation of the group or groups of mechanically intact or permeabilised living cells with a chemical substance in solution. In one aspect of the invention that is thoroughly described in Appendix I, the chemical substances are attached to an underlying matrix. In this aspect, the chemical substances may also be produced and secreted from, or attached to the plasma membrane surfaces of, a sheet of genetically engineered cells. In this aspect of the invention the chemical substances may also have been separated two-dimensionally in a non-denaturing gel using electrophoresis and the gel is directly put in close proximity or direct contact with the mechanically intact or permeabilised living cells so that the chemical substances can contact the cells through diffusion or convection.

The influence will modulate the intracellular processes. In one aspect the modulation could be an activation of the intracellular processes. In another aspect the modulation could be a deactivation of the intracellular processes. In yet another aspect, the influence could inhibit or promote the redistribution without directly affecting the metabolic activity of the component of the intracellular processes.

In one embodiment the invention is used to establish a dose-response relationship for one or many chemical substances. In one embodiment the invention is used as a basis for a screening program, where the effect of unknown influences such as a compound library,

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can be compared to influence of known reference compounds under standardised conditions.

In addition to the intensity, there are several parameters of fluorescence or luminescence that can be modulated by the effect of the influence on the underlying cellular phenomena, and can therefore be used in the invention. Some examples are resonance energy transfer, fluorescence lifetime, polarisation, and wavelength shift. Each of these methods requires a particular kind of filter in the emission light path to select the component of the light desired and reject other components. The recording of property of light could be in the form of an ordered array of values such as a CCD array or a vacuum tube device such as a vidicon. In addition, the translational mobility, or freedom of movement, of the luminophore attached to the protein of interest can be an important property affected by the influence on the underlying cellular phenomena, and can therefore be used in he invention.

In one embodiment of the invention, the spatially distributed light emitted by a luminophore is detected by a change in the resonance energy transfer between the luminophore and another luminescent entity capable of delivering energy to the luminophore, each of which has been selected or engineered to become part of, bound to or associated with particular components of the intracellular pathway. In this embodiment, either the luminophore or the luminescent entity capable of delivering energy to the luminophore undergoes redistribution in response to an influence. The resonance energy transfer would be measured as a change in the intensity of emission from the luminophore, preferably sensed by a single channel photodetector that responds only to the average intensity of the luminophore in a non-spatially resolved fashion.

In one embodiment of the invention, the spatially distributed light emitted by a luminophore includes the case of uniform spatial distribution of the light.

In one aspect of the invention, the luminophore is a fluorophore which redistributes through a non-homogenous excitation light field, resulting in a change in the intensity of the light emitted from the luminophore as a result of the change in the amount of excitation light intensity at different points in the field.

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In one embodiment of the invention, the recording of the spatially distributed light could be made at a single point in time after the application of the influence. In another embodiment, the recording could be made at two points in time, one point being before, and the other point being after the application of the influence. The result or variation is determined from the change in fluorescence compared to the fluorescence measured prior to the influence or modulation. In another embodiment of the invention, the recording could be performed at a series of points in time, in which the application of the influence occurs at some time after the first time point in the series of recordings, the recording being performed, e.g., with a predetermined time spacing of from 0.1 seconds to 1 hour, preferably from 1 to 60 seconds, more preferably from 1 to 30 seconds, in particular from 1 to 10 seconds, over a time span of from 1 second to 12 hours, such as from 10 seconds to 12 hours, e.g., from 10 seconds to one hour, such as from 60 seconds to 30 minutes or 20 minutes. The result or variation is determined from the change in fluorescence over time. The result or variation could also be determined as a change in the spatial distribution of the fluorescence over time.

In one embodiment the recording comprises a time series of total luminescence of the cells of one or several of the spatial limitations. In one embodiment the signal from all of the spatial limitations, one at a time, is measured by a recording being made in the individual spatial limitations by means of an apparatus to sequentially position each one of the limitations in the field of view of the detector and repeating the positioning and measurement process until all of the spatial limitations have been measured. The detector may be a photomultiplier tube. In a preferred embodiment of the present invention more than one spatial limitation is measured simultaneously. This may be done by means of a one- or two-dimensional array detector, whereby the multiple spatial limitations are imaged onto the array detector such that discrete subsets of the detecting units (pixels) in the array detector measure the signal from one and only one of the multiple spatial limitations, the signal from any one spatial limitation being the combined signal from those pixels that receive the image from one of the spatial limitations. This array detector may be a linear diode array, a video camera (according to any present or future standards and definitions of image acquisition and transmission) or a charge transfer device such as a charge-coupled device (CCD). In one embodiment the recording of signal requires

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illumination of the multiple spatial limitations to excite the luminophores so that they emit light. In one embodiment all of the spatial limitations are simultaneously illuminated during the measurement. In another embodiment the spatial limitations are singly illuminated only during the time in which they are being measured. In a preferred embodiment the illumination is provided by a laser that is scanned in a raster fashion over some or all of the spatial limitations being measured. The scanning may take place at a rate that is substantially faster than the measurement process such that the illumination appears to the measurement process to be continuous in time and spatially uniform over the region being measured.

The recording of spatially distributed luminescence emitted from the luminophore is 10 performed by an apparatus for measuring the distribution of fluorescence in the cells, and thereby any change in the distribution of fluorescence in the cells, which includes at a minimum the following component parts: (a) a light source, (b) a method for selecting the wavelength(s) of light from the source which will excite the luminescence of the 15 luminophore, (c) a device which can rapidly block or pass the excitation light into the rest of the system, (d) a series of optical elements for conveying the excitation light to the specimen, collecting the emitted fluorescence in a spatially resolved fashion, and forming an image from this fluorescence emission (or another type of intensity map relevant to the method of detection and measurement), (e) a bench or stand which holds the container of the cells being measured in a predetermined geometry with respect to the series of optical 20 elements, (f) a detector to record the spatially resolved fluorescence in the form of an image, (g) a computer or electronic system and associated software to acquire and store the recorded images, and to compute the degree of redistribution from the recorded images.

In a preferred embodiment of the invention the apparatus system is automated. In one embodiment the components in d and e mentioned above comprise a fluorescence microscope. In one embodiment the component in f mentioned above is a CCD camera. In one embodiment the component in f mentioned above is an array of photomultiplier tubes/devices.

In one embodiment the image is formed and recorded by an optical scanning system.

In one embodiment the optical scanning system is used to illuminate the bottom of a plate

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of microtiter type so that a time-resolved recording of changes in luminescence or fluorescence can be made from all spatial limitations simultaneously.

In a preferred embodiment the actual luminescence or fluorescence measurements are made in a FLIPR<sup>TM</sup> instrument, commercially available from Molecular Devices, Inc.

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In one embodiment of the invention the actual fluorescence measurements are made in a standard type of fluorometer for plates of microtiter type (fluorescence plate reader).

In one embodiment a liquid addition system is used to add a known or unknown compound to any or all of the cells in the cell holder at a time determined in advance. Preferably, the liquid addition system is under the control of the computer or electronic system. Such an automated system can be used for a screening program due to its ability to generate results from a larger number of test compounds than a human operator could generate using the apparatus in a manual fashion.

The methods whereby the detector layer of cells are physically contacted by the compounds can also be of another conceptual type where the compounds are delivered to the cells through a porous membrane by convection/diffusion or by directly contacting compounds attached to an inorganic or organic support (such as glass, plastic or the plasma membrane of intact living cells) with the cells. These methods are thoroughly described in Appendix I, but are also outlined in the following paragraphs.

In one aspect of the present invention where the detector layer of cells is a continuous or discontinuous sheet of cells without any separation into test units or wells. The compounds are printed onto a nonabsorbent sheet of porous material as a solution in solvent and allowed to dry. This printed sheet of compounds then defines the test pattern for the experiment as it is brought down in close proximity to or in direct contact with the underlying detector layer of cells. The compounds, now dissolved by the fluid layer on the cells, is brought in contact with the cells through the pores of the membrane by convection. The porous membrane onto which the compounds are printed is optically clear and preferably composed as stated in Appendix I. In another embodiment of this aspect of the present invention the detector layer of cells is a continuous or discontinuous sheet of cells, without any separation into test units or wells, growing on a porous and optically clear

membrane preferably of the types mentioned above. The porous membrane may allow the cells to send cellular processes through the pores of the membrane. The compounds are printed onto an optically clear substratum such as glass, plastic or quartz as solutions in solvent and allowed to dry. At the time of the experiment the cell sheet on the membrane, surrounded by a thin film of fluid, is layered ontop of the printed compound pattern. The compounds then dissolve and contact the cells via diffusion and convection. The compounds may be made using combinatorial chemistry techniques, and may be peptides. The compounds may be covalently attached to the optically clear substratum or porous membrane. The compounds may also be proteins, polypeptides or peptides secreted by or attached to the plasma membrane of genetically modified cells growing as a continuous or discontinuous sheet on a flat optically clear surface or an optically clear porous membrane.

The recording of the variation or result with respect to light emitted from the luminophore is performed by recording the spatially distributed light as one or more digital images, and the processing of the recorded variation to reduce it to one or more numbers representative of the degree of redistribution comprises a digital image processing procedure or combination of digital image processing procedures. The quantitative information which is indicative of the degree of the cellular response to the influence or the result of the influence on the intracellular pathway is extracted from the recording or recordings according to a predetermined calibration based on responses or results, recorded in the same manner, to known degrees of a relevant specific influence. This calibration procedure is developed according to principles described below (Developing an Image-based Assay Technique). Specific descriptions of the procedures for particular assays are given in the examples.

While the stepwise procedure necessary to reduce the image or images to the value representative of the response caused by the influence is particular to each assay, the individual steps are generally well-known methods of image processing. Some examples of the individual steps are point operations such as subtraction, ratioing, and thresholding, digital filtering methods such as smoothing, sharpening, and edge detection, spatial frequency methods such as Fourier filtering, image cross-correlation and image autocorrelation, object finding and classification (blob analysis), and colour space manipulations for visualisation. In addition to the algorithmic procedures, heuristic

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methods such as neural networks may also be used. In a preferred embodiment of the invention, a dose-response relationship is established based on quantification of the responses caused by a particular influence, representative of the underlying intracellular signalling process, using the methods described above and in examples 1-22 and 25. The dose-response relationship for the particular influence is then compared to the dose-response relationship obtained by performing the same assay in an instrument which allows parallel monitoring of all wells in a microtiter plate such as a FLIPR<sup>TM</sup> or an ordinary fluorescence plate reader for microtiter plates. If a good correlation between the dose-response relationships obtained from the two different measurement systems is obtained, it can be said that the parallel measurement mode has been validated (see examples 23 and 24). This implies that it can be used as the primary basis for a screening assay with the potential benefit of screening a significantly higher number of substances per unit of time for their influence on the response.

Imaging plate readers integrate the signal from each well into a single value per time point. Thus the data resulting from a single "run" of the instrument is a set of time series of single values, one for each well, with the injection of the test compound taking place at a known point in the time series. The primary advantage of this type of instrumentation is that it greatly increases the number of samples that can be processed in a given amount of time (the throughput). This is of great advantage when using the assay in a screening program for new pharmaceutical lead compounds.

The first step in the data analysis is to normalise the results from each well so that they can be compared with each other or with previously analysed known compounds. This always begins with correcting the signal by subtracting the instrument bias from all data points on a well-by-well basis. From this point, either of two techniques can be followed depending on the design of the assay:

Procedure 1: The average of the signal prior to the addition of the test compound is subtracted from all data points on a well-by-well basis.

Procedure 2: The data are corrected for any known background by subtracting the background value from all data points on a well-by-well basis. The resulting background-corrected data are normalised by dividing each data set by the average of the data values

prior to the injection of the test compound on a well-by-well basis.

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The corrected or normalised time series data sets are then further reduced by a technique that converts the time series to a single value. There are at least three such approaches:

- 1. For transient responses, the maximum deviation from the baseline is determined. This is also known as the "peak height" technique.
- 2. Alternatively, the signal is integrated over time between pre-defined limits. If the data were treated according to Procedure 2 above, then the offset is subtracted such that the integral of a non-response is zero within the limit of measurement error. This is also known as the "peak area" technique.
- 3. If the response is a cumulative one, e.g., an exponential change to a new level, the result is taken as the either the difference or the ratio between the signal after a predetermined time and the signal prior to the addition of the test compound.

All of the above procedures reduce the data for a given well to one or more single values. For screening purposes, these values will be searched for those that are greater than a certain statistically determined cut-off value. For characterisation, the values represent a quantitative response, and are further treated in sets by techniques such as dose-response curve fitting.

In another embodiment of the invention, the measurement of redistribution is accomplished indirectly by taking advantage of the fact that in order for redistribution to occur, the probe will experience some change in its freedom, or restriction, of movement within the intracellular milieu. The degree of translocation will correlate with the amount of freely mobile luminophore in the cytoplasm. At a point in time after the test compound has begun to have any influence it may have, the amount or fraction of restricted luminophore can be measured by disrupting or permeabilising the plasma membrane of the cells and allowing the freely mobile luminophore to diffuse away. If the detection volume of the detector is limited to the region immediately surrounding the cells, and the overall volume into which the freely mobile luminophore can diffuse is much larger, then the freely mobile luminophore essentially disappears from the detector's view and its signal is

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not recorded.

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In one aspect of the invention, the above mentioned measurement of redistribution is made on cells with permanently permeabilised plasma membranes immersed in a solution mimicking the cytoplasmic environment. In this way the influence of compounds that can normally not enter the cytoplasm of cells can be tested.

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The nucleic acid constructs used in the present invention encode in their nucleic acid sequences fusion polypeptides comprising a biologically active polypeptide that is a component of an intracellular signalling pathway, or a part thereof, and a GFP, preferably an F64L mutant of GFP, N- or C-terminally fused, optionally via a peptide linker, to the biologically active polypeptide or part thereof.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a protein kinase or a phosphatase.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a transcription factor or a part thereof which changes cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a protein, or a part thereof, which is associated with the cytoskeletal network and which changes cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a protein kinase or a part thereof which changes cellular localisation upon 20 activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a serine/threonine protein kinase or a part thereof capable of changing intracellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid 25 construct is a tyrosine protein kinase or a part thereof capable of changing intracellular localisation upon activation.

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In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a phospholipid-dependent serine/threonine protein kinase or a part thereof capable of changing intracellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a cAMP-dependent protein kinase or a part thereof capable of changing cellular localisation upon activation. In a preferred embodiment the biologically active polypeptide encoded by the nucleic acid construct is a PKAc-F64L-S65T-GFP fusion.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a cGMP-dependent protein kinase or a part thereof capable of changing cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a calmodulin-dependent serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a mitogen-activated serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation. In preferred embodiments the biologically active polypeptide encoded by the nucleic acid constructs are an ERK1-F64L-S65T-GFP fusion or an EGFP-ERK1 fusion.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a cyclin-dependent serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.

In one embodiment the biologically active polypeptide encoded by the nucleic acid construct is a protein phosphatase or a part thereof capable of changing cellular localisation upon activation.

In one preferred embodiment of the invention the nucleic acid constructs may be DNA constructs.

In one embodiment the biologically active polypeptide encoded by the nucleic acid

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construct. In one embodiment the gene encoding GFP in the nucleic acid construct is derived from Aequorea victoria. In a preferred embodiment the gene encoding GFP in the nucleic acid construct is EGFP or a GFP variant selected from F64L-GFP, F64L-Y66H-GFP and F64L-S65T-GFP.

In preferred embodiments of the invention the DNA constructs which can be identified by 5 any of the DNA sequences shown in SEQ ID NO: 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 78, 108, 110, 112, 114, 116, 118, 120, 122, 124, 126, 128, 130, 132, 134, 136, 138, 140, 142, 144, 146, 148, 150, and 152 or are variants of these sequences capable of encoding the same fusion polypeptide or a fusion polypeptide which is biologically equivalent thereto, e.g. an isoform, or a splice variant or a homologue from 10 another species.

The present invention describes a method that may be used to establish a screening program for the identification of biologically active substances that directly or indirectly affects intracellular signalling pathways and because of this property are potentially useful as medicaments. Based on measurements in living cells of the redistribution of spatially resolved luminescence from luminophores which undergo a change in distribution upon activation or deactivation of an intracellular signalling pathway the result of the individual measurement of each substance being screened indicates its potential biological activity.

In one embodiment of the invention the screening program is used for the identification of a biologically toxic substance as defined herein that exerts its toxic effect by interfering with an intracellular signalling pathway. Based on measurements in living cells of the redistribution of spatially resolved luminescence from luminophores which undergo a change in distribution upon activation or deactivation of an intracellular signalling pathway the result of the individual measurement of each substance being screened indicates its potential biologically toxic activity. In one embodiment of a screening program a compound that modulates a component of an intracellular pathway as defined herein, can be found and the therapeutic amount of the compound estimated by a method according to the method of the invention. In a preferred embodiment the present invention leads to the discovery of a new way of treating a condition or disease related to the intracellular function of a biologically active polypeptide comprising administration to a

patient suffering from said condition or disease of an effective amount of a compound which has been discovered by any method according to the invention. In another preferred embodiment of the invention a method is established for identification of a new drug target or several new drug targets among the group of biologically active polypeptides which are components of intracellular signalling pathways.

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In another embodiment of the invention an individual treatment regimen is established for the selective treatment of a selected patient suffering from an ailment where the available medicaments used for treatment of the ailment are tested on a relevant primary cell or cells obtained from said patient from one or several tissues, using a method comprising transfecting the cell or cells with at least one DNA sequence encoding a fluorescent probe according to the invention, transferring the transfected cell or cells back the said patient, or culturing the cell or cells under conditions permitting the expression of said probes and exposing it to an array of the available medicaments, then comparing changes in fluorescence patterns or redistribution patterns of the fluorescent probes in the intact living cells to detect the cellular response to the specific medicaments (obtaining a cellular action profile), then selecting one or more medicament or medicaments based on the desired activity and acceptable level of side effects and administering an effective amount of these medicaments to the selected patient.

The present invention describes a method that may be used to establish a screening program for back-tracking signal transduction pathways as defined herein. In one embodiment the screening program is used to establish more precisely at which level one or several compounds affect a specific signal transduction pathway by successively or in parallel testing the influence of the compound or compounds on the redistribution of spatially resolved luminescence from several of the luminophores which undergo a change in distribution upon activation or deactivation of the intracellular signalling pathway under study.

In general, a probe, i.e. a "GeneX"-GFP fusion or a GFP-"GeneX" fusion, is constructed using PCR with "GeneX"-specific primers followed by a cloning step to fuse "GeneX" in frame with GFP. The fusion may contain a short vector derived sequence between "GeneX" and GFP (e.g. part of a multiple cloning site region in the plasmid) resulting in a

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peptide linker between "GeneX" and GFP in the resulting fusion protein.

Some of the steps involved in the development of a probe include the following:

- Identify the sequence of the gene. This is most readily done by searching a depository of genetic information, e.g. the GenBank Sequence Database, which is widely available and routinely used by molecular biologists. In the specific examples below the GenBank Accession number of the gene in question is provided.
- Design the gene-specific primers. Inspection of the sequence of the gene allows design of gene-specific primers to be used in a PCR reaction. Typically, the top-strand primer encompasses the ATG start codon of the gene and the following ca. 20 nucleotides, while the bottom-strand primer encompasses the stop codon and the ca. 20 preceding nucleotides, if the gene is to be fused behind GFP, i.e. a GFP-"GeneX" fusion. If the gene is to be fused in front of GFP, i.e. a "GeneX"-GFP fusion, a stop codon must be avoided. Optionally, the full-length sequence of GeneX may not be used in the fusion, but merely the part that localizes and redistributes like GeneX in response to a signal. In addition to gene-specific sequences, the primers contain at least one recognition sequence for a restriction enzyme, to allow subsequent cloning of the PCR product. The sites are chosen so that they are unique in the PCR product and compatible with sites in the cloning vector. Furthermore, it may be necessary to include an exact number of nucleotides between the restriction enzyme site and the gene-specific sequence in order to establish the correct reading frame of the fusion gene and/or a translation initiation consensus sequence. Lastly, the primers always contain a few nucleotides in front of the restriction enzyme site to allow efficient digestion with the enzyme.
- Identify a source of the gene to be amplified. In order for a PCR reaction to produce a product with gene-specific primers, the gene-sequence must initially be present in the reaction, e.g. in the form of cDNA. Information in GenBank or the scientific literature will usually indicate in which tissue(s) the gene is expressed, and cDNA libraries from a great variety of tissues or cell types from various species are commercially available, e.g. from Clontech (Palo Alto), Stratagene (La Jolla) and Invitrogen (San Diego).

  Many genes are also available in cloned form from The American Type Tissue

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Collection (Virginia).

- Optimise the PCR reaction. Several factors are known to influence the efficiency and specificity of a PCR reaction, including the annealing temperature of the primers, the concentration of ions, notably Mg<sup>2+</sup> and K<sup>+</sup>, present in the reaction, as well as pH of the reaction. If the result of a PCR reaction is deemed unsatisfactory, it might be because the parameters mentioned above are not optimal. Various annealing temperatures should be tested, e.g. in a PCR machine with a built-in temperature gradient, available from e.g. Stratagene (La Jolla), and/or various buffer compositions should be tried, e.g. the OptiPrime buffer system from Stratagene (La Jolla).
- Clone the PCR product. The vector into which the amplified gene product will be cloned and fused with GFP will already have been taken into consideration when the primers were designed. When choosing a vector, one should at least consider in which cell types the probe subsequently will be expressed, so that the promoter controlling expression of the probe is compatible with the cells. Most expression vectors also contain one or more selective markers, e.g. conferring resistance to a drug, which is a useful feature when one wants to make stable transfectants. The selective marker should also be compatible with the cells to be used.

The actual cloning of the PCR product should present no difficulty as it typically will be a one-step cloning of a fragment digested with two different restriction enzymes into a vector digested with the same two enzymes. If the cloning proves to be problematic, it may be because the restriction enzymes did not work well with the PCR fragment. In this case one could add longer extensions to the end of the primers to overcome a possible difficulty of digestion close to a fragment end, or one could introduce an intermediate cloning step not based on restriction enzyme digestion. Several companies offer systems for this approach, e.g. Invitrogen (San Diego) and Clontech (Palo Alto).

Once the gene has been cloned and, in the process, fused with the GFP gene, the resulting product, usually a plasmid, should be carefully checked to make sure it is as expected. The most exact test would be to obtain the nucleotide sequence of the fusion-gene.

Once a DNA construct for a probe has been generated, its functionality and usefulness may

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be evaluated by transfecting it into cells capable of expressing the probe. The fluorescence of the cell is inspected soon after, typically the next day. At this point, two features of cellular fluorescence are noted: the intensity and the sub-cellular localisation.

The intensity should usually be at least as strong as that of unfused GFP in the cells. If it is not, the sequence or quality of the probe-DNA might be faulty, and should be carefully checked.

The sub-cellular localisation is an indication of whether the probe is likely to perform well. If it localises as expected for the gene in question, e.g. is excluded from the nucleus, it can immediately go on to a functional test. If the probe is not localised soon after the transfection procedure, it may be because of overexpression at this point in time, as the cell typically will have taken up very many copies of the plasmid, and localisation will occur in time, e.g. within a few weeks, as plasmid copy number and expression level decreases. If localisation does not occur after prolonged time, it may be because the fusion to GFP has destroyed a localisation function, e.g. masked a protein sequence essential for interaction with its normal cellular anchor-protein. In this case the opposite fusion might work, e.g. if GeneX-GFP does not work, GFP-GeneX might, as two different parts of GeneX will be affected by the proximity to GFP. If this does not work, the proximity of GFP at either end might be a problem, and it could be attempted to increase the distance by incorporating a longer linker between GeneX and GFP in the DNA construct.

If there is no prior knowledge of localisation, and no localisation is observed, it may be because the probe should not be localised at this point, because such is the nature of the protein fused to GFP. It should then be subjected to a functional test.

In a functional test, the cells expressing the probe are treated with at least one compound known to perturb, usually by activating, the signalling pathway on which the probe is expected to report by redistributing itself within the cell. If the redistribution is as expected, e.g. if prior knowledge tell that it should translocate from location X to location Y, it has passed the first critical test. In this case it can go on to further characterisation and quantification of the response.

If it does not perform as expected, it may be because the cell lacks at least one component

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of the signalling pathway, e.g. a cell surface receptor, or there is species incompatibility, e.g. if the probe is modelled on sequence information of a human gene product, and the cell is of hamster origin. In both instances one should identify other cell types for the testing process where these potential problems would not apply.

If there is no prior knowledge about the pattern of redistribution, the analysis of the redistribution will have to be done in greater depth to identify what the essential and indicative features are, and when this is clear, it can go on to further characterisation and quantification of the response. If no feature of redistribution can be identified, the problem might be as mentioned above, and the probe should be retested under more optimal cellular conditions.

If the probe does not perform under optimal cellular conditions, then it's back to the drawing board.

The process of developing an image-based redistribution assay begins with either the unplanned experimental observation that a redistribution phenomenon can be visualised, or the design of a probe specifically to follow a redistribution phenomenon already known to occur. In either event, the first and best exploratory technique is for a trained scientist or technician to observe the phenomenon. Even with the rapid advances in computing technology, the human eye-brain combination is still the most powerful pattern recognition system known, and requires no advance knowledge of the system in order to detect potentially interesting and useful patterns in raw data. This is especially if those data are presented in the form of images, which are the natural "data type" for human visual processing. Because human visual processing operates most effectively in a relatively narrow frequency range, i.e., we cannot see either very fast or very slow changes in our visual field, it may be necessary to record the data and play it back with either time dilation or time compression.

Some luminescence phenomena cannot be seen directly by the human eye. Examples include polarisation and fluorescence lifetime. However, with suitable filters or detectors, these signals can be recorded as images or sequences of images and displayed to the human in the fashion just described. In this way, patterns can be detected and the same methods can be applied.

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Once the redistribution has been determined to be a reproducible phenomenon, one or more data sets are generated for the purpose of developing a procedure for extracting the quantitative information from the data. In parallel, the biological and optical conditions are determined which will give the best quality raw data for the assay. This can become an iterative process; it may be necessary to develop a quantitative procedure in order to assess the effect on the assay of manipulating the assay conditions.

The data sets are examined by a person or persons with knowledge of the biological phenomenon and skill in the application of image processing techniques. The goal of this exercise is to determine or at least propose a method that will reduce the image or sequence of images constituting the record of a "response" to a value corresponding to the degree of the response. Using either interactive image processing software or an image processing toolbox and a programming language, the method is encoded as a procedure or algorithm that takes the image or images as input and generates the degree of response (in any units) as its output. Some of the criteria for evaluating the validity of a particular procedure are:

- Does the degree of the response vary in a biologically significant fashion, i.e., does
  it show the known or putative dependence on the concentration of the stimulating
  agent or condition?
- Is the degree of response reproducible, i.e., does the same concentration or level of stimulating agent or condition give the same response with an acceptable variance?
- Is the dynamic range of the response sufficient for the purpose of the assay? If not, can a change in the procedure or one of its parameters improve the dynamic range?
- Does the procedure exhibit any clear "pathologies", i.e., does it give ridiculous values for the response if there are commonly occurring imperfections in the imaging process? Can these pathologies be eliminated, controlled, or accounted for?
- Can the procedure deal with the normal variation in the number and/or size of cells in an image?

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In some cases the method may be obvious; in others, a number of possible procedures may suggest themselves. Even if one method appears clearly superior to others, optimisation of parameters may be required. The various procedures are applied to the data set and the criteria suggested above are determined, or the single procedure is applied repeatedly with adjustment of the parameter or parameters until the most satisfactory combination of signal, noise, range, etc. are arrived at. This is equivalent to the calibration of any type of single-channel sensor.

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The number of ways of extracting a single value from an image are extremely large, and thus an intelligent approach must be taken to the initial step of reducing this number to a small, finite number of possible procedures. This is not to say that the procedure arrived at is necessarily the best procedure - but a global search for the best procedure is simply out of the question due to the sheer number of possibilities involved.

Image-based assays are no different than other assay techniques in that their usefulness is characterised by parameters such as the specificity for the desired component of the sample, the dynamic range, the variance, the sensitivity, the concentration range over which the assay will work, and other such parameters. While it is not necessary to characterise each and every one of these before using the assay, they represent the only way to compare one assay with another.

The final step is then to see whether there exists a possibility to increase the throughput of the assay to improve its utility as the basis of a screening program. In order to do this, a dose-response relationship is established based on quantification of the responses caused by a particular influence, representative of the underlying intracellular signalling process, using the methods described above and in examples 1-22 and 25. The dose-response relationship for the particular influence is then compared to the dose-response relationship obtained by performing the same assay in an instrument which allows parallel monitoring of all wells in a microtiter plate such as a FLIPR<sup>TM</sup> or an ordinary imaging or fluorescence plate reader for microtiter plates. If a good correlation between the dose-response relationships obtained from the two different measurement systems is obtained, it can be said that the parallel measurement mode has been validated (see examples 23 and 24). This implies that it can be used as the primary basis for a screening program with the potential

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benefit of screening a significantly higher number of substances for their influence on the response per unit of time.

The process of developing an image-based assay is best illustrated by example. The development of such an assay for GLUT4 translocation is hereby described. GLUT4 is a member of the class of glucose transporter molecules that are important in cellular glucose uptake. It is known to translocate to the plasma membrane under some conditions of stimulation of glucose uptake. The ability to visualise the glucose uptake response non-invasively, without actually measuring glucose uptake, would be a very useful assay for anyone looking for, for example, treatments for type II diabetes.

A CHO cell line which stably expressed the human insulin receptor was used as the basis for a new cell line which stably expressed a fusion between GLUT4 and GFP. This cell line was expected to show translocation of GLUT4 to the plasma membrane as visualised by the movement of the GFP. The translocation could definitely be seen in the form of the appearance of local increases in the fluorescence in regions of the plasma membrane which had a characteristic shape or pattern. This is shown in Figure 12.

These objects became known as "snircles", and the phenomenon of their appearance as "snircling". In order to quantify their appearance, a method had to be found to isolate them as objects in the image field, and then enumerate them, measure their area, or determine some parameter about them which correlated in a dose-dependent fashion with the concentration of insulin to which the cells had been exposed. In order to separate the snircles, a binarization procedure was applied in which one copy of the image smoothed with a relatively severe gaussian kernel (sigma = 2.5) was subtracted from another copy to which only a relatively light gaussian smooth had been applied (sigma=0.5). The resultant image was rescaled to its min/max range, and an automatic threshold was applied to divide the image into two levels. The thresholded image contains a background of one value all found object with another value. The found objects were first filtered through a filter to remove objects far too large and far too small to be snircles. The remaining objects, which represent snircles and other artifacts from the image with approximately the same size and intensity characteristics as snircles, are passed into a classification procedure which has been previously trained with many images of snircles to recognize snircles and exclude the

other artifacts. The result of this procedure is a binary image that shows only the found snircles to the degree to which the classification procedure can accurately identify them. The total area of the snircles is then summed and this value is the quantitative measure of the degree of snircling for that image.

Another approach to the problem of quantifying GLUT 4 translocation has been performed 5 and validated using the same type of experimental protocol but a different image processing approach. In this case the objects of interest in the cells are not the appearance of snircles at the plasma membrane but the disappearance of GLUT4-GFP fluorescence from its intracellular site. With this method the bright area, consisting of GLUT4-GFP, centrally located in each cell is identified by a thresholding procedure. This demarcates a 10 certain area for the centrally located GLUT4-GFP. In the next step the total fluorescence intensity in this area is quantified on each image in the image series, i.e. over time. The response for each cell is defined as the difference in fluorescence intensity in the centrally located GLUT4-GFP area before and a fixed point in time after application of the influence. The dose-response relationship for insulin using the above described 15 quantitation procedure is shown in Figure 13. It can be seen that the ED50 value for insulin to reduce central GLUT4-GFP fluorescence is 0.3 nM.

In the present specification and claims, the term "an influence" covers any influence to which the cellular response comprises a redistribution. Thus, e.g., heating, cooling, high pressure, low pressure, humidifying, or drying are influences on the cellular response on which the resulting redistribution can be quantified, but as mentioned above, perhaps the most important influences are the influences of contacting or incubating the cells with substances which are known or suspected to exert an influence on the cellular response involving a redistribution contribution. In another embodiment of the invention the influence could be substances from a compound drug library.

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In the present context, the term "green fluorescent protein" is intended to indicate a protein which, when expressed by a cell, emits fluorescence upon exposure to light of the correct excitation wavelength (cf. [(Chalfie, M. et al. (1994) Science 263, 802-805)]). In the following, GFP in which one or more amino acids have been substituted, inserted or deleted is most often termed "modified GFP". "GFP" as used herein includes wild-type

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GFP derived from the jelly fish Aequorea victoria and modifications of GFP, such as the blue fluorescent variant of GFP disclosed by Heim et al. (1994). Proc.Natl.Acad.Sci. 91:26, pp 12501-12504, and other modifications that change the spectral properties of the GFP fluorescence, or modifications that exhibit increased fluorescence when expressed in cells at a temperature above about 30°C described in PCT/DK96/00051, published as WO 97/11094 on 27 March 1997 and hereby incorporated by reference, and which comprises a fluorescent protein derived from Aequorea Green Fluorescent Protein (GFP) or any functional analogue thereof, wherein the amino acid in position 1 upstream from the chromophore has been mutated to provide an increase of fluorescence intensity when the fluorescent protein of the invention is expressed in cells. Preferred GFP variants are F64L-GFP, F64L-Y66H-GFP and F64L-S65T-GFP. An especially preferred variant of GFP for use in all the aspects of this invention is EGFP (DNA encoding EGFP which is a F64L-S65T variant with codons optimized for expression in mammalian cells is available from Clontech, Palo Alto, plasmids containing the EGFP DNA sequence, cf. GenBank Acc. Nos. U55762, U55763).

The term "intracellular signalling pathway" and "signal transduction pathway" are 15 intended to indicate the co-ordinated intracellular processes whereby a living cell transduce an external or internal signal into cellular responses. Said signal transduction will involve an enzymatic reaction said enzymes include but are not limited to protein kinases, GTPases, ATPases, protein phosphatases, phospholipases and cyclic nucleotide phosphodiesterases. The cellular responses include but are not limited to gene 20 transcription, secretion, proliferation, mechanical activity, metabolic activity, cell death.

The term "second messenger" is used to indicate a low molecular weight component involved in the early events of intracellular signal transduction pathways.

The term "luminophore" is used to indicate a chemical substance that has the property of emitting light either inherently or upon stimulation with chemical or physical means. This includes but is not limited to fluorescence, bioluminescence, phosphorescence, and chemiluminescence.

The term "mechanically intact living cell" is used to indicate a cell which is considered living according to standard criteria for that particular type of cell such as maintenance of normal membrane potential, energy metabolism, proliferative capability, and has not

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experienced any physically invasive treatment designed to introduce external substances into the cell such as microinjection.

In the present context, the term "permeabilised living cell" is used to indicate cells where a pore forming agent such as Streptolysin O or Staphylococcus Aureus α-toxin has been applied and thereby incorporated into the plasma membrane in the cells. This creates proteinaceous pores with a defined pore size in the plasma membranes of the exposed cells. Pores could also be made by electroporation, i.e. exposing the cells to high voltage discharges, a procedure that creates small holes in the plasma membrane by coagulating integral membrane proteins. Treatment with a mild detergent such as saponin may accomplish the same thing. Common to all these treatments are that pores are formed only in the plasma membrane without affecting the integrity of cytoplasmic structural elements and organelles. The term living in this context means that the permeabilised cells bathed in a solution mimicking the intracellular milieu still have functional organelles, such as actively respiring mitochondria and endoplasmic reticulum that can take up and release calcium ions, and functional structural elements. The benefit of this method is that substances that normally can not traverse the plasma membrane, but most likely exert their influence intracellularly, can be introduced and their influence studied without cumbersome microinjection of the substances into single cells. Using this method the response to an influence can be recorded from many cells simultaneously.

In the present context, the term "permeabilisation" is intended to indicate the selective disruption of the plasma membrane barrier so that soluble substances freely mobile in the cytosol are lost from the cells. The permeabilisation can be achieved as described above under "permeabilised living cells" or by using other chemical detergents such as Triton X-100 or digitonin in carefully titrated amounts.

The term "physiologically relevant", when applied to an experimentally determined redistribution of an intracellular component, as measured by a change in the luminescence properties or distribution, is used to indicate that said redistribution can be explained in terms of the underlying biological phenomenon which gives rise to the redistribution.

The terms "image processing" and "image analysis" are used to describe a large family of digital data analysis techniques or combination of such techniques which reduce ordered

arrays of numbers (images) to quantitative information describing those ordered arrays of numbers. When said ordered arrays of numbers represent measured values from a physical process, the quantitative information derived is therefore a measure of the physical process.

The term "fluorescent probe" is used to indicate a fluorescent fusion polypeptide comprising a GFP or any functional part thereof which is N- or C-terminally fused to a biologically active polypeptide as defined herein, optionally via a peptide linker consisting of one or more amino acid residues, where the size of the linker peptide in itself is not critical as long as the desired functionality of the fluorescent probe is maintained. A fluorescent probe according to the invention is expressed in a cell and basically mimics the physiological behaviour of the biologically active polypeptide moiety of the fusion polypeptide.

The term "mammalian cell" is intended to indicate any living cell of mammalian origin. The cell may be an established cell line, many of which are available from The American Type Culture Collection (ATCC, Virginia, USA) or a primary cell with a limited life span 15 derived from a mammalian tissue, including tissues derived from a transgenic animal, or a newly established immortal cell line derived from a mammalian tissue including transgenic tissues, or a hybrid cell or cell line derived by fusing different cell types of mammalian origin e.g. hybridoma cell lines. The cells may optionally express one or more non-native gene products, e.g. receptors, enzymes, enzyme substrates, prior to or in 20 addition to the fluorescent probe. Preferred cell lines include but are not limited to those of fibroblast origin, e.g. BHK, CHO, BALB, or of endothelial origin, e.g. HUVEC, BAE (bovine artery endothelial), CPAE (cow pulmonary artery endothelial), HLMVEC (human lung microvascular endothelial cells) or of pancreatic origin, e.g. RIN, INS-1, MIN6, bTC3, aTC6, bTC6, HIT, or of hematopoietic origin, e.g.primary isolated human 25 monocytes, macrophages, neutrophils, basophils, eosinophils and lyphocyte populations, AML-193, HL-60, RBL-1, adipocyte origin, e.g. 3T3-L1, neuronal/neuroendocrine origin, e.g. AtT20, PC12, GH3, muscle origin, e.g. SKMC, A10, C2C12, renal origin, e.g. HEK 293, LLC-PK1.

30 The term "hybrid polypeptide" is intended to indicate a polypeptide which is a fusion of at

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least a portion of each of two proteins, in this case at least a portion of the green fluorescent protein, and at least a portion of a catalytic and/or regulatory domain of a protein kinase. Furthermore a hybrid polypeptide is intended to indicate a fusion polypeptide comprising a GFP or at least a portion of the green fluorescent protein that contains a functional fluorophore, and at least a portion of a biologically active polypeptide as defined herein provided that said fusion is not the PKCα-GFP, PKCγ-GFP, and PKCε-GFP disclosed by Schmidt *et al.* and Sakai *et al.*, respectively. Thus, GFP may be N- or C-terminally tagged to a biologically active polypeptide, optionally via a linker portion or linker peptide consisting of a sequence of one or more amino acids. The hybrid polypeptide or fusion polypeptide may act as a fluorescent probe in intact living cells carrying a DNA sequence encoding the hybrid polypeptide under conditions permitting expression of said hybrid polypeptide.

The term "kinase" is intended to indicate an enzyme that is capable of phosphorylating a cellular component.

The term "protein kinase" is intended to indicate an enzyme that is capable of phosphorylating serine and/or threonine and/or tyrosine in peptides and/or proteins.

The term "phosphatase" is intended to indicate an enzyme that is capable of dephosphorylating phosphoserine and/or phosphothreonine and/or phosphotyrosine in peptides and/or proteins.

The term "cyclic nucleotide phosphodiesterase" is intended to indicate an enzyme that is capable of inactivating the second messengers cAMP and cGMP by hydrolysis of their 3'-ester bond.

In the present context, the term "biologically active polypeptide" is intended to indicate a polypeptide affecting intracellular processes upon activation, such as an enzyme which is active in intracellular processes or a portion thereof comprising a desired amino acid sequence which has a biological function or exerts a biological effect in a cellular system. In the polypeptide one or several amino acids may have been deleted, inserted or replaced to alter its biological function, e.g. by rendering a catalytic site inactive. Preferably, the biologically active polypeptide is selected from the group consisting of proteins taking part

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in an intracellular signalling pathway, such as enzymes involved in the intracellular phosphorylation and dephosphorylation processes including kinases, protein kinases and phosphorylases as defined herein, but also proteins making up the cytoskeleton play important roles in intracellular signal transduction and are therefore included in the meaning of "biologically active polypeptide" herein. More preferably, the biologically active polypeptide is a protein which according to its state as activated or non-activated changes localisation within the cell, preferably as an intermediary component in a signal transduction pathway. Included in this preferred group of biologically active polypeptides are cAMP dependent protein kinase A.

The term "a substance having biological activity" is intended to indicate any sample that has a biological function or exerts a biological effect in a cellular system. The sample may be a sample of a biological material such as a sample of a body fluid including blood, plasma, saliva, milk, urine, or a microbial or plant extract, an environmental sample containing pollutants including heavy metals or toxins, or it may be a sample containing a compound or mixture of compounds prepared by organic synthesis or genetic techniques.

The phrase "any change in fluorescence" means any change in absorption properties, such as wavelength and intensity, or any change in spectral properties of the emitted light, such as a change of wavelength, fluorescence lifetime, intensity or polarisation, or any change in the intracellular localisation of the fluorophore. It may thus be localised to a specific cellular component (e.g. organelle, membrane, cytoskeleton, molecular structure) or it may be evenly distributed throughout the cell or parts of the cell.

The term "organism" as used herein indicates any unicellular or multicellular organism preferably originating from the animal kingdom including protozoans, but also organisms that are members of the plant kingdoms, such as algae, fungi, bryophytes, and vascular plants are included in this definition.

The term "nucleic acid" is intended to indicate any type of poly- or oligonucleic acid sequence, such as a DNA sequence, a cDNA sequence, or an RNA sequence.

The term "biologically equivalent" as it relates to protein is intended to mean that a first protein is equivalent to a second protein if the cellular functions of the two proteins may

substitute for each other, e.g. if the two proteins are closely related isoforms encoded by different genes, if they are splicing variants, or allelic variants derived from the same gene, if they perform identical cellular functions in different cell types, or in different species. The term "biologically equivalent" as it relates to DNA is intended to mean that a first DNA sequence encoding a polypeptide is equivalent to a second DNA sequence encoding a polypeptide if the functional proteins encoded by the two genes are biologically equivalent.

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The phrase "back-tracking of a signal transduction pathway" is intended to indicate a process for defining more precisely at what level a signal transduction pathway is affected, either by the influence of chemical compounds or a disease state in an organism. Consider a specific signal transduction pathway represented by the bioactive polypeptides A - B - C - D, with signal transduction from A towards D. When investigating all components of this signal transduction pathway compounds or disease states that influence the activity or redistribution of only D can be considered to act on C or downstream of C whereas compounds or disease states that influence the activity or redistribution of C and D, but not of A and B can be considered to act downstream of B.

The term "fixed cells" is used to mean cells treated with a cytological fixative such as glutaraldehyde or formaldehyde, treatments that serve to chemically cross-link and stabilise soluble and insoluble proteins within the structure of the cell. Once in this state, such proteins cannot be lost from the structure of the now-dead cell.

In the present context a "screening assay" is intended to mean any measurement protocol, including materials, cells, instruments, chemicals, reagents, detection units, calibration and quantification procedures used to measure a response from mechanically intact or permeabilised living cells relevant to influences on an intracellular pathway.

The term "dose-response relationship" and "screening programme" is in the present context intended to mean a clear correlation between the quantified response of cells in a screening assay to application of an influence, such as a compound, and the concentration of the applied influence. The response to the influence may be both an up-regulation and a down-regulation of the quantified parameter used in the screening assay.

In the present context, the term "physiology" is intended to mean the normal function of biological and biochemical processes inside cells, between cells and in the whole organism or animal.

## 5 BRIEF DESCRIPTION OF THE DRAWINGS

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Figure 1. CHO cells expressing the PKAc-F64L-S65T-GFP hybrid protein have been treated in HAM's F12 medium with 50 μM forskolin at 37°C. The images of the GFP fluorescence in these cells have been taken at different time intervals after treatment, which were: a) 40 seconds b) 60 seconds c) 70 seconds d) 80 seconds. The fluorescence changes from a punctate to a more even distribution within the (non-nuclear) cytoplasm.

Figure 2. Time-lapse analysis of forskolin induced PKAc-F64L-S65T-GFP redistribution. CHO cells, expressing the PKAc-F64L-S65T-GFP fusion protein were analysed by time-lapse fluorescence microscopy. Fluorescence micrographs were acquired at regular intervals from 2 min before to 8 min after the addition of agonist. The cells were challenged with 1  $\mu$ M forskolin immediately after the upper left image was acquired (t=0). Frames were collected at the following times: i) 0, ii) 1, iii) 2, iv) 3, v) 4 and vi) 5 minutes. Scale bar 10  $\mu$ m.

Figure 3. Time-lapse analyses of PKAc-F64L-S65T-GFP redistribution in response to various agonists. The effects of 1 μM forskolin (A), 50 μM forskolin (B), 1mM dbcAMP (C) and 100 μM IBMX (D) (additions indicated by open arrows) on the localisation of the PKAc-F64L-S65T-GFP fusion protein were analysed by time-lapse fluorescence microscopy of CHO/PKAc-F64L-S65T-GFP cells. The effect of addition of 10 μM forskolin (open arrow), followed shortly by repeated washing with buffer (solid arrow), on the localisation of the PKAc-F64L-S65T-GFP fusion protein was analysed in the same cells (E). In a parallel experiment, the effect of adding 10 μM forskolin and 100 μM IBMX (open arrow) followed by repeated washing with buffer containing 100 μM IBMX

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(solid arrow) was analysed (F). Removing forskolin caused PKAc-F64L-S65T-GFP fusion protein to return to the cytoplasmic aggregates while this is prevented by the continued presence of IBMX (F). The effect of 100 nM glucagon (Fig 3G, open arrow) on the localisation of the PKAc-F64L-S65T-GFP fusion protein is also shown for BHK/GR, PKAc-F64L-S65T-GFP cells. The effect of 10 μM norepinephrine (H), solid arrow, on the localisation of the PKAc-F64L-S65T-GFP fusion protein was analysed similarly, in transiently transfected CHO, PKAc-F64L-S65T-GFP cells, pretreated with 10 μM forskolin, open arrow, to increase [cAMP]. N.B. in Fig 3H the x-axis counts the image numbers, with 12 seconds between images. The raw data of each experiment consisted of 60 fluorescence micrographs acquired at regular intervals including several images acquired before the addition of buffer or agonist. The charts (A-G) each show a quantification of the response seen through all the 60 images, performed as described in analysis method 2. The change in total area of the highly fluorescent aggregates, relative to the initial area of fluorescent aggregates is plotted as the ordinate in all graphs in Figure 3, versus time for each experiment. Scale bar 10 μm.

Figure 4. Dose-response curve (two experiments) for forskolin-induced redistribution of the PKAc-F64L-S65T-GFP fusion.

Figure 5. Time from initiation of a response to half maximal (t<sub>1/2max</sub>) and maximal (t<sub>max</sub>) PKAc-F64L-S65T-GFP redistribution. The data was extracted from curves such as that shown in "Figure 2." All t<sub>1/2max</sub> and t<sub>max</sub> values are given as mean±SD and are based on a total of 26-30 cells from 2-3 independent experiments for each forskolin concentration. Since the observed redistribution is sustained over time, the t<sub>max</sub> values were taken as the earliest time point at which complete redistribution is reached. Note that the values do not relate to the degree of redistribution.

Figure 6. Parallel dose-response analyses of forskolin induced cAMP elevation and PKAc-

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F64L-S65T-GFP redistribution. The effects of buffer or 5 increasing concentrations of forskolin on the localisation of the PKAc-F64L-S65T-GFP fusion protein in CHO/PKAc-F64L-S65T-GFP cells, grown in a 96 well plate, were analysed as described above. Computing the ratio of the SD's of fluorescence micrographs taken of the same field of cells, prior to and 30 min after the addition of forskolin, gave a reproducible measure of PKAc-F64L-S65T-GFP redistribution. The graph shows the individual 48 measurements and a trace of their mean±s.e.m at each forskolin concentration. For comparison, the effects of buffer or 8 increasing concentrations of forskolin on [cAMP], was analysed by a scintillation proximity assay of cells grown under the same conditions. The graph shows a trace of the mean ± s.e.m of 4 experiments expressed in arbitrary units.

Figure 7. BHK cells stably transfected with the human muscarinic (hM1) receptor and the PKC $\alpha$ -F64L-S65T-GFP fusion. Carbachol (100  $\mu$ M added at 1.0 second) induced a transient redistribution of PKC $\alpha$ -F64L-S65T-GFP from the cytoplasm to the plasma membrane. Images were taken at the following times: a) 1 second before carbachol addition, b) 8.8 seconds after addition and c) 52.8 seconds after addition.

Figure 8. BHK cells stably transfected with the hM1 receptor and PKC $\alpha$ -F64L-S65T-GFP fusion were treated with carbachol (1  $\mu$ M, 10  $\mu$ M, 100  $\mu$ M). In single cells intracellular [Ca²+] was monitored simultaneously with the redistribution of PKC $\alpha$ -F64L-S65T-GFP. Dashed line indicates the addition times of carbachol. The top panel shows changes in the intracellular Ca²+ concentration of individual cells with time for each treatment. The middle panel shows changes in the average cytoplasmic GFP fluorescence for individual cells against time for each treatment. The bottom panel shows changes in the fluorescence of the periphery of single cells, within regions that specifically include the circumferential edge of a cell as seen in normal projection, the best regions for monitoring changes in the fluorescence intensity of the plasma membrane.

Figure 9.

- a) The hERK1-F64L-S65T-GFP fusion expressed in HEK293 cells treated with 100 μM of the MEK1 inhibitor PD98059 in HAM F-12 (without serum) for 30 minutes at 37 °C. The nuclei empty of fluorescence during this treatment.
- 5 b) The same cells as in (a) following treatment with 10 % foetal calf serum for 15 minutes at 37 °C.
  - c) Time profiles for the redistribution of GFP fluorescence in HEK293 cells following treatment with various concentrations of EGF in Hepes buffer (HAM F-12 replaced with Hepes buffer directly before the experiment). Redistribution of fluorescence is expressed as the change in the ratio value between areas in nucleus and cytoplasm of single cells. Each time profile is the mean for the changes seen in six single cells.
  - d) Bar chart for the end-point measurements, 600 seconds after start of EGF treatments, of fluorescence change (nucleus:cytoplasm) following various concentrations of EGF.

# 15 Figure 10.

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- a) The SMAD2-EGFP fusion expressed in HEK293 cells starved of serum overnight in HAM F-12. HAM F-12 was then replaced with Hepes buffer pH 7.2 immediately before the experiment. Scale bar is 10 μm.
- b) HEK 293 cells expressing the SMAD2-EGFP fusion were treated with various concentration of TGF-beta as indicated, and the redistribution of fluorescence monitored against time. The time profile plots represent increases in fluorescence within the nucleus, normalised to starting values in each cell measured. Each trace is the time profile for a single cell nucleus.
- c) A bar chart representing the end-point change in fluorescence within nuclei (after 850 seconds of treatment) for different concentrations of TGF-beta. Each bar is the value for a single nucleus in each treatment.

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Figure 11. The VASP-F64L-S65T-GFP fusion in CHO cells stably transfected with the human insulin receptor. The cells were starved for two hours in HAM F-12 without serum, then treated with 10% foetal calf serum. The image shows the resulting redistribution of fluorescence after 15 minutes of treatment. GFP fluorescence becomes localised in structures identified as focal adhesions along the length of actin stress fibres.

Figure 12. Time lapse recording GLUT4-GFP redistribution in CHO-HIR cells. Time indicates minutes after the addition of 100 nM insulin.

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- Figure 13. Dose-response relationships for the influence of insulin on the disappearance of total fluorescence from the centrally located area of GLUT4-GFP. Data points indicate mean±SE.
- Figure 14. Dose-response relationship for the translocation of PKCα-GFP in BHKhM1 cells stimulated with the muscarininc agonist carbamylcholine using a FLIPR™ to do the actual experiments.
- Figure 15. Dose-response relationship for the translocation of PKAc-GFP in CHO/PKAc-20 F64L-S65T-GFP cells stimulated with forskolin using a FLIPR™ to do the actual experiments.
  - Figure 16. Dose-response relationship for the disappearance of fluorescence from permeabilised CHO/PKAc-F64L-S65T-GFP when previously exposed to different doses of forskolin.

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### **EXAMPLES**

### EXAMPLE 1

Construction, testing and implementation of an assay for cAMP based on PKA activation in real time within living cells.

Useful for monitoring the activity of signalling pathways that lead to altered concentrations of cAMP, e.g. activation of G-protein coupled receptors which couple to G-proteins of the  $G_s$  or  $G_t$  class.

The catalytic subunit of the murine cAMP dependent protein kinase (PKAc) was fused C-terminally to a F64L-S65T derivative of GFP. The resulting fusion (PKAc-F64L-S65T-GFP) was used for monitoring *in vivo* the translocation and thereby the activation of PKA.

To construct the PKAc-F64L-S65T-GFP fusion, convenient restriction endonuclease sites were introduced into the cDNAs encoding murine PKAc (Gen Bank Accession number: M12303) and F64L-S65T-GFP (sequence disclosed in WO 97/11094) by polymerase chain reaction (PCR). The PCR reactions were performed according to standard protocols with the following primers:

## 5'PKAc:

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TTggACACAAgCTTTggACACCCTCAggATATgggCAACgCCgCCgCCGCCAAg (SEQ ID NO:3),

### 20 3'PKAc:

 $\label{eq:gtcatcttctc} g A g T C T T C A g g C g C C C A A A C T C A g T A A A C T C C T T g C C A C C (SEQ ID NO:4) \,,$ 

5'GFP: TTggACACAAgCTTTggACACggCgCCCATgAgTAAAggAgAAGAACTTTTC (SEQ ID NO:1),

25 3'GFP: gTCATCTTCTCgAgTCTTACTCCTgAggTTTgTATAgTTCATCCATgCCATgT (SEQ ID NO:2).

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The PKAc amplification product was then digested with HindIII+AscI and the F64L-S65T-GFP product with AscI+XhoI. The two digested PCR products were subsequently ligated with a HindIII+XhoI digested plasmid (pZeoSV® mammalian expression vector, Invitrogen, San Diego, CA, USA). The resulting fusion construct (SEQ ID NO:68 & 69) was under control of the SV40 promoter.

## Transfection and cell culture conditions:

Chinese hamster ovary cells (CHO), were transfected with the plasmid containing the PKAc-F64L-S65T-GFP fusion using the calcium phosphate precipitate method in HEPES-buffered saline (Sambrook *et al.*, 1989). Stable transfectants were selected using 1000 μg Zeocin/ml (Invitrogen) in the growth medium (DMEM with 1000 mg glucose/l, 10 % fetal bovine serum (FBS), 100 μg penicillin-streptomycin mixture ml<sup>-1</sup>, 2 mM L-glutamine purchased from Life Technologies Inc., Gaithersburg, MD, USA). Untransfected CHO cells were used as the control. To assess the effect of glucagon on fusion protein translocation, the PKAc-F64L-S65T-GFP fusion was stably expressed in baby hamster kidney cells overexpressing the human glucagon receptor (BHK/GR cells). Untransfected BHK/GR cells were used as the control. Expression of GR was maintained with 500 μg G418/ml (*Neo* marker) and PKAc-F64L-S65T-GFP was maintained with 500 μg Zeocin/ml (*Sh ble* marker). CHO cells were also simultaneously co-transfected with vectors containing the PKAc-F64L-S65T-GFP fusion and the human α2a adrenoceptor (hARa2a).

For fluorescence microscopy, cells were allowed to adhere to Lab-Tek chambered coverglasses (Nalge Nunc Int., Naperville, IL, USA) for at least 24 hours and cultured to about 80% confluence. Prior to experiments, the cells were cultured over night without selection pressure in HAM F-12 medium with glutamax (Life Technologies), 100 µg penicillin-streptomycin mixture ml<sup>-1</sup> and 0.3 % FBS. This medium has low autofluorescence enabling fluorescence microscopy of cells straight from the incubator.

Monitoring activity of PKA activity in real time:

Image aquisition of live cells were gathered using a Zeiss Axiovert 135M fluorescence microscope fitted with a Fluar 40X, NA: 1.3 oil immersion objective and coupled to a Photometrics CH250 charged coupled device (CCD) camera. The cells were illuminated with a 100 W HBO arc lamp. In the light path was a 470±20 nm excitation filter, a 510 nm dichroic mirror and a 515±15 nm emission filter for minimal image background. The cells were maintained at 37°C with a custom built stage heater.

Images were processed and analysed in the following manner:

# Method 1: Stepwise procedure for quantitation of translocation of PKA:

- 1. The image was corrected for dark current by performing a pixel-by-pixel subtraction of a dark image (an image taken under the same conditions as the actual image, except the camera shutter is not allowed to open).
  - 2. The image was corrected for non-uniformity of the illumination by performing a pixel-by-pixel ratio with a flat field correction image (an image taken under the same conditions as the actual image of a uniformly fluorescent specimen).
- 3. The image histogram, i.e., the frequency of occurrence of each intensity value in the image, was calculated.
  - 4. A smoothed, second derivative of the histogram was calculated and the second zero is determined. This zero corresponds to the inflection point of the histogram on the high side of the main peak representing the bulk of the image pixel values.
- 5. The value determined in step 4 was subtracted from the image. All negative values were discarded.
  - 6. The variance (square of the standard deviation) of the remaining pixel values was determined. This value represents the "response" for that image.
  - 7. Scintillation proximity assay (SPA) for independent quantitation of cAMP.

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- 1. The fluorescent aggregates are segmented from each image using an automatically found threshold based on the maximisation of the information measure between the object and background. The *a priori* entropy of the image histogram is used as the information measure.
- 5 2. The area of each image occupied by the aggregates is calculated by counting pixels in the segmented areas.
  - 3. The value obtained in step 2 for each image in a series, or treatment pair, is normalised to the value found for the first (unstimulated) image collected. A value of zero (0) indicates no redistribution of fluorescence from the starting condition. A value of one
- (1) by this method equals full redistribution.

Cells were cultured in HAM F-12 medium as described above, but in 96-well plates. The medium was exchanged with Ca<sup>2+</sup>-HEPES buffer including 100 µM IBMX and the cells were stimulated with different concentrations of forskolin for 10 min. Reactions were stopped with addition of NaOH to 0.14 M and the amount of cAMP produced was measured with the cAMP-SPA kit, RPA538 (Amersham) as described by the manufacturer.

Manipulating intracellular levels of cAMP to test the PKAc-F64L-S65T-GFP fusion.

The following compounds were used to vary cAMP levels: Forskolin, an activator of adenylate cyclase; dbcAMP, a membrane permeable cAMP analog which is not degraded by phosphodiesterase; IBMX, an inhibitor of phosphodiesterase.

CHO cells stably expressing the PKAc-F64L-S65T-GFP, showed a dramatic translocation of the fusion protein from a punctate distribution to an even distribution throughout the cytoplasm following stimulation with 1  $\mu$ M forskolin (n=3), 10  $\mu$ M forskolin (n=4) and 50  $\mu$ M forskolin (n=4) (Fig 1), or dbcAMP at 1mM (n=6).

Fig. 2 shows the progression of response in time following treatment with 1 µM forskolin.

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Fig. 3 gives a comparison of the average temporal profiles of fusion protein redistribution and a measure of the extent of each response to the three forskolin concentrations (Fig. 3A, E, B), and to 1 mM dbcAMP (fig 3C) which caused a similar but slower response, and to addition of 100  $\mu$ M IBMX (n=4, Fig. 3D) which also caused a slow response, even in the absence of adenylate cyclase stimulation. Addition of buffer (n=2) had no effect (data not shown).

As a control for the behaviour of the fusion protein, F64L-S65T-GFP alone was expressed in CHO cells and these were also given 50 µM forskolin (n=5); the uniform diffuse distribution characteristic of GFP in these cells was unaffected by such treatment (data not shown).

The forskolin-induced translocation of PKAc-F64L-S65T-GFP showed a dose-response relationship (Fig 4 and 6), see quantitative procedures above.

Reversibility of PKAc-F64L-S65T-GFP translocation.

- The release of the PKAc probe from its cytoplasmic anchoring hotspots was reversible. Washing the cells repeatedly (5-8 times) with buffer after 10µM forskolin treatment completely restored the punctate pattern within 2-5 min (n=2, Fig. 3E). In fact the fusion protein returned to a pattern of fluorescent cytoplasmic aggregates virtually indistinguishable from that observed before forskolin stimulation.
- To test whether the return of fusion protein to the cytoplasmic aggregates reflected a decreased [cAMP], cells were treated with a combination of 10 μM forskolin and 100 μM IBMX (n=2) then washed repeatedly (5-8 times) with buffer containing 100 μM IBMX (Fig. 3F). In these experiments, the fusion protein did not return to its prestimulatory localisation after removal of forskolin.

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Testing the PKA-F64L-S65T-GFP probe with physiologically relevant agents.

To test the probe's response to receptor activation of adenylate cyclase, BHK cells stably transfected with the glucagon receptor and the PKA-F64L-S65T-GFP probe were exposed to glucagon stimulation. The glucagon receptor is coupled to a  $G_s$  protein which activates adenylate cyclase, thereby increasing the cAMP level. In these cells, addition of 100 nM glucagon (n=2) caused the release of the PKA-F64L-S65T-GFP probe from the cytoplasmic aggregates and a resulting translocation of the fusion protein to a more even cytoplasmic distribution within 2-3 min (Fig. 3G). Similar but less pronounced effects were seen at lower glucagon concentrations (n=2, data not shown). Addition of buffer (n=2) had no effect over time (data not shown).

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10 Transiently transfected CHO cells expressing hARα2a and the PKA-F64L-S65T-GFP probe were treated with 10 μM forskolin for 7.5 minutes, then, in the continued presence of forskolin, exposed to 10 μM norepinephrine to stimulate the exogenous adrenoreceptors, which couple to a G<sub>1</sub> protein, which inhibit adenylate cyclase. This treatment led to reappearance of fluorescence in the cytoplasmic aggregates indicative of a decrease in [cAMP] (Fig. 3H).

# Fusion protein translocation correlated with [cAMP],

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As described above, the time it took for a response to come to completion was dependent on the forskolin dose (Fig. 5) In addition the degree of responses was also dose-dependent. To test the PKA-F64L-S65T-GFP fusion protein translocation in a semi high through-put system, CHO cells stably transfected with the PKA-F64L-S65T-GFP fusion was stimulated with buffer and 5 increasing doses of forskolin (n=8). Using the image analysis algorithm described above (Method 1), a dose-response relationship was observed in the range from 0.01-50 μM forskolin (Fig. 6). A half-maximal stimulation was observed at about 2 μM forskolin. In parallel, cells were stimulated with buffer and 8 increasing concentrations of forskolin (n=4) in the range 0.01-50 μM. The amount of cAMP produced was measured in an SPA assay. A steep increase was observed between 1 and 5 μM forskolin coincident with the steepest part of the curve for fusion protein translocation (also Fig. 6).

#### EXAMPLE 2

# Quantitation of redistribution in real-time within living cells.

Probe for detection of PKC activity in real time within living cells:

5 Construction of PKC-GFP fusion:

The probe was constructed by ligating two restriction enzyme treated polymerase chain reaction (PCR) amplification products of the cDNA for murine PKCα (GenBank Accession number: M25811) and F64L-S65T-GFP (sequence disclosed in WO 97/11094) respectively. Taq® polymerase and the following oligonucleotide primers were used for PCR;

5'mPKCa:

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TTggACACAAgCTTTggACACCCTCAggATATggCTgACgTTTACCCggCCAACg (SEQ ID NO:5),

3'mPKCa:

gTCATCTTCTCgAgTCTTTCAggCgCgCCCTACTgCACTTTgCAAgATTgggTgC (SEQ ID NO:6),

5'F64L-S65T-GFP:

TTggACACAAgCTTTggACACggCgCCCATgAgTAAAggAgAAACTTTTC (SEQ ID NO:1),

20 3'F64L-S65T-GFP:

gTCATCTTCTCgAgTCTTACTCCTgAggTTTgTATAgTTCATCCATgCCATgT (SEQ ID NO:2).

The hybrid DNA strand was inserted into the pZeoSV® mammalian expression vector as a HindIII-XhoI casette as described in example 1.

25 BHK cells expressing the human M1 receptor under the control of the inducible metallothionine promoter and maintained with the dihydrofolate reductase marker were

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transfected with the PKCa-F64L-S65T-GFP probe using the calcium phosphate precipitate method in HEPES buffered saline (HBS [pH 7.10]). Stable transfectants were selected using 1000 µg Zeocin®/ml in the growth medium (DMEM with 1000 mg glucose/l, 10 % foetal bovine serum (FBS), 100 µg penicillin-streptomycin mixture ml-1, 2 mM l- glutamine). The hM1 receptor and PKCa-F64L-S65T-GFP fusion protein were maintained with 500 nM methotrexate and 500 µg Zeocin®/ml respectively. 24 hours prior to any experiment, the cells were transferred to HAM F-12 medium with glutamax, 100 µg penicillin-streptomycin mixture ml<sup>-1</sup> and 0.3 % FBS. This medium relieves selection pressure, gives a low induction of signal transduction pathways and has a low autofluorescence at the relevant wavelength enabling fluorescence microscopy of cells straight from the incubator.

### Method 1: Monitoring the PKCα activity in real time:

Digital images of live cells were gathered using a Zeiss Axiovert 135M fluorescence microscope fitted with a 40X, NA: 1.3 oil immersion objective and coupled to a Photometrics CH250 charged coupled device (CCD) camera. The cells were illuminated with a 100 W arc lamp. In the light path was a 470±20 nm excitation filter, a 510 nm dichroic mirror and a 515±15 nm emission filter for minimal image background. The cells were kept and monitored to be at 37°C with a custom built stage heater.

20 Images were analyzed using the IPLab software package for Macintosh.

Upon stimulation of the M1-BHK cells, stably expressing the PKCα-F64L-S65T-GFP fusion, with carbachol we observed a dose-dependent transient translocation from the cytoplasm to the plasma membrane (Fig. 7a,b,c). Simultaneous measurement of the cytosolic free calcium concentration shows that the carbachol-induced calcium mobilisation precedes the translocation (Fig. 8).

Stepwise procedure for quantitation of translocation of PKCa:

- 1. The image was corrected for dark current by performing a pixel-by-pixel subtraction of a dark image (an image taken under the same conditions as the actual image, except the camera shutter is not allowed to open).
- The image was corrected for non-uniformity of the illumination by performing a
   pixel-by-pixel ratio with a flat field correction image (an image taken under the same conditions as the actual image of a uniformly fluorescent specimen).
  - 3. A copy of the image was made in which the edges are identified. The edges in the image are found by a standard edge-detection procedure convolving the image with a kernel which removes any large-scale unchanging components (i.e., background) and accentuates any small-scale changes (i.e., sharp edges). This image was then converted to a binary image by threshholding. Objects in the binary image which are too small to represent the edges of cells were discarded. A dilation of the binary image was performed to close any gaps in the image edges. Any edge objects in the image which were in contact with the borders of the image are discarded. This binary image represents the edge mask.
  - 4. Another copy of image was made via the procedure in step 3. This copy was further processed to detect objects which enclose "holes" and setting all pixels inside the holes to the binary value of the edge, i.e., one. This image represents the whole cell mask.
- 5. The original image was masked with the edge mask from step 3 and the sum total of all pixel values is determined.
  - 6. The original image was masked with the whole cell mask from step 4 and the sum total of all pixel values was determined.
- 7. The value from step 5 was divided by the value from step 6 to give the final result, the fraction of fluorescence intensity in the cells which was localized in the edges.

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Probes for detection of mitogen activated protein kinase Erk1 redistribution.

Useful for monitoring signalling pathways involving MAPK, e.g. to identify compounds which modulate the activity of the pathway in living cells.

Erk1, a serine/threonine protein kinase, is a component of a signalling pathway that is activated by e.g. many growth factors.

Probes for detection of ERK-1 activity in real time within living cells:

The extracellular signal regulated kinase (ERK-1, a mitogen activated protein kinase, MAPK) is fused N- or C-terminally to a derivative of GFP. The resulting fusions expressed in different mammalian cells are used for monitoring *in vivo* the nuclear translocation, and thereby the activation, of ERK1 in response to stimuli that activate the MAPK pathway.

a) Construction of murine ERK1 - F64L-S65T-GFP fusion:

Convenient restriction endonuclease sites are introduced into the cDNAs encoding murine ERK1 (GenBank Accession number: Z14249) and F64L-S65T-GFP (sequence disclosed in WO 97/11094) by polymerase chain reaction (PCR). The PCR reactions are performed according to standard protocols with the following primers:

#### 5'ERK1:

TTggACACAAgCTTTggACACCCTCAggATATggCggCggCggCggCggCTCCgggggg Cggg (SEQ ID NO:7),

20 3'ERK1:

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5'F64L-S65T-GFP:

TTggACACAAgCTTTggACACggCgCCCATgAgTAAAggAgAAACTTTTC

(SEQ ID NO:1)

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3'F64L-S65T-GFP:

gTCATCTTCTCgAgTCTTACTCCTgAggTTTgTATAgTTCATCCATgCCATgT (SEQ ID NO:2)

To generate the mERK1-F64L-S65T-GFP (SEQ ID NO:56 & 57) fusion the ERK1 amplification product is digested with HindIII+AscI and the F64L-S65T-GFP product with AscI+XhoI. To generate the F64L-S65T-GFP-mERK1 fusion the ERK1 amplification product is then digested with HindIII+Bsu36I and the F64L-S65T-GFP product with Bsu36I+XhoI. The two pairs of digested PCR products are subsequently ligated with a HindIII+XhoI digested plasmid (pZeoSV® mammalian expression vector, Invitrogen, San Diego, CA, USA). The resulting fusion constructs are under control of the SV40 promoter.

b) The human Erk1 gene (GenBank Accession number: X60188) was amplified using PCR according to standard protocols with primers Erk1-top (SEQ ID NO:9) and Erk1-bottom/+stop (SEQ ID NO:10). The PCR product was digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with EcoR1 and BamH1. This produces an EGFP-Erk1 fusion (SEQ ID NO:38 &39) under the control of a CMV promoter.

The plamid containing the EGFP-Erk1 fusion was transfected into HEK293 cells employing the FUGENE transfection reagent (Boehringer Mannheim). Prior to experiments the cells were grown to 80%-90% confluency 8 well chambers in DMEM with 10% FCS. The cells were washed in plain HAM F-12 medium (without FCS), and then incubated for 30-60 minutes in plain HAM F-12 (without FCS) with 100 micromolar PD98059, an inhibitor of MEK1, a kinase which activates Erk1; this step effectively empties the nucleus of EGFP-Erk1. Just before starting the experiment, the HAM F-12 was replaced with Hepes buffer following a wash with Hepes buffer. This removes the PD98059 inhibitor; if blocking of MEK1 is still wanted (e.g. in control experiments), the inhibitor is included in the Hepes buffer.

The experimental setup of the microscope was as described in example 1.

60 images were collected with 10 seconds between each, and with the test compound added after image number 10.

Addition of EGF (1-100 nM) caused within minutes a redistribution of EGFP-Erk1 from the cytoplasm into the nucleus (Fig. 9a,b).

The response was quantitated as described below and a dose-dependent relationship between EGF concentration and nuclear translocation of EGFP-Erk1 was found (Fig. 9c,d). Redistribution of GFP fluorescence is expressed in this example as the change in the ratio value between areas in nuclear versus cytoplasmic compartments of the cell. Each time profile is the average of nuclear to cytoplasmic ratios from six cells in each treatment.

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### **EXAMPLE 4**

### Probes for detection of Erk2 redistribution.

Useful for monitoring signalling pathways involving MAPK, e.g. to identify compounds which modulate the activity of the pathway in living cells.

- 15 Erk2, a serine/threonine protein kinase, is closely related to Erk1 but not identical; it is a component of a signalling pathway that is activated by e.g. many growth factors.
  - a) The rat Erk2 gene (GenBank Accession number: M64300) was amplified using PCR according to standard protocols with primers Erk2-top (SEQ ID NO:11) and Erk2-bottom/+stop (SEQ ID NO:13) The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-Erk2 fusion (SEQ ID NO:40 &41) under the control of a CMV promoter.
  - b) The rat Erk2 gene (GenBank Accession number: M64300) was amplified using PCR according to standard protocols with primers (SEQ ID NO:11) Erk2-top and Erk2-bottom/-stop (SEQ ID NO:12). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank

Accession number U55762) digested with Xho1 and BamH1. This produces an Erk2-EGFP fusion (SEQ ID NO:58 &59) under the control of a CMV promoter.

The resulting plasmids were transfected into CHO cells and BHK cells. The cells were grown under standard conditions. Prior to experiments, the cells were starved in medium without serum for 48-72 hours. This led to a predominantly cytoplasmic localisation of both probes, especially in BHK cells. 10% fetal calf serum was added to the cells and the fluorescence of the cells was recorded as explained in example 3. Addition of serum caused the probes to redistribute into the nucleus within minutes of addition of serum.

## 10 EXAMPLE 5

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### Probes for detection of Smad2 redistribution.

Useful for monitoring signalling pathways activated by some members of the transforming growth factor-beta family, e.g. to identify compounds which modulate the activity of the pathway in living cells.

- Smad 2, a signal transducer, is a component of a signalling pathway that is induced by some members of the TGFbeta family of cytokines.
  - a) The human Smad2 gene (GenBank Accession number: AF027964) was amplified using PCR according to standard protocols with primers Smad2-top (SEQ ID NO:24) and Smad2-bottom/+stop (SEQ ID NO:26). The PCR product was digested with restriction enzymes EcoR1 and Acc65I, and ligated into pEGFP-C1 (Clontech; Palo Alto; GenBank Accession number U55763) digested with EcoR1 and Acc65I. This produces an EGFP-Smad2 fusion (SEQ ID NO:50&51) under the control of a CMV promoter.
- b) The human Smad2 gene (GenBank Accession number: AF027964) was amplified using PCR according to standard protocols with primers Smad2-top (SEQ ID NO:24) and
   Smad2-bottom/-stop (SEQ ID NO:25). The PCR product was digested with restriction enzymes EcoR1 and Acc65I, and ligated into pEGFP-N1 (Clontech, Palo Alto;

GenBank Accession number U55762) digested with EcoR1 and Acc65I. This produces a Smad2-EGFP fusion (SEQ ID NO:74 &75) under the control of a CMV promoter.

The plasmid containing the EGFP-Smad2 fusion was transfected into HEK293 cells, where it showed a cytoplasmic distribution. Prior to experiments the cells were grown in 8 well Nunc chambers in DMEM with 10% FCS to 80% confluence and starved overnight in HAM F-12 medium without FCS.

For experiments, the HAM F-12 medium was replaced with Hepes buffer pH 7.2.

The experimental setup of the microscope was as described in example 1.

90 images were collected with 10 seconds between each, and with the test compound added after image number 5.

After serum starvation of cells, each nucleus contains less GFP fluorescence than the surrounding cytoplasm (Fig. 10a). Addition of TGFbeta caused within minutes a redistribution of EGFP-Smad2 from the cytoplasma into the nucleus (Fig. 10b).

The redistribution of fluorescence within the treated cells was quantified simply as the fractional increase in nuclear fluorescence normalised to the starting value of GFP fluorescence in the nucleus of each unstimulated cell.

### **EXAMPLE 6**

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### Probe for detection of VASP redistribution.

Useful for monitoring signalling pathways involving rearrangement of cytoskeletal elements, e.g. to identify compounds which modulate the activity of the pathway in living cells.

VASP, a phosphoprotein, is a component of cytoskeletal structures, which redistributes in response to signals that affect focal adhesions.

The human VASP gene (GenBank Accession number: Z46389) was amplified using PCR according to standard protocols with primers VASP-top (SEQ ID NO:94) and VASP-bottom/+stop (SEQ ID NO:95). The PCR product was digested with restriction enzymes Hind3 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Hind3and BamH1. This produces an EGFP-VASP fusion (SEQ ID NO:124 &125) under the control of a CMV promoter.

The resulting plasmid was transfected into CHO cells expressing the human insulin receptor using the calcium-phosphate transfection method. Prior to experiments, cells were grown in 8 well Nunc chambers and starved overnight in medium without FCS.

Experiments are performed in a microscope setup as described in example 1.

10% FCS was added to the cells and images were collected. The EGFP-VASP fusion was redistributed from a somewhat even distribution near the periphery into more localised structures, identified as focal adhesion points (Fig. 11).

A large number of further GFP fusions have been made or are in the process of being made, as apparent from the following Examples 7-22 which also suggest suitable host cells and substances for activation of the cellular signalling pathways to be monitored and analyzed.

# **EXAMPLE 7**

# 20 Probe for detection of actin redistribution.

Useful for monitoring signalling pathways involving rearrangement or formation of actin filaments, e.g. to identify compounds which modulate the activity of pathways leading to cytoskeletal rearrangements in living cells.

Actin is a component of cytoskeletal structures, which redistributes in response to very many cellular signals.

The actin binding domain of the human alpha-actinin gene (GenBank Accession number:

X15804) was amplified using PCR according to standard protocols with primers ABD-top (SEQ ID NO:90) and ABD-bottom/-stop (SEQ ID NO:91). The PCR product was digested with restriction enzymes Hind3 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Hind3 and BamH1. This produced an actin-binding-domain-EGFP fusion (SEQ ID NO:128 &129) under the control of a CMV promoter.

The resulting plasmid was transfected into CHO cells expressing the human insulin receptor. Cells were stimulated with insulin that caused the actin binding domain-EGFP probe to become redistributed into morphologically distinct membrane-associated structures.

### **EXAMPLE 8**

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### Probes for detection of p38 redistribution.

Useful for monitoring signalling pathways responding to various cellular stress situations,
e.g. to identify compounds which modulate the activity of the pathway in living cells, or as
a counterscreen.

p38, a serine/threonine protein kinase, is a component of a stress-induced signalling pathway which is activated by many types of cellular stress, e.g. TNFalpha, anisomycin, UV and mitomycin C.

- a) The human p38 gene (GenBank Accession number: L35253) was amplified using PCR according to standard protocols with primers p38-top (SEQ ID NO:14) and p38-bottom/+stop (SEQ ID NO: 16). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produced an EGFP-p38 fusion (SEQ ID NO:46 & 47) under the control of a CMV promoter.
  - b) The human p38 gene (GenBank Accession number: L35253) was amplified using PCR according to standard protocols with primers p38-top (SEQ ID NO:13) and p38-

bottom/-stop (SEQ ID NO:15). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produced a p38-EGFP fusion (SEQ ID NO:64 & 65) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. HEK293, in which the EGFP-p38 probe and/or the p38-EGFP probe should change its cellular distribution from predominantly cytoplasmic to nuclear within minutes in response to activation of the signalling pathway with e.g. anisomycin.

### 10 EXAMPLE 9

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### Probes for detection of Jnk1 redistribution.

Useful for monitoring signalling pathways responding to various cellular stress situations, e.g. to identify compounds which modulate the activity of the pathway in living cells, or as a counterscreen.

- Jnk1, a serine/threonine protein kinase, is a component of a stress-induced signalling pathway different from the p38 described above, though it also is activated by many types of cellular stress, e.g. TNFalpha, anisomycin and UV.
  - a) The human Jnk1 gene (GenBank Accession number: L26318) was amplified using PCR according to standard protocols with primers Jnk-top (SEQ ID NO:17) and Jnk-bottom/+stop (SEQ ID NO:19). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produced an EGFP-Jnk1 fusion (SEQ ID NO:44 &45) under the control of a CMV promoter.
  - b) The human Jnk1 gene (GenBank Accession number: L26318) was amplified using PCR according to standard protocols with primers Jnk-top (SEQ ID NO:17) and Jnk-bottom/-stop (SEQ ID NO:18). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank

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Accession number U55762) digested with Xho1 and BamH1. This produced a Jnk1-EGFP fusion (SEQ ID NO:62 &63) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. HEK293, in which the EGFP-Jnk1 probe and/or the Jnk1-EGFP probe should change its cellular distribution from predominantly cytoplasmic to nuclear in response to activation of the signalling pathway with e.g. anisomycin.

### **EXAMPLE 10**

### Probes for detection of PKG redistribution.

- Useful for monitoring signalling pathways involving changes in cyclic GMP levels, e.g. to identify compounds which modulate the activity of the pathway in living cells.
  - PGK, a cGMP-dependent serine/threonine protein kinase, mediates the guanylyl-cyclase/cGMP signal.
  - a) The human PKG gene (GenBank Accession number: Y07512) is amplified using PCR according to standard protocols with primers PKG-top (SEQ ID NO:81) and PKG-bottom/+stop (SEQ ID NO:83). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-PKG fusion (SEQ ID NO:134 &135) under the control of a CMV promoter.
- b) The human PKG gene (GenBank Accession number: Y07512) is amplified using PCR according to standard protocols with primers PKG-top (SEQ ID NO:81) and PKG-bottom/-stop (SEQ ID NO: 82). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produces a PKG-EGFP fusion (SEQ ID NO:136 &137) under the control of a CMV promoter.
  - The resulting plasmids are transfected into a suitable cell line, e.g. A10, in which the EGFP-PKG probe and/or the PKG-EGFP probe should change its cellular distribution

from cytoplasmic to one associated with cytoskeletal elements within minutes in response to treatment with agents which raise nitric oxide (NO) levels.

## **EXAMPLE 11**

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# 5 Probes for detection of IkappaB kinase redistribution.

Useful for monitoring signalling pathways leading to NFkappaB activation, e.g. to identify compounds which modulate the activity of the pathway in living cells.

IkappaB kinase, a serine/threonine kinase, is a component of a signalling pathway which is activated by a variety of inducers including cytokines, lymphokines, growth factors and stress.

- a) The alpha subunit of the human IkappaB kinase gene (GenBank Accession number: AF009225) is amplified using PCR according to standard protocols with primers IKK-top (SEQ ID NO:96) and IKK-bottom/+stop (SEQ ID NO:98). The PCR product is digested with restriction enzymes EcoR1 and Acc65I, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with EcoR1and Acc65I. This produces an EGFP-IkappaB-kinase fusion (SEQ ID NO:120 &121) under the control of a CMV promoter.
- b) The alpha subunit of the human IkappaB kinase gene (GenBank Accession number: AF009225) is amplified using PCR according to standard protocols with primers IKK-top (SEQ ID NO:96) and IKK-bottom/-stop (SEQ ID NO:97). The PCR product is digested with restriction enzymes EcoR1 and Acc651, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and Acc651. This produces an IkappaB-kinase-EGFP fusion (SEQ ID NO:122 &123) under the control of a CMV promoter.
- The resulting plasmids are transfected into a suitable cell line, e.g. Jurkat, in which the EGFP-IkappaB-kinase probe and/or the IkappaB-kinase-EGFP probe should achieve a more cytoplasmic distribution within seconds following stimulation with e.g. TNFalpha.

### **EXAMPLE 12**

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#### Probes for detection of CDK2 redistribution.

Useful for monitoring signalling pathways of the cell cycle, e.g. to identify compounds that modulate the activity of the pathway in living cells.

CDK2, a cyclin-dependent serine/threonine kinase, is a component of the signalling system that regulates the cell cycle.

- a) The human CDK2 gene (GenBank Accession number: X61622) is amplified using PCR according to standard protocols with primers CDK2-top (SEQ ID NO:102) and CDK2-bottom/+stop (SEQ ID NO: 104). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-CDK2 fusion (SEQ ID NO:114 &115) under the control of a CMV promoter.
- b) The human CDK2 gene (GenBank Accession number: X61622) is amplified using PCR according to standard protocols with primers CDK2-top (SEQ ID NO:102) and CDK2-bottom/-stop (SEQ ID NO:103). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produces a CDK2-EGFP fusion (SEQ ID NO:112 &113) under the control of a CMV promoter.
- The resulting plasmids are transfected into a suitable cell line, e.g. HEK293 in which the EGFP-CDK2 probe and/or the CDK2-EGFP probe should change its cellular distribution from cytoplasmic in contact-inhibited cells, to nuclear location in response to activation with a number of growth factors, e.g. IGF.

### 25 EXAMPLE 13

Probes for detection of Grk5 redistribution.

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Useful for monitoring signalling pathways involving desensitisation of G-protein coupled receptors, e.g. to identify compounds which modulate the activity of the pathway in living cells.

Grk5, a G-protein coupled receptor kinase, is a component of signalling pathways involving membrane bound G-protein coupled receptors.

- a) The human Grk5 gene (GenBank Accession number: L15388) is amplified using PCR according to standard protocols with primers Grk5-top (SEQ ID NO:27) and Grk5-bottom/+stop (SEQ ID NO:29). The PCR product is digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with EcoR1 and BamH1. This produces an EGFP-Grk5 fusion (SEQ ID NO:42 &43) under the control of a CMV promoter.
- b) The human Grk5 gene (GenBank Accession number: L15388) is amplified using PCR according to standard protocols with primers Grk5-top (SEQ ID NO:27) and Grk5-bottom/-stop (SEQ ID NO:28). The PCR product is digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and BamH1. This produces a Grk5-EGFP fusion (SEQ ID NO:60 &61) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. HEK293 expressing a rat dopamine D1A receptor, in which the EGFP-Grk5 probe and/or the Grk5-EGFP probe should change its cellular distribution from predominantly cytoplasmic to peripheral in response to activation of the signalling pathway with e.g. dopamine.

# **EXAMPLE 14**

### 25 Probes for detection of Zap70 redistribution.

Useful for monitoring signalling pathways involving the T cell receptor, e.g. to identify compounds which modulate the activity of the pathway in living cells.

Zap70, a tyrosine kinase, is a component of a signalling pathway which is active in e.g. T-cell differentiation.

- a) The human Zap70 gene (GenBank Accession number: L05148) is amplified using PCR according to standard protocols with primers Zap70-top (SEQ ID NO:105) and Zap70-bottom/+stop (SEQ ID NO:107). The PCR product is digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-C1 (GenBank Accession number U55763) digested with EcoR1 and BamH1. This produces an EGFP-Zap70 fusion (SEQ ID NO:108 &109) under the control of a CMV promoter.
- b) The human Zap70 gene (GenBank Accession number: L05148) is amplified using PCR according to standard protocols with primers Zap70-top (SEQ ID NO:105) and Zap70-bottom/-stop (SEQ ID NO:106). The PCR product is digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and BamH1. This produces a Zap70-EGFP fusion (SEQ ID NO:110 &111) under the control of a CMV promoter.
- The resulting plasmids are transfected into a suitable cell line, e.g. Jurkat, in which the EGFP-Zap70 probe and/or the Zap70-EGFP probe should change its cellular distribution from cytoplasmic to membrane-associated within seconds in response to activation of the T cell receptor signalling pathway with e.g. antibodies to CD3epsilon.

# 20 EXAMPLE 15

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# Probes for detection of p85 redistribution.

Useful for monitoring signalling pathways involving PI-3 kinase, e.g. to identify compounds which modulate the activity of the pathway in living cells.

p85alpha is the regulatory subunit of PI3-kinase which is a component of many pathways involving membrane-bound tyrosine kinase receptors and G-protein-coupled receptors.

a) The human p85alpha gene (GenBank Accession number: M61906) was amplified using PCR according to standard protocols with primers p85-top-C (SEQ ID NO:22) and p85-

bottom/+stop (SEQ ID NO:23). The PCR product was digested with restriction enzymes Bgl2 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Bgl2 and BamH1. This produced an EGFP-p85alpha fusion (SEQ ID NO:48 &49) under the control of a CMV promoter.

b) The human p85alpha gene (GenBank Accession number: M61906) was amplified using PCR according to standard protocols with primers p85-top-N (SEQ ID NO:20) and p85-bottom/-stop (SEQ ID NO:21). The PCR product was digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and BamH1. This produced a p85alpha-EGFP fusion (SEQ ID NO:66 &67) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. CHO expressing the human insulin receptor, in which the EGFP-p85 probe and/or the p85-EGFP probe may change its cellular distribution from cytoplasmic to membrane-associated within minutes in response to activation of the receptor with insulin.

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# **EXAMPLE 16**

# Probes for detection of protein-tyrosine phosphatase redistribution.

Useful for monitoring signalling pathways involving tyrosine kinases, e.g. to identify compounds which modulate the activity of the pathway in living cells.

- 20 Protein-tyrosine phosphatase1C, a tyrosine-specific phosphatase, is an inhibitory component in signalling pathways involving e.g. some growth factors.
  - a) The human protein-tyrosine phosphatase 1C gene (GenBank Accession number: X62055) is amplified using PCR according to standard protocols with primers PTP-top (SEQ ID NO:99) and PTP-bottom/+stop (SEQ ID NO:101). The PCR product is digested with restriction enzymes Xho1 and EcoR1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and EcoR1. This produces an EGFP-PTP fusion (SEQ ID NO:116 & 117) under the control

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of a CMV promoter.

b) The human protein-tyrosine phosphatase 1C gene (GenBank Accession number: X62055) is amplified using PCR according to standard protocols with primers PTP-top (SEQ ID NO:99) and PTP-bottom/-stop (SEQ ID NO:100). The PCR product is digested with restriction enzymes Xho1 and EcoR1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and EcoR1. This produces a PTP-EGFP fusion (SEQ ID NO:118 & 119) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. MCF-7 in which the

EGFP-PTP probe and/or the PTP-EGFP probe should change its cellular distribution from
cytoplasm to the plasma menbrane within minutes in response to activation of the growth
inhibitory signalling pathway with e.g. somatostatin.

### **EXAMPLE 17**

# 15 Probes for detection of Smad4 redistribution.

Useful for monitoring signalling pathways involving most members of the transforming growth factor-beta family, e.g. to identify compounds which modulate the activity of the pathway in living cells.

Smad4, a signal transducer, is a common component of signalling pathways induced by various members of the TGFbeta family of cytokines.

- a) The human Smad4 gene (GenBank Accession number: U44378) was amplified using PCR according to standard protocols with primers Smad4-top and Smad4-bottom/+stop (SEQ ID NO:35). The PCR product was digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with EcoR1 and BamH1. This produce an EGFP-Smad4 fusion (SEQ ID NO:52 & 53) under the control of a CMV promoter.
- b) The human Smad4 gene (GenBank Accession number: U44378) was amplified using

PCR according to standard protocols with primers Smad4-top (SEQ ID NO:33) and Smad4-bottom/-stop (SEQ ID NO:34). The PCR product was digested with restriction enzymes EcoR1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and BamH1. This produced a Smad4-EGFP fusion (SEQ ID NO:76 & 77) under the control of a CMV promoter.

The resulting plasmids are transfected into a cell line, e.g. HEK293 in which the EGFP-Smad4 probe and/or the Smad4-EGFP probe should change its cellular distribution within minutes from cytoplasmic to nuclear in response to activation of the signalling pathway with e.g. TGFbeta.

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### **EXAMPLE 18**

# Probes for detection of Stat5 redistribution.

Useful for monitoring signalling pathways involving the activation of tyrosine kinases of the Jak family, e.g. to identify compounds that modulate the activity of the pathway in living cells.

Stat5, signal transducer and activator of transcription, is a component of signalling pathways that are induced by e.g. many cytokines and growth factors.

- a) The human Stat5 gene (GenBank Accession number: L41142) was amplified using PCR according to standard protocols with primers Stat5-top (SEQ ID NO:30) and Stat5-bottom/+stop (SEQ ID NO:32). The PCR product was digested with restriction enzymes Bgl2 and Acc65I, and ligated into pEGFP-C1 (Clontech; Palo Alto; GenBank Accession number U55763) digested with Bgl2 and Acc65I. This produced an EGFP-Stat5 fusion (SEQ ID NO:54 & 55) under the control of a CMV promoter.
- b) The human Stat5 gene (GenBank Accession number: L41142) was amplified using PCR according to standard protocols with primers Stat5-top (SEQ ID NO:30) and Stat5-bottom/-stop (SEQ ID NO:331). The PCR product was digested with restriction

enzymes Bgl2 and Acc65I, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Bgl2 and Acc65I. This produced a Stat5-EGFP fusion (SEQ ID NO:78 & 79) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. MIN6 in which the EGFP-Stat5 probe and/or the Stat5-EGFP probe should change its cellular distribution from cytoplasmic to nuclear within minutes in response to activation signalling pathway with e.g. prolactin.

#### **EXAMPLE 19**

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## 10 Probes for detection of NFAT redistribution.

Useful for monitoring signalling pathways involving activation of NFAT, e.g. to identify compounds which modulate the activity of the pathway in living cells.

NFAT, an activator of transcription, is a component of signalling pathways involved in e.g. immune responses.

- a) The human NFAT1 gene (GenBank Accession number: U43342) is amplified using PCR according to standard protocols with primers NFAT-top (SEQ ID NO:84) and NFAT-bottom/+stop (SEQ ID NO:86). The PCR product is digested with restriction enzymes Xho1 and EcoR1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and EcoR1. This produces an EGFP-NFAT fusion (SEQ ID NO:130 & 131) under the control of a CMV promoter.
  - b) The human NFAT gene (GenBank Accession number: U43342) is amplified using PCR according to standard protocols with primers NFAT-top (SEQ ID NO:84) and NFAT-bottom/-stop (SEQ ID NO:85). The PCR product is digested with restriction enzymes Xho1 and EcoR1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and EcoR1. This produces an NFAT-EGFP fusion (SEQ ID NO:132 & 133) under the control of a CMV promoter.

The resulting plasmids are transfected into a suitable cell'line, e.g. Jurkat, in which the

EGFP-NFAT probe and/or the NFAT-EGFP probe should change its cellular distribution from cytoplasmic to nuclear within minutes in response to activation of the signalling pathway with e.g. antibodies to CD3epsilon.

### 5 EXAMPLE 20

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### Probes for detection of NFkappaB redistribution.

Useful for monitoring signalling pathways leading to activation of NFkappaB, e.g. to identify compounds which modulate the activity of the pathway in living cells.

NFkappaB, an activator of transcription, is a component of signalling pathways that are responsive to a varity of inducers including cytokines, lymphokines, and some immunosuppressive agents.

- a) The human NFkappaB p65 subunit gene (GenBank Accession number: M62399) is amplified using PCR according to standard protocols with primers NFkappaB-top (SEQ ID NO:87) and NFkappaB-bottom/+stop (SEQ ID NO:89). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-NFkappaB fusion (SEQ ID NO:142 & 143) under the control of a CMV promoter.
- b) The human NFkappaB p65 subunit gene (GenBank Accession number: M62399) is
   amplified using PCR according to standard protocols with primers NFkappaB-top (SEQ ID NO:87) and NFkappaB-bottom/-stop (SEQ ID NO:88). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produces an NFkappaB-EGFP fusion (SEQ ID NO:140 & 141) under the control of a
   CMV promoter.

The resulting plasmids are transfected into a suitable cell line, e.g. Jurkat, in which the EGFP-NFkappaB probe and/or the NFkappaB-EGFP probe should change its cellular

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distribution from cytoplasmic to nuclear in response to activation of the signalling pathway with e.g. TNFalpha.

### **EXAMPLE 21**

## 5 Probe for detection of RhoA redistribution.

Useful for monitoring signalling pathways involving RhoA, e.g. to identify compounds which modulate the activity of the pathway in living cells.

RhoA, a small GTPase, is a component of many signalling pathways, e.g. LPA induced cytoskeletal rearrangements.

The human RhoA gene (GenBank Accession number: L25080) was amplified using PCR according to standard protocols with primers RhoA-top (SEQ ID NO:92) and RhoA-bottom/+stop (SEQ ID NO:93). The PCR product was digested with restriction enzymes Hind3 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Hind3and BamH1. This produced an EGFP-RhoA fusion (SEQ ID NO:126 &127) under the control of a CMV promoter.

The resulting plasmid is transfected into a suitable cell line, e.g. Swiss3T3, in which the EGFP-RhoA probe should change its cellular distribution from a reasonably homogenous to a peripheral distribution within minutes of activation of the signalling pathway with e.g. LPA.

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### **EXAMPLE 22**

# Probes for detection of PKB redistribution.

Useful for monitoring signalling pathways involving PKB e.g. to identify compounds which modulate the activity of the pathway in living cells.

25 PKB, a serine/threonine kinase, is a component in various signalling pathways, many of

which are activated by growth factors.

- a) The human PKB gene (GenBank Accession number: M63167) is amplified using PCR according to standard protocols with primers PKB-top (SEQ ID NO:36) and PKB-bottom/+stop (SEQ ID NO:80). The PCR product is digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-C1 (Clontech, Palo Alto; GenBank Accession number U55763) digested with Xho1 and BamH1. This produces an EGFP-PKB fusion (SEQ ID NO:138 & 139) under the control of a CMV promoter.
- b) The human PKB gene (GenBank Accession number: M63167) was amplified using

  PCR according to standard protocols with primers PKB-top (SEQ ID NO:36) and PKBbottom/-stop (SEQ ID NO:37). The PCR product was digested with restriction
  enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank
  Accession number U55762) digested with Xho1 and BamH1. This produced a PKBEGFP fusion (SEQ ID NO:70 &71) under the control of a CMV promoter.
- The resulting plasmids are transfected into a suitable cell line, e.g. CHO expressing the human insulin receptor, in which the EGFP-PKB probe and/or the PKB-EGFP probe cycles between cytoplasmic and membrane locations during the activation-deactivation process following addition of insulin. The transition should be apparent within minutes.

### 20 EXAMPLE 23

Measurement of the real-time redistribution of protein kinase C  $\alpha$  isoform-GFP fusion (PKC $\alpha$ -GFP) in response to carbamylcholine stimulation of the muscarinic M1 receptor; 96 parallel redistribution measurements in microtiter plates.

BHK cells were stably expressing a recombinant human muscarinic typ 1 receptor, under the selection with 500 μg/ml Methotrexate, and also a PKCα-GFP construct (KaA 048), under the selection of 500 nM Zeocin. The cells were grown in 96-well plates (Packard ViewPlate, black with transparent bottom), washed and preincubated in a Hank's Buffered

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Salt solution (HBSS) without phenol red, with 20 mM HEPES and 5.5 mM glucose.

The plate was measured in a FLIPR<sup>TM</sup> (Fluorescence Imaging Plate Reader) from Molecular Devices. The 488 nm emission line from an argon ion laser, run at between 0.4 and 0.8 W output, was used to excite fluorescence form the GFP. Emission wavelengths were collected through a 510 to 565 nm band pass filter.

The cells were challenged with three doses of carbamylcholine, an M1 receptor agonist known from previous studies to give a microscopically detectable redistribution of the PKCα-GFP construct [(Almholt et al. 1997)]. Measurements were made every 10 seconds for 5 minutes. After data handling including normalisation of baseline fluorescence for the different wells, background subtraction and averaging the 6 wells used for each concentration the data presented in figure 14 were obtained. It can clearly be seen (Fig 14) that carbamylcholine gave a time- and dose-dependent, and transient, decrease in fluorescence very similar to the time- and dose-dependent profile seen in microscopic fluorescence measurements [(see Almholt et al. 1997)]. This experiment was repeated twice on the same batch of cells with similar results.

### **EXAMPLE 24**

Measurement of the real-time redistribution of cyclic-AMP dependent protein kinase catalytic subuit-GFP fusion (C-GFP<sup>LT</sup>) in response to forskolin stimulation of the adenylate cyclase; 96 parallel redistribution measurements in microtiter plates.

CHO cells were stably transfected with hybrid DNA for the PKA catalytic subunit-F64L+S65T GFP (C-GFP<sup>LT</sup>) fusion protein, and were typically under continuous selection with 1000  $\mu$ g/ml zeocin (Invitrogen). The cells were grown without selection for 2 days in 96-well plates (Packard ViewPlate, black with transparent bottom), washed and preincubated in a Hank's Buffered Salt solution (HBSS) without phenol red, with 20 mM HEPES and 5.5 mM glucose.

The plate was measured in a FLIPR™ (Fluorescence Imaging Plate Reader) from Molecular Devices. The 488 nm emission line from an argon ion laser, run at between 0.4

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and 0.8 W output, was used to excite fluorescence from the GFP. Emission wavelengths were collected through a 510 to 565 nm band pass filter.

The cells were challenged with three doses of forskolin (Fig 15), an adenylate cyclase agonist known from previous studies to give a microscopically detectable redistribution of the C-GFP<sup>LT</sup> construct [(Almholt *et al.* 1998)]. Measurements were made every 10 seconds for over 6 minutes from the point of addition of forskolin. After data handling including normalisation of baseline fluorescence for the different wells, background subtraction and averaging the 6 wells used for each concentration the data presented below were obtained. It can clearly be seen in figure 15 that forskolin gave a time- and dose-dependent decrease in fluorescence very similar to the time- and dose-dependent profile seen in microscopic fluorescence measurements [(see Almholt *et al.* 1998)]. This experiment was repeated twice on the same batch of cells with similar results.

## **EXAMPLE 25**

Measurement of the redistribution response of cyclic-AMP dependent protein kinase catalytic subuit-GFP fusion (C-GFP<sup>LT</sup>) after forskolin stimulation of the adenylate cyclase; measurement of the change in total fluorescence upon permeabilisation of agonist-treated cells.

CHO cells were stably transfected with hybrid DNA for the PKA catalytic subunit- F64L+S65T GFP (C-GFP<sup>LT</sup>) fusion protein, and were typically under continuous selection with 1000  $\mu$ g/ml zeocin (Invitrogen). For the experiments reported here, cells were grown without selection to 90% confluence in 8-well tissue culture-treated Lab-Tek® chambered coverglass units (chambers, obtained from Nunc, Inc. Illinois, USA). Immediately prior to the experiment growth medium was washed from the cells and replaced with 200  $\mu$ l HEPES buffer per well.

For the results reported here, chambers were measured using a cooled CCD camera (KAF1400 chip, Photometrics Ltd., USA) attached to an inverted microscope (Diaphot 300, Nikon, Japan) equipped with a x40 oil-immersion Fluar lens, NA 1.4. Cells were

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illuminated with 450-490 nm light from a 50 W HBO lamp, and emitted light collected between 510-560 nm.

The cells were challenged with four doses of forskolin, an adenylate cyclase agonist known from previous studies to give a microscopically detectable redistribution of the C-GFP<sup>LT</sup> construct [(Almholt *et al.* 1998)]. Images were collected at 10-second intervals for a period of 10 minutes for each treatment. Six minutes after the addition of forskolin or buffer control, Triton-X100 was added to a final concentration of 0.1%. The detergent releases freely mobile C-GFP<sup>LT</sup> from the cells. The change in fluorescence resulting from this loss was measured after 1 minute of equilibration. After data handling including background subtraction and normalisation to pre-detergent values, the data presented in figure 16 were obtained. Permeabilisation caused decreases in fluorescence, the magnitude of which were dependent on the forskolin treatments. The dose-dependent profile for forskolin activation of the cAMP system as revealed by this method was very similar to that registered by other methods (see Almholt *et al.* 1998). This experiment was repeated twice on the same batch of cells with similar results.

### EXAMPLE 26

# Probe for detection of PKCbeta2 redistribution.

Useful for monitoring signalling pathways involving protein kinase C, e.g. for identifying compounds which modulate the activity of the pathway in living cells.

PKCbeta2, a serine/threonine protein kinase, is closely related to PKCalpha but not identical; it is a component of a signalling pathway that is activated by elevation of intracellular calcium concomitant with an increase in diacylglycerol species.

a) The human PKCbeta2 gene (GenBank Accession number: X07109) was amplified using PCR according to standard protocols with primers PKCbeta2-top (SEQ ID NO:162) and PKCbeta2-bottom (SEQ ID NO:163). The PCR product was digested with restriction enzymes Xho1 and BamH1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Xho1 and BamH1. This produces a PKCbeta2-

EGFP fusion (SEQ ID NO:146 & 147) under the control of a CMV promoter.

The resulting plasmids are transfected into BHK cells transfected with a human muscarinic acetylcholine receptor type M1. The cells are grown under standard conditions. The fluorescence of the cells is recorded as explained in example 3. Addition of  $1\mu M$  - $100\mu M$  carbachol causes a transient redistribution of fluorescence within the cells whereby it changes from a cytosolic location to the plasma membrane.

### **EXAMPLE 27**

# Probes for detection of PDE4D redistribution.

10 Useful for monitoring signalling pathways involving Protein Kinase A, e.g. to identify compounds which modulate the activity of the pathway in living cells.

PDE4D3, PDE4D4 and PDE4D5 are closely related splicing variants of PDE4D, a cAMP dependent phosphodiesterase. They are components of signalling pathways which involves cAMP.

The human PDE4D3, PDE4D4 and PDE4D5 genes (GenBank Accession numbers: L20970, L20969 and AF012073) are amplified using PCR according to standard protocols with the common bottom primer PDE4D-bottom (SEQ ID NO:159) and PDE4D3-top (SEQ ID NO:156), PDE4D4-top (SEQ ID NO:157) and PDE4D5-top respectively (SEQ ID NO:158) The PCR products are digested with restriction enzymes Hind3 and EcoR1, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Hind3 and EcoR1. This produces a PDE4D3-EGFP fusion (SEQ ID NO:154 & 155), a PDE4D4-EGFP fusion (SEQ ID NO:150 & 151) and a PDE4D5-EGFP fusion (SEQ ID NO:148 & 149), all three under the control of a CMV promoter.

The resulting plasmids are transfected into MVLEC cells. The cells are grown under standard conditions. The fluorescence of the cells is recorded as explained in example 3. Addition of test compounds may cause a redistribution of fluorescence within the cells from an organised cytosolic distribution to a more random one.

### EXAMPLE 28

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## Probes for detection of PDE5 redistribution.

Useful for monitoring signalling pathways involving Protein Kinase G, e.g. to identify compounds which modulate the activity of the pathway in living cells.

PDE5 is a cGMP specific phosphodiesterase. It is a component of a signalling pathway which is activated by e.g. nitric oxide.

a) The human PDE5 gene (GenBank Accession numbers: AJ004865) is amplified using PCR according to standard protocols with primers PDE5-top (SEQ ID NO:160) and PDE5-bottom (SEQ ID NO:161). The PCR product is digested with restriction enzymes EcoR1 and Acc65I, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with EcoR1 and Acc65I. This produces a PDE5-EGFP fusion (SEQ ID NO 144 & 145) under the control of a CMV promoter.

The resulting plasmids are transfected into e.g. A10 cells. The cells are grown under standard conditions. The fluorescence of the cells is recorded as explained in example 3. Addition of test compounds may cause a redistribution of fluorescence within the cells from an organized cytosolic distribution to a more random one.

## **EXAMPLE 29**

## 20 Probe for detection of Ikappa-kinase redistribution.

The human IKKbeta (GenBank Acc. No. AF031416) is amplified using PCR according to standard protocols with primers IKKbeta-top (SEQ ID NO:164) and IKKbeta-bottom (SEQ ID NO:165). The PCR product is digested with restriction enzymes Hind3 and Acc65I, and ligated into pEGFP-N1 (Clontech, Palo Alto; GenBank Accession number U55762) digested with Hind3 and Acc65I. This produces a IKKbeta-EGFP fusion (SEQ ID NO 152 & 153) under the control of a CMV promoter.

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## **EXAMPLE 30**

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# Construction of catalytically inactive Erk1 probes.

A catalytically inactive probe has the advantage that it interferes less with the normal physiology of the cell while retaining its ability to report on activation of a cellular signalling pathway by redistribution.

The Erk1 probes described above in Example 3 were subjected to site specific mutagenesis which specifically replaced the lysine at amino acid residue number 71 in the native Erk1 sequence with arginine. This mutation is known to inactivate the catalytic activity of Erk1. The redistribution patterns of the inactive Erk1 probes were identical to the original Erk1 probes, i.e. they reported on activation of the pathway by redistributing from the cytoplasm into the nucleus. The establishment of stable cell lines expressing the probe was facilitated.

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### **CLAIMS**

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- 1. A method for extracting quantitative information relating to an influence on a cellular response, the method comprising recording variation, caused by the influence on mechanically intact or permeabilised living cells, in spatially distributed light emitted from a luminophore, the luminophore being present in the cells and being capable of being redistributed in a manner which is related with the degree of the influence, and/or of being modulated by a component which is capable of being redistributed in a manner which is related to the degree of the influence, resulting in a modulation of the luminescence characteristics of the luminophore, and processing the recorded variation in the luminescence characteristics to provide quantitative information correlating the recorded variation to the degree of the influence on the cellular response.
- 2. A method according to claim 1 for extracting quantitative information relating to an influence on an intracellular pathway involving redistribution of at least one component associated with the pathway, or part thereof, the method comprising recording the result of the influence on mechanically intact or permeabilised living cells, as manifested in spatially distributed light emitted from a luminophore which is present in the cells and which is capable of being redistributed, by modulation of the pathway, in a manner which is related to the redistribution of the at least one component of the intracellular pathway, processing the recorded result to provide quantitative information correlating the change in the measured property of the light to the degree of the influence on the intracellular pathway.
  - 3. A method according to claim 1 or 2, wherein the quantitative information which is indicative of the degree of the cellular response to the influence or the result of the influence on the intracellular pathway is extracted from the recorded variation according to a predetermined calibration based on responses or results, recorded in the same manner, to known degrees of a relevant specific influence.
  - 4. A method according to any of claims 1-3, wherein the influence comprises contact between the mechanically intact or permeabilised living cells and a chemical substance and/or incubation of the mechanically intact or permeabilised living cells with a chemical substance.

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5. A method according to any of claims 1-4, wherein the influence is a substance whose effect on an intracellular pathway is to be determined.

- 6. A method according to any of claims 1-5, wherein the cells comprise a group of cells contained within a spatial limitation.
- 5 7. A method according to any of claims 1-5, wherein the cells comprise multiple groups of cells contained within multiple spatial limitations.
  - 8. A method according to any of claims 1-7, wherein the cells comprise multiple groups of cells that are qualitatively the same but are subjected to different influences.
- 9. A method according to any of claims 1-7, wherein the cells comprise multiple groups of cells that are qualitatively different but are subjected to the same influence.
  - 10. A method according to any of claims 1-9, wherein the recording is performed by means of a detector capable of measuring total luminescence in a non-spatially resolved fashion, the recording comprising a time series of measurements of the total luminescence of the cells of one or several of the spatial limitations.
- 11. A method according to claim 10, wherein the signal is measured from individual spatial limitations one at a time, the recording being made in the individual spatial limitation by means of an apparatus to sequentially position each one of the limitations in the field of view of the detector, and repeating the positioning and measuring process until all of the spatial limitations have been measured.
- 20 12. A method according to claim 11, wherein the detector is a photomultiplier tube (PMT).
  - 13. A method according to any of claims 1-9, wherein more than one of the spatial limitations are measured simultaneously.
- 14. A method according to claim 13, wherein the multiple spatial limitations are measured simultaneously by means of a one- or two-dimensional array detector, whereby the multiple spatial limitations are imaged onto the array detector such that discrete subsets of the detecting units (pixels) in the array detector measure the signal from one and

only one of the multiple spatial limitations, the signal from any one spatial limitation being the combined signal from those pixels that receive the image from one of the spatial limitations.

- 15. A method according to claim 14, wherein the detector is a linear diode array.
- 5 16. A method according to claim 14, wherein the detector is a video camera.
  - 17. A method according to claim 14, wherein the detector is a charge transfer device.
  - 18. A method according to claim 17, wherein the charge transfer device is a charge-coupled device.
- 19. A method according to any of claims 1-18, wherein the luminophore must be illuminated in order to emit light.
  - 20. A method according to any of claims 13-18, wherein all of the multiple spatial limitations are simultaneously illuminated during the measurement operation.
  - 21. A method according to any of claims 10-18, wherein the individual spatial limitations are singly illuminated only during the time period in which they are being measured.
- 15 22. A method according to any of claims 10-18, wherein the illumination is provided by a laser which is scanned in a raster fashion over some or all of the spatial limitations being measured, the scanning taking place at a rate substantially faster than the measurement process such that the illumination appears to the measurement process to be continuous in time and spatially uniform over the region being measured.
- 23. A method according to any of claims 1-22, wherein the spatial limitations are spatial limitations arranged in one or more arrays on a common carrier.
  - 24. A method according to claim 23, wherein the spatial limitations are wells in a plate of microtiter type.
- 25. A method according to any of claims 1-22 wherein the spatial limitations are domainsdefined on a substrate on which the cells are present.

- 26. A method according to claim 25 wherein the domains are domains established by the presence of the cells on the substrate in a pattern defining the domains.
- 27. A method according to claim 25 wherein the domains are domains established by the spatial pattern of the influence as it is applied to or contacted with the cells.
- 28. A method according to any of claims 1-27, wherein the recording is performed at a series of points in time, in which the application of the influence occurs at some time after the first time point in the series of recordings, the recording being performed, e.g., with a predetermined time spacing of from 0.1 seconds to 1 hour, preferably from 1 to 60 seconds, more preferably from 1 to 30 seconds, in particular from 1 to 10 seconds, over a time span of from 1 second to 12 hours, such as from 10 seconds to 12 hours, e.g., from 10 seconds to one hour, such as from 60 seconds to 30 minutes or 20 minutes.
  - 29. A method according to claim 28, wherein the recording is made at two points in time, one point being before, and the other point being after the application of the influence.
- 30. A method according to any of claims 1-29, wherein the cells are fixed at a point in time after the application of the influence at which the response has been predetermined to be significant, and the recording is made at an arbitrary later time.
  - 31. A method according to any of claims 1-30, wherein the luminophore is a luminophore that is capable of being redistributed in a manner that is physiologically relevant to the degree of the influence.
  - 32. A method according to any of claims 1-30, wherein the luminophore is a luminophore which is capable of associating with a component which is capable of being redistributed in manner which is physiologically relevant to the degree of the influence.
- 33. A method according to any of claims 1-30, wherein the luminophore is a luminophore
   which is capable of being redistributed in a manner which is experimentally
   determined to be correlated to the degree of the influence.
  - 34. A method according to any of claims 1-30, wherein the luminophore is a luminophore

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which is capable of being redistributed, by modulation of the intracellular pathway, in substantially the same manner as the at least one component of the intracellular pathway.

- 35. A method according to any of claims 1-30, wherein the luminophore is a luminophore which is capable of being quenched upon spatial association with a component which is redistributed by modulation of the pathway, the quenching being measured as a decrease in the intensity of the luminescence.
- 36. A method according to any of claims 1-30, wherein the variation in spatially distributed light emitted by the luminophore is detected by a change in the resonance energy transfer between the luminophore and another luminescent entity capable of delivering energy to the luminophore, each of which has been selected or engineered to become part of, bound to or associated with particular components of the intracellular pathway, and one of which undergoes redistribution in response to the influence, thereby changing the amount of resonance energy transfer, the change in the resonance energy transfer being measured as a change in the intensity of emission from the luminophore.
  - 37. A method according to any of claims 1-35, wherein the intensity of the light being recorded is a function of the fluorescence lifetime, polarisation, wavelength shift, or other property which is modulated as a result of the underlying cellular response.
- 38. A method according to any of claims 1-37, wherein the light to be measured passes through a filter which selects the desired component of the light to be measured and rejects other components.
  - 39. A method according to any of claims 2-38, wherein the intracellular pathway is an intracellular signalling pathway.
- 40. A method according to any of claims 1-39, wherein the luminophore is a fluorophore.
  - 41. A method according to any of claims 1-40, wherein the luminophore is a polypeptide encoded by and expressed from a nucleotide sequence harboured in the cells.

- 42. A method according to any of claims 1-41 for detecting intracellular redistribution of a biologically active polypeptide affecting intracellular processes upon activation, the method comprising
- a) culturing one or more cells containing a nucleotide sequence coding for a hybrid
   polypeptide comprising a GFP which is N- or C-terminally tagged, optionally through
  a linker, to a biologically active polypeptide under conditions permitting expression of
  the nucleotide sequence,
  - b) modulating the activity of the biologically active polypeptide by incubating the cells with a substance having biological activity, and
- or variation with respect to the fluorescence, such result or variation being indicative of the redistribution of a biologically active polypeptide in said cells.
  - 43. A method according to claim 42, wherein the luminophore is a hybrid polypeptide comprising a fusion of at least a portion of each of two polypeptides one of which comprises a luminescent polypeptide and the other one of which comprises a biologically active polypeptide, as defined herein.
  - 44. A method according to claim 43, wherein the luminescent polypeptide is a GFP as defined herein.
- 45. A method according to claim 44, wherein the GFP is selected from the group consisting of green fluorescent proteins having the F64L mutation as defined herein.
  - 46. A method according to claim 45, wherein the GFP is a GFP variant selected from the group consisting of F64L-GFP, F64L-Y66H-GFP, F64L-S65T-GFP, and EGFP.
  - 47. A method according to claim 42, wherein the nucleotide sequence is a DNA sequence.
  - 48. A method according to claims 42-47, wherein the modulation is activation.
- 49. A method according to claims 42-47, wherein the modulation is deactivation.
  - 50. A method according to any of claims 1-49, wherein the cells are selected from the

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group consisting of fungal cells, such as yeast cells; invertebrate cells including insect cells; and vertebrate cells, such as mammalian cells.

- 51. A method according to claim 50, wherein the mechanically intact or permeabilised living cells are mammalian cells which, during the time period over which the influence is observed, are incubated at a temperature of 30°C or above, preferably at a temperature of from 32°C to 39°C, more preferably at a temperature of from 35°C to 38°C, and most preferably at a temperature of about 37°C.
- 52. A method according to any of claims 1-51, wherein the mechanically intact or permeabilised living cells are part of a matrix of identical or non-identical cells.
- 53. A method according to any of claims 41-52, wherein the nucleotide sequence has been introduced into the cells in the form of a nucleic acid construct coding for a fusion polypeptide comprising a biologically active polypeptide that is a component of an intracellular signalling pathway, or a part thereof, and a GFP.
- 54. A method according to claim 53, wherein the nucleic acid construct is a nucleic acid construct coding for a fusion polypeptide comprising a biologically active polypeptide that is a component of an intracellular signalling pathway, or a part thereof, and an F64L mutant of GFP.
  - 55. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 53 or 54, wherein the biologically active polypeptide is a protein kinase or a phosphatase.
  - 56. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 53 55, wherein the GFP is N- or C-terminally tagged, optionally via a peptide linker, to the biologically active polypeptide or part thereof.
- 57. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 53, 54 or 56, wherein the biologically active polypeptide is a transcription factor or a part thereof which changes cellular localisation upon activation.

- 58. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 53, 54 or 56, wherein the biologically active polypeptide is a protein, or a part thereof, which is associated with the cytoskeletal network and which changes cellular localisation upon activation.
- 5 59. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to any of claims 53-56, wherein the biologically active polypeptide is a protein kinase or a part thereof which changes cellular localisation upon activation.
- 60. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a serine/threonine protein kinase or a part thereof capable of changing intracellular localisation upon activation.
  - 61. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a tyrosine protein kinase or a part thereof capable of changing intracellular localisation upon activation.
  - 62. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a phospholipid-dependent serine/threonine protein kinase or a part thereof capable of changing intracellular localisation upon activation.
- 20 63. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a cAMP-dependent protein kinase or a part thereof capable of changing cellular localisation upon activation.
- 64. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 63 which codes for a PKAc-F64L-S65T-GFP fusion.
  - 65. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a cGMP-dependent protein kinase or a part thereof capable of changing cellular localisation upon

activation.

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66. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a calmodulin-dependent serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.

- 67. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a mitogen-activated serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.
- 68. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 67, which codes for an ERK1-F64L-S65T-GFP fusion.
  - 69. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 67, which codes for an EGFP-ERK1 fusion.
- 15 70. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 59, wherein the protein kinase is a cyclin-dependent serine/threonine protein kinase or a part thereof capable of changing cellular localisation upon activation.
- 71. A method according to claim 53 or 54, wherein the nucleic acid construct is a nucleic acid construct according to claim 55 or 56, wherein the biologically active polypeptide is a protein phosphatase or a part thereof capable of changing cellular localisation upon activation.
  - 72. A method according to claim 53 -71, wherein the nucleic acid construct is a nucleic acid construct which is a DNA construct.
- 73. A method according to claim 53 -72, wherein the nucleic acid construct is a nucleic acid construct according to any of claims 53-72 wherein the gene encoding GFP is derived from Aequorea victoria.

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- 74. A method according to claim 73, wherein the nucleic acid construct is a nucleic acid construct according to claim 73 in which the gene encoding GFP is the gene encoding EGFP as defined herein.
- 75. A method according to claim 73, wherein the nucleic acid construct is a nucleic acid construct according to claim 73 in which the gene encoding a GFP is a gene encoding a GFP variant selected from F64L-GFP, F64L-Y66H-GFP and F64L-S65T-GFP.
- 76. A method according to claims 72 and 74, wherein the nucleic acid construct is a DNA construct according to claims 72 and 74 or, where applicable, 75, which is a construct as identified by any of the DNA sequences shown in SEQ ID NO: 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, 78, 108, 110, 112, 114, 116, 118, 120, 122, 124, 126, 128, 130, 132, 134, 136, 138, 140, 142, 144, 146, 148, 150, and 152 or is a variant thereof capable of encoding the same fusion polypeptide or a fusion polypeptide which is biologically equivalent thereto, as defined herein.
- 77. A method comprising a cell containing a nucleic acid construct according to any of claims 53-76 and capable of expressing the sequence encoded by the construct.
- 78. A method comprising a cell according to claim 77, which is a eukaryotic cell.
- 79. A method comprising a cell according to claim 77, which is selected from the group consisting of fungal cells, such as yeast cells; invertebrate cells, including insect cells, and vertebrate cells, such as mammalian cells.
- 80. A method according to any of claims 1-79, as used in a screening program as defined herein.
  - 81. A method according claim 80, wherein the method is a screening program for the identification of a biologically active substance as defined herein that directly or indirectly affects an intracellular signalling pathway and is potentially useful as a medicament, wherein the result of the individual measurement of each substance being screened which indicates its potential biological activity is based on measurement of the redistribution of spatially resolved luminescence in living cells and which undergoes a change in distribution upon activation of an intracellular signalling

pathway.

- 82. A method according to claim 80, wherein the method is a screening program for the identification of a biologically toxic substance as defined herein that exerts its toxic effect by interfering with an intracellular signalling pathway, wherein the result of the individual measurement of each substance being screened which indicates its potential biologically toxic activity is based on measurement of the redistribution of said fluorescent probe in living cells and which undergoes a change in distribution upon activation of an intracellular signalling pathway.
- 83. A method according to any of claims 1-82 wherein a fluorescent probe is used in backtracking of signal transduction pathways as defined herein.
  - 84. A method according to any of claims 1-83, for treating a condition or disease related to the intracellular function of a protein kinase comprising administering to a patient suffering from said condition or disease an effective amount of a compound which has been discovered by any method.
- 15 85. A compound that modulates a component of an intracellular pathway as defined herein, as determined by any method according to any of claims 1-83.
  - 86. A medical composition comprising a therapeutic amount of a compound identified according to any method according to any of claims 1-83.
- medical treatment comprising obtaining a primary cells from said patient, transfecting the cells with at least one DNA sequence encoding a fluorescent probe according to any of the preceding claims, culturing the cells under conditions permitting the expression of said probes and exposing it to an array of medicaments suspected of being capable of alleviating said ailment, then comparing changes in fluorescence patterns or redistribution patterns of the fluorescent probes in the intact living cells to detect the cellular response to the specific medicaments (obtaining a cellular action profile), then selecting a medicament(s) based on desired activity and acceptable level of side effects and administering an effective amount of said medicament(s) to said

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patient.

88. A method according to any of claims 1-83 of identifying a drug target among the group of biologically active polypeptides that are components of intracellular signalling pathways.

## **ABSTRACT**

An improved method and tools for quantifying the effect of an influence on cellular response is described. In particular, an improved method is described for detecting intracellular translocation or redistribution of biologically active polypeptides. The invention also describes several ways of contacting the cells with a substance influencing a cellular response and extracting quantitative information relating to the response in a highly parallel fashion. The method may be used as a very efficient procedure for testing or discovering the influence of a substance on a physiological process using commercially available parallel, high volume assay techniques, for example in connection with screening for new drugs, testing of substances for toxicity, and identifying drug targets for known or novel drugs.

### SEQUENCE LISTING

- (1) GENERAL INFORMATION
- (i) APPLICANT: NovoNordisk, BioImage
- (ii) TITLE OF THE INVENTION: An Improved Method of Detecting Cellular Translocation of Biologically Active Polypeptides Using Fluorescense Imaging
- (iii) NUMBER OF SEQUENCES: 165
- (iv) CORRESPONDENCE ADDRESS:
  - (A) ADDRESSEE: NovoNordisk, BioImage
  - (B) STREET: Mørkhøjbygade 28
  - (C) CITY: Søborg
  - (D) STATE: DK
  - (E) COUNTRY: DENMARK
  - (F) ZIP: 2860
- (v) COMPUTER READABLE FORM:
  - (A) MEDIUM TYPE: Diskette
  - (B) COMPUTER: IBM Compatible
  - (C) OPERATING SYSTEM: DOS
- (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (viii) ATTORNEY/AGENT INFORMATION:
  - (A) NAME: , PV&P R
  - (B) REGISTRATION NUMBER:
  - (C) REFERENCE/DOCKET NUMBER:
  - (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 53 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
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- (2) INFORMATION FOR SEQ ID NO:2:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 53 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
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GTCATCTTCT CGAGTCTTAC TCCTGAGGTT TGTATAGTTC ATCCATGCCA TGT	53
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(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
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/ ' \ ADALTHYOD AND ADDATOR OF	
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(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
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(A) LENGTH: 30 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
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(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(5) 10102001. 11.001	
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(B) TYPE: nucleic acid	
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(D) TOPOLOGY: linear	

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TAGGATCCAT AGATCTGTAT CCTGG	25
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(vi) ORIGINAL SOURCE:  (A) ORGANISM: p85-top-C	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
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(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
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(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1896 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence(B) LOCATION: 1...1891

(D) OTHER INFORMATION:

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 			GAC Asp			_	_		_	9	6
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			CTG Leu							19	2
			CAG Gln 70							24	0
 			AAG Lys							28	88
			AAG Lys							33	36
			GAC Asp							38	34
			GAC Asp							43	32
			AAC Asn 150							4.8	30
 			TTC Phe					_	Ser	52	28

	AAC ACC CCC ATC GGC GAC Asn Thr Pro Ile Gly Asp 190	
	CTG AGC ACC CAG TCC GCC Leu Ser Thr Gln Ser Ala 205	•
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	ATG GAC GAG CTG TAC AAG Met Asp Glu Leu Tyr Lys 235	
J Ala Gln Ala Ser	AAT TCA ACC ATG GCG GCG Asn Ser Thr Met Ala Ala 250 255	a Ala
	CCC CGT AGA ACC GAG GGC Pro Arg Arg Thr Glu Gly 270	
	ATG GTG AAG GGG CAG CCC Met Val Lys Gly Gln Pro 285	
	CAG TAC ATC GGC GAG GGC Gln Tyr Ile Gly Glu Gly 300	
	CAC GTG CGC AAG ACT CGG His Val Arg Lys Thr Arg 315	
e Ser Pro Phe Glu	CAT CAG ACC TAC TGC CAC His Gln Thr Tyr Cys Gln 330 33	n Arg
	CGC TTC CGC CAT GAG AA' Arg Phe Arg His Glu Ass 350	
	TCC ACC CTG GAA GCC ATG Ser Thr Leu Glu Ala Me 365	
	GAG ACT GAC CTG TAC AAG Glu Thr Asp Leu Tyr Ly 380	
	CAT ATC TGC TAC TTC CT His Ile Cys Tyr Phe Le 395	
	CAC TCC GCC AAC GTG CT His Ser Ala Asn Val Le	

CGA GAT CTA AAG CCC TCC AAC CTG CTC AGC AAC ACC ACC TGC GAC CTT Arg Asp Leu Lys Pro Ser Asn Leu Leu Ser Asn Thr Thr Cys Asp Leu AAG ATT TGT GAT TTC GGC CTG GCC CGG ATT GCC GAT CCT GAG CAT GAC Lys Ile Cys Asp Phe Gly Leu Ala Arg Ile Ala Asp Pro Glu His Asp CAC ACC GGC TTC CTG ACG GAG TAT GTG GCT ACG CGC TGG TAC CGG GCC His Thr Gly Phe Leu Thr Glu Tyr Val Ala Thr Arg Trp Tyr Arg Ala CCA GAG ATC ATG CTG AAC TCC AAG GGC TAT ACC AAG TCC ATC GAC ATC Pro Glu Ile Met Leu Asn Ser Lys Gly Tyr Thr Lys Ser Ile Asp Ile TGG TCT GTG GGC TGC ATT CTG GCT GAG ATG CTC TCT AAC CGG CCC ATC Trp Ser Val Gly Cys Ile Leu Ala Glu Met Leu Ser Asn Arg Pro Ile TTC CCT GGC AAG CAC TAC CTG GAT CAG CTC AAC CAC ATT CTG GGC ATC Phe Pro Gly Lys His Tyr Leu Asp Gln Leu Asn His Ile Leu Gly Ile CTG GGC TCC CCA TCC CAG GAG GAC CTG AAT TGT ATC ATC AAC ATG AAG Leu Gly Ser Pro Ser Gln Glu Asp Leu Asn Cys Ile Ile Asn Met Lys GCC CGA AAC TAC CTA CAG TCT CTG CCC TCC AAG ACC AAG GTG GCT TGG Ala Arg Asn Tyr Leu Gln Ser Leu Pro Ser Lys Thr Lys Val Ala Trp GCC AAG CTT TTC CCC AAG TCA GAC TCC AAA GCC CTT GAC CTG CTG GAC Ala Lys Leu Phe Pro Lys Ser Asp Ser Lys Ala Leu Asp Leu Leu Asp CGG ATG TTA ACC TTT AAC CCC AAT AAA CGG ATC ACA GTG GAG GAA GCG Arg Met Leu Thr Phe Asn Pro Asn Lys Arg Ile Thr Val Glu Glu Ala CTG GCT CAC CCC TAC CTG GAG CAG TAC TAT GAC CCG ACG GAT GAG CCA Leu Ala His Pro Tyr Leu Glu Gln Tyr Tyr Asp Pro Thr Asp Glu Pro GTG GCC GAG GAG CCC TTC ACC TTC GCC ATG GAG CTG GAT GAC CTA CCT Val Ala Glu Glu Pro Phe Thr Phe Ala Met Glu Leu Asp Asp Leu Pro AAG GAG CGG CTG AAG GAG CTC ATC TTC CAG GAG ACA GCA CGC TTC CAG Lys Glu Arg Leu Lys Glu Leu Ile Phe Gln Glu Thr Ala Arg Phe Gln CCC GGA GTG CTG GAG GCC C CCTAG Pro Gly Val Leu Glu Ala Pro

#### (2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 631 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 30 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 45 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 85 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 125 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 140 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 175 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 190 180 185 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 220 210 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 230 235 Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Thr Met Ala Ala Ala 245 250 Ala Ala Gln Gly Gly Gly Gly Glu Pro Arg Arg Thr Glu Gly Val 265 260 Gly Pro Gly Val Pro Gly Glu Val Glu Met Val Lys Gly Gln Pro Phe 280 285 Asp Val Gly Pro Arg Tyr Thr Gln Leu Gln Tyr Ile Gly Glu Gly Ala 300 295 Tyr Gly Met Val Ser Ser Ala Tyr Asp His Val Arg Lys Thr Arg Val 305 310 315 Ala Ile Lys Lys Ile Ser Pro Phe Glu His Gln Thr Tyr Cys Gln Arg 330 Thr Leu Arg Glu Ile Gln Ile Leu Leu Arg Phe Arg His Glu Asn Val 350 345 340

Ile Gly Ile Arg Asp Ile Leu Arg Ala Ser Thr Leu Glu Ala Met Arg 360 Asp Val Tyr Ile Val Gln Asp Leu Met Glu Thr Asp Leu Tyr Lys Leu 375 Leu Lys Ser Gln Gln Leu Ser Asn Asp His Ile Cys Tyr Phe Leu Tyr 390 395 Gln Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala Asn Val Leu His 405 410 Arg Asp Leu Lys Pro Ser Asn Leu Leu Ser Asn Thr Thr Cys Asp Leu 425 420 430 Lys Ile Cys Asp Phe Gly Leu Ala Arg Ile Ala Asp Pro Glu His Asp 440 445 His Thr Gly Phe Leu Thr Glu Tyr Val Ala Thr Arg Trp Tyr Arg Ala 455 460 Pro Glu Ile Met Leu Asn Ser Lys Gly Tyr Thr Lys Ser Ile Asp Ile 470 475 Trp Ser Val Gly Cys Ile Leu Ala Glu Met Leu Ser Asn Arg Pro Ile 485 490 Phe Pro Gly Lys His Tyr Leu Asp Gln Leu Asn His Ile Leu Gly Ile 505 500 Leu Gly Ser Pro Ser Gln Glu Asp Leu Asn Cys Ile Ile Asn Met Lys 515 520 525 Ala Arg Asn Tyr Leu Gln Ser Leu Pro Ser Lys Thr Lys Val Ala Trp 535 540 Ala Lys Leu Phe Pro Lys Ser Asp Ser Lys Ala Leu Asp Leu Leu Asp 550 555 Arg Met Leu Thr Phe Asn Pro Asn Lys Arg Ile Thr Val Glu Glu Ala 565 570 Leu Ala His Pro Tyr Leu Glu Gln Tyr Tyr Asp Pro Thr Asp Glu Pro 580 585 590 Val Ala Glu Glu Pro Phe Thr Phe Ala Met Glu Leu Asp Asp Leu Pro 600 605 Lys Glu Arg Leu Lys Glu Leu Ile Phe Gln Glu Thr Ala Arg Phe Gln 615 Pro Gly Val Leu Glu Ala Pro

### (2) INFORMATION FOR SEQ ID NO:40:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1818 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...1815
  - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu

1 5 10 15

					GAC Asp											96
					GCC Ala											144
					CTG Leu											192
					CAG Gln 70											240
					AAG Lys											288
					AAG Lys											336
					GAC Asp											384
					GAC Asp											432
					AAC Asn 150											480
															Ser	528
														Asp	GGC	576
CCC Pro	GTG Val	CTG Leu 195	Leu	CCC Pro	GAC Asp	AAC Asn	CAC His 200	Tyr	CTG Leu	AGC Ser	ACC Thr	Gln 205	Ser	GCC	CTG Leu	624
AGC Ser	AAA Lys 210	Asp	CCC	AAC Asn	GAG Glu	AAG Lys 215	Arg	GAT Asp	CAC His	ATG Met	GTC Val 220	Leu	CTG Leu	GAC Glu	TTC Phe	672
	Thr					Thr					Glu				TCC Ser 240	720
GGA Gly	CTC Leu	AGA Arg	TCT	CGA Arg	GTA Val	ACC Thr	ATG Met	GCG Ala	GCG Ala	GCG	GCG Ala	GCC Ala	GCG Ala	GGC Gly	CCG Pro	768

245 250 GAG ATG GTC CGC GGG CAG GTG TTC GAC GTG GGG CCG CGC TAC ACT AAT Glu Met Val Arg Gly Gln Val Phe Asp Val Gly Pro Arg Tyr Thr Asn 265 CTC TCG TAC ATC GGA GAA GGC GCC TAC GGC ATG GTT TGT TCT GCT TAT Leu Ser Tyr Ile Gly Glu Gly Ala Tyr Gly Met Val Cys Ser Ala Tyr 280 GAT AAT CTC AAC AAA GTT CGA GTT GCT ATC AAG AAA ATC AGT CCT TTT 912 Asp Asn Leu Asn Lys Val Arg Val Ala Ile Lys Lys Ile Ser Pro Phe 295 300 GAG CAC CAG ACC TAC TGT CAG AGA ACC CTG AGA GAG ATA AAA ATC CTA 960 Glu His Gln Thr Tyr Cys Gln Arg Thr Leu Arg Glu Ile Lys Ile Leu 315 310 CTG CGC TTC AGA CAT GAG AAC ATC ATC GGC ATC AAT GAC ATC ATC CGG 1008 Leu Arg Phe Arg His Glu Asn Ile Ile Gly Ile Asn Asp Ile Ile Arg 325 GCA CCA ACC ATT GAG CAG ATG AAA GAT GTA TAT ATA GTA CAG GAC CTC 1056 Ala Pro Thr Ile Glu Gln Met Lys Asp Val Tyr Ile Val Gln Asp Leu 340 345 ATG GAG ACA GAT CTT TAC AAG CTC TTG AAG ACA CAG CAC CTC AGC AAT 1104 Met Glu Thr Asp Leu Tyr Lys Leu Leu Lys Thr Gln His Leu Ser Asn 355 360 365 GAT CAT ATC TGC TAT TTT CTT TAT CAG ATC CTG AGA GGA TTA AAG TAT 1152 Asp His Ile Cys Tyr Phe Leu Tyr Gln Ile Leu Arg Gly Leu Lys Tyr 370 375 ATA CAT TCA GCT AAT GTT CTG CAC CGT GAC CTC AAG CCT TCC AAC CTC 1200 Ile His Ser Ala Asn Val Leu His Arg Asp Leu Lys Pro Ser Asn Leu 395 385 390 CTG CTG AAC ACC ACT TGT GAT CTC AAG ATC TGT GAC TTT GGC CTT GCC Leu Leu Asn Thr Thr Cys Asp Leu Lys Ile Cys Asp Phe Gly Leu Ala 405 410 CGT GTT GCA GAT CCA GAC CAT GAT CAT ACA GGG TTC TTG ACA GAG TAT 1296 Arg Val Ala Asp Pro Asp His Asp His Thr Gly Phe Leu Thr Glu Tyr 420 425 GTA GCC ACG CGT TGG TAC AGA GCT CCA GAA ATT ATG TTG AAT TCC AAG Val Ala Thr Arg Trp Tyr Arg Ala Pro Glu Ile Met Leu Asn Ser Lys 435 440 GGT TAT ACC AAG TCC ATT GAT ATT TGG TCT GTG GGC TGC ATC CTG GCA Gly Tyr Thr Lys Ser Ile Asp Ile Trp Ser Val Gly Cys Ile Leu Ala GAG ATG CTA TCC AAC AGG CCT ATC TTC CCA GGA AAG CAT TAC CTT GAC 1440

Glu Met Leu Ser Asn Arg Pro Ile Phe Pro Gly Lys His Tyr Leu Asp

475

480

470

CTG Leu								1488
 AAT Asn								1536
CAC His								1584
 AAA Lys 530								1632
AGG Arg								1680
 тат Тух								1728
 ATG Met								1776
 GAA Glu						TAA		1818

### (2) INFORMATION FOR SEQ ID NO:41:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 605 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 1 5 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 30 20 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 45 35 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu

```
Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu
           100
                              105
Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly
                         120
                                            125
       115
Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr
                     135
                                         140
Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn
                 150
                                     155
Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser
              165
                                 170
Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly
          180
                             185
                                                190
Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu
                         200
      195
                                            205
Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe
                      215
                                         220
Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser
                   230
                                      235
Gly Leu Arg Ser Arg Val Thr Met Ala Ala Ala Ala Ala Ala Gly Pro
              245
                                 250
Glu Met Val Arg Gly Gln Val Phe Asp Val Gly Pro Arg Tyr Thr Asn
           260
                              265
                                                 270
Leu Ser Tyr Ile Gly Glu Gly Ala Tyr Gly Met Val Cys Ser Ala Tyr
                          280
Asp Asn Leu Asn Lys Val Arg Val Ala Ile Lys Lys Ile Ser Pro Phe
                     295
                                         300
Glu His Gln Thr Tyr Cys Gln Arg Thr Leu Arg Glu Ile Lys Ile Leu
                 310
                                     315
Leu Arg Phe Arg His Glu Asn Ile Ile Gly Ile Asn Asp Ile Ile Arg
              325
                                  330
Ala Pro Thr Ile Glu Gln Met Lys Asp Val Tyr Ile Val Gln Asp Leu
           340
                              345
                                                 350
Met Glu Thr Asp Leu Tyr Lys Leu Leu Lys Thr Gln His Leu Ser Asn
                         360
                                             365
      355
Asp His Ile Cys Tyr Phe Leu Tyr Gln Ile Leu Arg Gly Leu Lys Tyr
                     375
Ile His Ser Ala Asn Val Leu His Arg Asp Leu Lys Pro Ser Asn Leu
         390
                                     395
Leu Leu Asn Thr Thr Cys Asp Leu Lys Ile Cys Asp Phe Gly Leu Ala
             405
                                 410
Arg Val Ala Asp Pro Asp His Asp His Thr Gly Phe Leu Thr Glu Tyr
                              425
Val Ala Thr Arg Trp Tyr Arg Ala Pro Glu Ile Met Leu Asn Ser Lys
                          440
                                             445
        435
Gly Tyr Thr Lys Ser Ile Asp Ile Trp Ser Val Gly Cys Ile Leu Ala
                    455
                                          460
Glu Met Leu Ser Asn Arg Pro Ile Phe Pro Gly Lys His Tyr Leu Asp
                                      475
Gln Leu Asn His Ile Leu Gly Ile Leu Gly Ser Pro Ser Gln Glu Asp
              485
                                  490
Leu Asn Cys Ile Ile Asn Leu Lys Ala Arg Asn Tyr Leu Leu Ser Leu
                             505
          500
Pro His Lys Asn Lys Val Pro Trp Asn Arg Leu Phe Pro Asn Ala Asp
                          520
                                              525
Ser Lys Ala Leu Asp Leu Leu Asp Lys Met Leu Thr Phe Asn Pro His
             535
                                         540
Lys Arg Ile Glu Val Glu Gln Ala Leu Ala His Pro Tyr Leu Glu Gln
                 550
                                       555
```

### (2) INFORMATION FOR SEQ ID NO:42:

### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2529 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

### (ii) MOLECULE TYPE: cDNA

- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2526
  - (D) OTHER INFORMATION:

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

		GAG Glu						48
		GAC Asp						96
		GCC Ala						144
		CTG Leu						192
 	 	CAG Gln 70						240
		AAG Lys						288
		AAG Lys						336
		GAC Asp						384
		GAC Asp						432

AAC TAC AAC AGC CAC AAC GTC TAT ATC ATG GCC GAC AAG CAG AAG AAC Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn GGC ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser GTG CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly CCC GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG GAG TTC Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe GTG ACC GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TCC Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser GGA CTC AGA TCT CGA GCT CAA GCT TCG AAT TCG TCA ATG GAG CTG GAA Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Ser Met Glu Leu Glu AAC ATC GTG GCC AAC ACG GTC TTG CTG AAA GCC AGG GAA GGG GGC GGA Asn Ile Val Ala Asn Thr Val Leu Leu Lys Ala Arg Glu Gly Gly GGA AAG CGC AAA GGG AAA AGC AAG AAG TGG AAA GAA ATC CTG AAG TTC Gly Lys Arg Lys Gly Lys Ser Lys Lys Trp Lys Glu Ile Leu Lys Phe CCT CAC ATT AGC CAG TGT GAA GAC CTC CGA AGG ACC ATA GAC AGA GAT Pro His Ile Ser Gln Cys Glu Asp Leu Arg Arg Thr Ile Asp Arg Asp TAC TGC AGT TTA TGT GAC AAG CAG CCA ATC GGG AGG CTG CTT TTC CGG Tyr Cys Ser Leu Cys Asp Lys Gln Pro Ile Gly Arg Leu Leu Phe Arg CAG TTT TGT GAA ACC AGG CCT GGG CTG GAG TGT TAC ATT CAG TTC CTG Gln Phe Cys Glu Thr Arg Pro Gly Leu Glu Cys Tyr Ile Gln Phe Leu GAC TCC GTG GCA GAA TAT GAA GTT ACT CCA GAT GAA AAA CTG GGA GAG Asp Ser Val Ala Glu Tyr Glu Val Thr Pro Asp Glu Lys Leu Gly Glu AAA GGG AAG GAA ATT ATG ACC AAG TAC CTC ACC CCA AAG TCC CCT GTT Lys Gly Lys Glu Ile Met Thr Lys Tyr Leu Thr Pro Lys Ser Pro Val

			GGC Gly						1152
			TGC Cys 390						1200
			AGG Arg	_					1248
			TTT Phe						1296
		_	TTC Phe				_		1344
			GCC Ala						1392
			GAG Glu 470						1440
			GAG Glu						1488
			GCC Ala						1536
			ATG Met						1584
			GGC Gly						1632
			GGC Gly 550						1680
			CCT Pro						1728
			CTG Leu						1776
			GTG Val						1824

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595 600 605 CTG AAC AAC CAG AGG TAC GGC CTG AGC CCC GAC TAC TGG GGC CTT GGC 1872 Leu Asn Asn Gln Arg Tyr Gly Leu Ser Pro Asp Tyr Trp Gly Leu Gly 615 620 TGC CTC ATC TAT GAG ATG ATC GAG GGC CAG TCG CCG TTC CGC GGC CGT 1920 Cys Leu Ile Tyr Glu Met Ile Glu Gly Gln Ser Pro Phe Arg Gly Arg 630 635 AAG GAG AAG GTG AAG CGG GAG GAG GTG GAC CGC CGG GTC CTG GAG ACG 1968 Lys Glu Lys Val Lys Arg Glu Glu Val Asp Arg Arg Val Leu Glu Thr 645 650 GAG GAG GTG TAC TCC CAC AAG TTC TCC GAG GAG GCC AAG TCC ATC TGC 2016 Glu Glu Val Tyr Ser His Lys Phe Ser Glu Glu Ala Lys Ser Ile Cys 660 665 AAG ATG CTG CTC ACG AAA GAT GCG AAG CAG AGG CTG GGC TGC CAG GAG 2064 Lys Met Leu Leu Thr Lys Asp Ala Lys Gln Arg Leu Gly Cys Gln Glu 680 GAG GGG GCT GCA GAG GTC AAG AGA CAC CCC TTC TTC AGG AAC ATG AAC 2112 Glu Gly Ala Ala Glu Val Lys Arg His Pro Phe Phe Arg Asn Met Asn 690 695 TTC AAG CGC TTA GAA GCC GGG ATG TTG GAC CCT CCC TTC GTT CCA GAC 2160 Phe Lys Arg Leu Glu Ala Gly Met Leu Asp Pro Pro Phe Val Pro Asp 705 710 715 CCC CGC GCT GTG TAC TGT AAG GAC GTG CTG GAC ATC GAG CAG TTC TCC 2208 Pro Arg Ala Val Tyr Cys Lys Asp Val Leu Asp Ile Glu Gln Phe Ser 725 ACT GTG AAG GGC GTC AAT CTG GAC CAC ACA GAC GAC GAC TTC TAC TCC 2256 Thr Val Lys Gly Val Asn Leu Asp His Thr Asp Asp Asp Phe Tyr Ser 740 745 AAG TTC TCC ACG GGC TCT GTG TCC ATC CCA TGG CAA AAC GAG ATG ATA 2304 Lys Phe Ser Thr Gly Ser Val Ser Ile Pro Trp Gln Asn Glu Met Ile 755 760 765 GAA ACA GAA TGC TTT AAG GAG CTG AAC GTG TTT GGA CCT AAT GGT ACC 2352 Glu Thr Glu Cys Phe Lys Glu Leu Asn Val Phe Gly Pro Asn Gly Thr CTC CCG CCA GAT CTG AAC AGA AAC CAC CCT CCG GAA CCG CCC AAG AAA 2400 Leu Pro Pro Asp Leu Asn Arg Asn His Pro Pro Glu Pro Pro Lys Lys GGG CTG CTC CAG AGA CTC TTC AAG CGG CAG CAT CAG AAC AAT TCC AAG 2448 Gly Leu Leu Gln Arg Leu Phe Lys Arg Gln His Gln Asn Asn Ser Lys 805 810 AGT TCG CCC AGC TCC AAG ACC AGT TTT AAC CAC CAC ATA AAC TCA AAC 2496 Ser Ser Pro Ser Ser Lys Thr Ser Phe Asn His His Ile Asn Ser Asn

CAT GTC AGC TCG AAC TCC ACC GGA AGC AGC TAG 2529
His Val Ser Ser Asn Ser Thr Gly Ser Ser

#### (2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 842 amino acids
  - (B) TYPE: amino acid

835

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 30 20 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 45 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 60 55 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 70 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 95 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 125 120 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 140 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 175 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 190 180 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 230 235 Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Ser Met Glu Leu Glu 245 250 Asn Ile Val Ala Asn Thr Val Leu Leu Lys Ala Arg Glu Gly Gly 260 270 265 Gly Lys Arg Lys Gly Lys Ser Lys Lys Trp Lys Glu Ile Leu Lys Phe 280 285 Pro His Ile Ser Gln Cys Glu Asp Leu Arg Arg Thr Ile Asp Arg Asp 300 295 Tyr Cys Ser Leu Cys Asp Lys Gln Pro Ile Gly Arg Leu Leu Phe Arg 310 315

Gln Phe Cys Glu Thr Arg Pro Gly Leu Glu Cys Tyr Ile Gln Phe Leu 325 330 Asp Ser Val Ala Glu Tyr Glu Val Thr Pro Asp Glu Lys Leu Gly Glu 345 Lys Gly Lys Glu Ile Met Thr Lys Tyr Leu Thr Pro Lys Ser Pro Val 360 365 Phe Ile Ala Gln Val Gly Gln Asp Leu Val Ser Gln Thr Glu Glu Lys 375 380 Leu Leu Gln Lys Pro Cys Lys Glu Leu Phe Ser Ala Cys Ala Gln Ser 390 395 Val His Glu Tyr Leu Arg Gly Glu Pro Phe His Glu Tyr Leu Asp Ser 405 410 Met Phe Phe Asp Arg Phe Leu Gln Trp Lys Trp Leu Glu Arg Gln Pro 425 420 Val Thr Lys Asn Thr Phe Arg Gln Tyr Arg Val Leu Gly Lys Gly Gly 435 440 445 Phe Gly Glu Val Cys Ala Cys Gln Val Arg Ala Thr Gly Lys Met Tyr 455 460 Ala Cys Lys Arg Leu Glu Lys Lys Arg Ile Lys Lys Arg Lys Gly Glu 470 475 Ser Met Ala Leu Asn Glu Lys Gln Ile Leu Glu Lys Val Asn Ser Gln 485 490 Phe Val Val Asn Leu Ala Tyr Ala Tyr Glu Thr Lys Asp Ala Leu Cys 500 505 510 Leu Val Leu Thr Ile Met Asn Gly Gly Asp Leu Lys Phe His Ile Tyr 520 525 515 Asn Met Gly Asn Pro Gly Phe Glu Glu Glu Arg Ala Leu Phe Tyr Ala 535 540 Ala Glu Ile Leu Cys Gly Leu Glu Asp Leu His Arg Glu Asn Thr Val 550 555 Tyr Arg Asp Leu Lys Pro Glu Asn Ile Leu Leu Asp Asp Tyr Gly His 565 570 Ile Arg Ile Ser Asp Leu Gly Leu Ala Val Lys Ile Pro Glu Gly Asp 585 Leu Ile Arg Gly Arg Val Gly Thr Val Gly Tyr Met Ala Pro Glu Val 600 605 Leu Asn Asn Gln Arg Tyr Gly Leu Ser Pro Asp Tyr Trp Gly Leu Gly 615 620 Cys Leu Ile Tyr Glu Met Ile Glu Gly Gln Ser Pro Phe Arg Gly Arg 630 635 Lys Glu Lys Val Lys Arg Glu Glu Val Asp Arg Arg Val Leu Glu Thr 645 650 Glu Glu Val Tyr Ser His Lys Phe Ser Glu Glu Ala Lys Ser Ile Cys 660 665 Lys Met Leu Leu Thr Lys Asp Ala Lys Gln Arg Leu Gly Cys Gln Glu - 680 Glu Gly Ala Ala Glu Val Lys Arg His Pro Phe Phe Arg Asn Met Asn 700 695 Phe Lys Arg Leu Glu Ala Gly Met Leu Asp Pro Pro Phe Val Pro Asp 710 715 Pro Arg Ala Val Tyr Cys Lys Asp Val Leu Asp Ile Glu Gln Phe Ser 730 Thr Val Lys Gly Val Asn Leu Asp His Thr Asp Asp Asp Phe Tyr Ser 740 745 Lys Phe Ser Thr Gly Ser Val Ser Ile Pro Trp Gln Asn Glu Met Ile 760 765 Glu Thr Glu Cys Phe Lys Glu Leu Asn Val Phe Gly Pro Asn Gly Thr 

### (2) INFORMATION FOR SEQ ID NO:44:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1902 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...1899
  - (D) OTHER INFORMATION:

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

_			GAG Glu					_	48
			GAC Asp						96
			GCC Ala						144
	 	 	CTG Leu	 	 	 	 		 192
			CAG Gln 70						240
			AAG Lys						288
			AAG Lys						336
			GAC Asp						384

		GAG Glu								432
		CAC His					_			480
		AAC Asn 165								528
		GAC Asp								576
		CCC Pro								624
		AAC Asn				_		_		672
		GGG Gly								720
		CGA Arg 245								768
		AGT Ser								816
									ATA Ile	864
		ТАТ Туг								912
		CCA Pro								960
									GGC Gly	1008
	 		_					Gln	GAT Asp	1056
_									ATT	1104

355 360 365

					TCC Ser			_		1152
 					GCT Ala					1200
			_		TCT Ser 410		_		_	1248
					GGA Gly					1296
					GCA Ala	_	_	 _		1344
					TGG Trp					1392
					TTT Phe					1440
					CTT Leu 490				GAA Glu	1488
									AGA Arg	1536
									Leu	1584
									AGG Arg	1632
Leu									Ser 560	1680
 	 								CCT Pro	1728
								Leu	GAT Asp	1776

GAA AGG GAA CAC ACA ATA GAA GAG TGG AAA GAA TTG ATA TAT AAG GAA
Glu Arg Glu His Thr Ile Glu Glu Trp Lys Glu Leu Ile Tyr Lys Glu
595 600 605

GTT ATG GAC TTG GAG GAG AGA ACC AAG AAT GGA GTT ATA CGG GGG CAG

Val Met Asp Leu Glu Glu Arg Thr Lys Asn Gly Val Ile Arg Gly Gln

610 620

CCC TCT CCT TTA GCA CAG GTG CAG CAG TGA 1902
Pro Ser Pro Leu Ala Gln Val Gln Gln
625 630

### (2) INFORMATION FOR SEQ ID NO:45:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 633 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
  (v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 70 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 125 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 · 200 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 225 230 235 240 Gly Leu Arg Ser Arg Ala Arg Ala Ile Met Ser Arg Ser Lys Arg Asp 245 250

Asn Asn Phe Tyr Ser Val Glu Ile Gly Asp Ser Thr Phe Thr Val Leu 265 Lys Arg Tyr Gln Asn Leu Lys Pro Ile Gly Ser Gly Ala Gln Gly Ile 275 280 Val Cys Ala Ala Tyr Asp Ala Ile Leu Glu Arg Asn Val Ala Ile Lys 290 295 300 Lys Leu Ser Arg Pro Phe Gln Asn Gln Thr His Ala Lys Arg Ala Tyr 305 310 315 Arg Glu Leu Val Leu Met Lys Cys Val Asn His Lys Asn Ile Ile Gly 325 330 Leu Leu Asn Val Phe Thr Pro Gln Lys Ser Leu Glu Glu Phe Gln Asp 340 345 350 Val Tyr Ile Val Met Glu Leu Met Asp Ala Asn Leu Cys Gln Val Ile 355 360 365 Gln Met Glu Leu Asp His Glu Arg Met Ser Tyr Leu Leu Tyr Gln Met 380 370 375 Leu Cys Gly Ile Lys His Leu His Ser Ala Gly Ile Ile His Arg Asp 395 385 390 Leu Lys Pro Ser Asn Ile Val Val Lys Ser Asp Cys Thr Leu Lys Ile 410 405 Leu Asp Phe Gly Leu Ala Arg Thr Ala Gly Thr Ser Phe Met Met Thr 420 425 430 Pro Tyr Val Val Thr Arg Tyr Tyr Arg Ala Pro Glu Val Ile Leu Gly 440 445 Met Gly Tyr Lys Glu Asn Val Asp Leu Trp Ser Val Gly Cys Ile Met 460 455 Gly Glu Met Val Cys His Lys Ile Leu Phe Pro Gly Arg Asp Tyr Ile 465 470 475 480 Asp Gln Trp Asn Lys Val Ile Glu Gln Leu Gly Thr Pro Cys Pro Glu 485 490 Phe Met Lys Lys Leu Gln Pro Thr Val Arg Thr Tyr Val Glu Asn Arg 505 510 Pro Lys Tyr Ala Gly Tyr Ser Phe Glu Lys Leu Phe Pro Asp Val Leu 525 515 520 Phe Pro Ala Asp Ser Glu His Asn Lys Leu Lys Ala Ser Gln Ala Arg 530 535 540 Asp Leu Leu Ser Lys Met Leu Val Ile Asp Ala Ser Lys Arg Ile Ser 550 555 Val Asp Glu Ala Leu Gln His Pro Tyr Ile Asn Val Trp Tyr Asp Pro 565 570 575 Ser Glu Ala Glu Ala Pro Pro Pro Lys Ile Pro Asp Lys Gln Leu Asp 580 585 Glu Arg Glu His Thr Ile Glu Glu Trp Lys Glu Leu Ile Tyr Lys Glu 595 600 605 Val Met Asp Leu Glu Glu Arg Thr Lys Asn Gly Val Ile Arg Gly Gln 610 615 Pro Ser Pro Leu Ala Gln Val Gln Gln 630

### (2) INFORMATION FOR SEQ ID NO:46:

### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1824 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

### (ix) FEATURE:

- (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...1821
- (D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

Me	et '	GTG Val	AGC Ser	AA	s Gl	C GAG	G GA	G CTY u Lev	G TTV	e Thi	C GGG	G GT y Va	G GTO	G CCC	C ATC	CTG	48
GI		GAG	CTC	GAG	5 C GG(	C GAG	C GT	A AAC	c GG(	10 C CAC	C AAC	G TT	C AGO	GTC	15 TCC	: GGC	96
Va	11 (	Glu	Leu	Ası 20	o Gly	y Ası	Va:	l Asr	1 Gly 25	/ His	s Lys	s Phe	e Ser	Val	Ser	Gly	30
GA G1	G (	GGC Gly	GAG Glu 35	G17	C GAT	r GCC	ACC Thi	TAC Tyr 40	Gly	AAC Lys	CTC	G ACC	C CTG Leu 45	AAC Lys	TTC Phe	ATC : Ile	144
TG Cy	s T	ACC Thr 50	ACC Thr	GGC	AAC Lys	CTC Leu	CCC Pro 55	GTG Val	Pro	TGC Tri	CCC Pro	ACC Thr	CTC Leu	GTG Val	ACC Thr	ACC Thr	192
CT Le 65	G A	ACC	TAC Tyr	GGC Gly	GTG Val	Gln 70	TGC Cys	TTC Phe	AGC	CGC Arg	ТАС Тут 75	Pro	GAC Asp	CAC	ATG Met	AAG Lys 80	240
CA( Gl	G C	AC Iis	GAC Asp	TTC Phe	Phe 85	AAG Lys	TCC Ser	GCC Ala	ATG Met	CCC Pro 90	GAA Glu	GGC Gly	TAC Tyr	GTC Val	CAG Gln 95	GAG Glu	288
CG( Arg	A S	CC hr	ATC Ile	TTC Phe 100	TTC Phe	AAG Lys	GAC Asp	GAC Asp	GGC Gly 105	AAC Asn	TAC Tyr	AAG Lys	ACC Thr	CGC Arg 110	GCC Ala	GAG Glu	336
GT( Va]	G A	ys	TTC Phe 115	GAG Glu	GGC Gly	GAC Asp	ACC Thr	CTG Leu 120	GTG Val	AAC Asn	CGC Arg	ATC Ile	GAG Glu 125	CTG Leu	AAG Lys	GGC Gly	384
ATC Ile	• A	AC sp 30	TTC Phe	AAG Lys	GAG Glu	GAC Asp	GGC Gly 135	AAC Asn	ATC Ile	CTG Leu	GGG Gly	CAC His 140	AAG Lys	CTG Leu	GAG Glu	TAC Tyr	432
AAC Asn 145	T	AC I	AAC Asn	AGC Ser	CAC His	AAC Asn 150	GTC Val	TAT Tyr	ATC Ile	ATG Met	GCC Ala 155	GAC Asp	AAG Lys	CAG Gln	AAG Lys	AAC Asn 160	480
GGC Gly	A'	rc z le I	AAG Lys	GTG Val	AAC Asn 165	TTC Phe	AAG Lys	ATC Ile	CGC Arg	CAC His 170	AAC Asn	ATC Ile	GAG Glu	GAC Asp	GGC Gly 175	AGC Ser	528
GTG Val	CZ G]	AG (	Leu .	GCC Ala 180	GAC Asp	CAC His	TAC Tyr	Gln	CAG Gln 185	AAC Asn	ACC Thr	CCC Pro	ATC Ile	GGC Gly 190	GAC Asp	GGC Gly	576
CCC Pro	G1 Va	G C	CTG ( Leu 1	CTG Leu	CCC Pro	GAC Asp	AAC Asn	CAC His	TAC Tyr	CTG Leu	AGC Ser	ACC Thr	CAG Gln	TCC Ser	GCC Ala	CTG Leu	624

AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe GTG ACC GCC GCG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TCC Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser GGA CTC AGA TCT CGA GGG AAA ATG TCT CAG GAG AGG CCC ACG TTC TAC Gly Leu Arg Ser Arg Gly Lys Met Ser Gln Glu Arg Pro Thr Phe Tyr CGG CAG GAG CTG AAC AAG ACA ATC TGG GAG GTG CCC GAG CGT TAC CAG Arg Gln Glu Leu Asn Lys Thr Ile Trp Glu Val Pro Glu Arg Tyr Gln AAC CTG TCT CCA GTG GGC TCT GGC GCC TAT GGC TCT GTG TGT GCT GCT Asn Leu Ser Pro Val Gly Ser Gly Ala Tyr Gly Ser Val Cys Ala Ala TTT GAC ACA AAA ACG GGG TTA CGT GTG GCA GTG AAG AAG CTC TCC AGA Phe Asp Thr Lys Thr Gly Leu Arg Val Ala Val Lys Lys Leu Ser Arg CCA TTT CAG TCC ATC ATT CAT GCG AAA AGA ACC TAC AGA GAA CTG CGG Pro Phe Gln Ser Ile Ile His Ala Lys Arg Thr Tyr Arg Glu Leu Arg TTA CTT AAA CAT ATG AAA CAT GAA AAT GTG ATT GGT CTG TTG GAC GTT Leu Leu Lys His Met Lys His Glu Asn Val Ile Gly Leu Leu Asp Val TTT ACA CCT GCA AGG TCT CTG GAG GAA TTC AAT GAT GTG TAT CTG GTG Phe Thr Pro Ala Arg Ser Leu Glu Glu Phe Asn Asp Val Tyr Leu Val ACC CAT CTC ATG GGG GCA GAT CTG AAC AAC ATT GTG AAA TGT CAG AAG Thr His Leu Met Gly Ala Asp Leu Asn Asn Ile Val Lys Cys Gln Lys CTT ACA GAT GAC CAT GTT CAG TTC CTT ATC TAC CAA ATT CTC CGA GGT Leu Thr Asp Asp His Val Gln Phe Leu Ile Tyr Gln Ile Leu Arg Gly CTA AAG TAT ATA CAT TCA GCT GAC ATA ATT CAC AGG GAC CTA AAA CCT Leu Lys Tyr Ile His Ser Ala Asp Ile Ile His Arg Asp Leu Lys Pro AGT AAT CTA GCT GTG AAT GAA GAC TGT GAG CTG AAG ATT CTG GAT TTT Ser Asn Leu Ala Val Asn Glu Asp Cys Glu Leu Lys Ile Leu Asp Phe GGA CTG GCT CGG CAC ACA GAT GAT GAA ATG ACA GGC TAC GTG GCC ACT 

Gly Leu Ala Arg His Thr Asp Asp Glu Met Thr Gly Tyr Val Ala Thr 

					CTG Leu						1344
					TGC Cys						1392
					GAC Asp						1440
					GGG Gly 490						1488
					ATT Ile				_		1536
					ATT Ile					GCT Ala	1584
					TTG Leu		Asp			ATT	1632
										GAT Asp 560	1680
										AGC Ser	1728
					Ser				Glu	GTC Val	1776
	Val			Asp	CAA Gln			Glu		TGA	1824

### (2) INFORMATION FOR SEQ ID NO:47:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 607 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- . (xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu · 105 110 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 125 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 175 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 190 180 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 200 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 230 235 Gly Leu Arg Ser Arg Gly Lys Met Ser Gln Glu Arg Pro Thr Phe Tyr 250 . 245 Arg Gln Glu Leu Asn Lys Thr Ile Trp Glu Val Pro Glu Arg Tyr Gln 265 Asn Leu Ser Pro Val Gly Ser Gly Ala Tyr Gly Ser Val Cys Ala Ala 285 275 280 Phe Asp Thr Lys Thr Gly Leu Arg Val Ala Val Lys Lys Leu Ser Arg 290 295 300 Pro Phe Gln Ser Ile Ile His Ala Lys Arg Thr Tyr Arg Glu Leu Arg 310 315 Leu Leu Lys His Met Lys His Glu Asn Val Ile Gly Leu Leu Asp Val 330 325 Phe Thr Pro Ala Arg Ser Leu Glu Glu Phe Asn Asp Val Tyr Leu Val 340 345 Thr His Leu Met Gly Ala Asp Leu Asn Asn Ile Val Lys Cys Gln Lys . 360 365 Leu Thr Asp Asp His Val Gln Phe Leu Ile Tyr Gln Ile Leu Arg Gly 375 380 Leu Lys Tyr Ile His Ser Ala Asp Ile Ile His Arg Asp Leu Lys Pro 390 395 Ser Asn Leu Ala Val Asn Glu Asp Cys Glu Leu Lys Ile Leu Asp Phe 410 Gly Leu Ala Arg His Thr Asp Asp Glu Met Thr Gly Tyr Val Ala Thr 425 430 Arg Trp Tyr Arg Ala Pro Glu Ile Met Leu Asn Trp Met His Tyr Asn 445 435 440 Gln Thr Val Asp Ile Trp Ser Val Gly Cys Ile Met Ala Glu Leu Leu 455 460

Thr 465	Gly	Arg	Thr	Leu	Phe 470	Pro	Gly	Thr	Asp	His 475	Ile	Asp	Gln	Leu	Lys 480
Leu	Ile	Leu	Arg	Leu 485	Val	Gly	Thr	Pro	Gly 490	Ala	Glu	Leu	Leu	Lys 495	Lys
Ile	Ser	Ser	Glu 500	Ser	Ala	Arg	Asn	Tyr 505	Ile	Gln	Ser	Leu	Thr 510	Gln	Met
Pro	Lys	Met 515	Asn	Phe	Ala	Asn	Val 520	Phe	Ile	Gly	Ala	Asn 525	Pro	Leu	Ala
Val	Asp 530	Leu	Leu	Glu	Lys	Met 535	Leu	Val	Leu	Asp	Ser 540	Asp	Lys	Arg	Ile
Thr 545	Ala	Ala	Gln	Ala	Leu 550	Ala	His	Ala	Tyr	Phe 555	Ala	Gln	Tyr	His	Asp 560
Pro	Asp	Asp	Glu	Pro 565	Val	Ala	Asp	Pro	Tyr 570	Asp	Gln	Ser	Phe	G1u 575	Ser
Arg	Asp	Leu	Leu 580	Ile	Asp	Glu	Trp	Lys 585	Ser	Leu	Thr	Tyr	Asp 590	Glu	Val
Ile	Ser	Phe 595	Val	Pro	Pro	Pro	Leu 600	Asp	Gln	Glu	Glu	Met 605	Glu	Ser	

### (2) INFORMATION FOR SEQ ID NO:48:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2907 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2904
  - (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

 		GAG Glu	 	 	 	 	 	48
		GAC Asp						96
	_	GCC Ala						144
 		CTG Leu	 		 	 	 	192
		CAG Gln 70						240
		AAG Lys						288

85 90 95 CGC ACC ATC TTC TTC AAG GAC GAC GGC AAC TAC AAG ACC CGC GCC GAG 336 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 GTG AAG TTC GAG GGC GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC 384 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 115 120 ATC GAC TTC AAG GAG GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC 432 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 130 135 AAC TAC AAC AGC CAC AAC GTC TAT ATC ATG GCC GAC AAG CAG AAG AAC Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 GGC ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 170 165 GTG CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC 576 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 180 185 CCC GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205 AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC 672 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 GTG ACC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TCC 720 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 235 230 GGA CTC AGA TCT ATG AGT GCT GAG GGG TAC CAG TAC AGA GCG CTG TAT Gly Leu Arg Ser Met Ser Ala Glu Gly Tyr Gln Tyr Arg Ala Leu Tyr 250 GAT TAT AAA AAG GAA AGA GAA GAA GAT ATT GAC TIG CAC TIG GGT GAC 816 Asp Tyr Lys Lys Glu Arg Glu Glu Asp Ile Asp Leu His Leu Gly Asp 260 265 ATA TTG ACT GTG AAT AAA GGG TCC TTA GTA GCT CTT GGA TTC AGT GAT 864 Ile Leu Thr Val Asn Lys Gly Ser Leu Val Ala Leu Gly Phe Ser Asp 280 GGA CAG GAA GCC AGG CCT GAA GAA ATT GGC TGG TTA AAT GGC TAT AAT 912 Gly Gln Glu Ala Arg Pro Glu Glu Ile Gly Trp Leu Asn Gly Tyr Asn 300 295 GAA ACC ACA GGG GAA AGG GGG GAC TTT CCG GGA ACT TAC GTA GAA TAT 960 Glu Thr Thr Gly Glu Arg Gly Asp Phe Pro Gly Thr Tyr Val Glu Tyr

315

ATT GGA A	rg Lys I							1008
CGG CCT C								1056
GAA CAA C Glu Gln G 3			o Asp					1104
CCT GAC A Pro Asp I 370								1152
AAG AAA G Lys Lys G 385					_			1200
AAC CTG G Asn Leu A	Ala Glu I							1248
GAC TTG G Asp Leu G					_			1296
TAT CTC C Tyr Leu L 4		Asn P						1344
GAA ATG A Glu Met I 450						_	_	1392
CAG CTA I Gln Leu I 465								1440
TGG CTT A	Thr Leu (							1488
ACC TCC A								1536
AGC CCT A Ser Pro M		Phe S						1584
AAC CTC A Asn Leu I 530								1632
CGA CAG C Arg Gln I								1680

545	550		555	560
		•	ATG TCC TTA CAA Met Ser Leu Gln 570	
			GAA GTG AAT GAA Glu Val Asn Glu	
	Asp Gly Thr		CGA GAT GCG TCT Arg Asp Ala Ser 605	
			AAA GGG GGA AAT Lys Gly Gly Asn 620	
			TAT GGC TTC TCT Tyr Gly Phe Ser 635	
			AAC CAC TAC CGG Asn His Tyr Arg 650	
			GTG AAA TTA CTT Val Lys Leu Leu	
	Gln Gln Asp		AAA GAA GAT AAT Lys Glu Asp Asn 685	
			ACT CAG TTT CAA Thr Gln Phe Gln 700	
			TAT ACC CGC ACA Tyr Thr Arg Thr 715	
			GCA TTT AAT GAA Ala Phe Asn Glu 730	
			GAG CGG TAC AGC Glu Arg Tyr Ser	
	Phe Lys Arg		GAG AAA GAA ATA Glu Lys Glu Ile 765	
			CGA ATC AGT GAA Arg Ile Ser Glu 780	

AGT A Ser A 785									2400
CGA G Arg G									2448
CTG A									2496
GTT C									2544
GAC C Asp G									2592
GAG A Glu I 865									2640
CTG I									2688
AAA C Lys C									2736
CAT T									2784
TAT A									2832
ACC Thr S									2880
CCA (		_	_		TGA				2907

### (2) INFORMATION FOR SEQ ID NO:49:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 968 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

### (v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser Gly Leu Arg Ser Met Ser Ala Glu Gly Tyr Gln Tyr Arg Ala Leu Tyr Asp Tyr Lys Lys Glu Arg Glu Glu Asp Ile Asp Leu His Leu Gly Asp Ile Leu Thr Val Asn Lys Gly Ser Leu Val Ala Leu Gly Phe Ser Asp Gly Gln Glu Ala Arg Pro Glu Glu Ile Gly Trp Leu Asn Gly Tyr Asn Glu Thr Thr Gly Glu Arg Gly Asp Phe Pro Gly Thr Tyr Val Glu Tyr Ile Gly Arg Lys Lys Ile Ser Pro Pro Thr Pro Lys Pro Arg Pro Pro 325 . 330 Arg Pro Leu Pro Val Ala Pro Gly Ser Ser Lys Thr Glu Ala Asp Val Glu Gln Gln Ala Leu Thr Leu Pro Asp Leu Ala Glu Gln Phe Ala Pro Pro Asp Ile Ala Pro Pro Leu Leu Ile Lys Leu Val Glu Ala Ile Glu Lys Lys Gly Leu Glu Cys Ser Thr Leu Tyr Arg Thr Gln Ser Ser Asn Leu Ala Glu Leu Arg Gln Leu Leu Asp Cys Asp Thr Pro Ser Val Asp Leu Glu Met Ile Asp Val His Val Leu Ala Asp Ala Phe Lys Arg 

Tyr Leu Leu Asp Leu Pro Asn Pro Val Ile Pro Ala Ala Val Tyr Ser Glu Met Ile Ser Leu Ala Pro Glu Val Gln Ser Ser Glu Glu Tyr Ile Gln Leu Leu Lys Lys Leu Ile Arg Ser Pro Ser Ile Pro His Gln Tyr Trp Leu Thr Leu Gln Tyr Leu Leu Lys His Phe Phe Lys Leu Ser Gln Thr Ser Ser Lys Asn Leu Leu Asn Ala Arg Val Leu Ser Glu Ile Phe Ser Pro Met Leu Phe Arg Phe Ser Ala Ala Ser Ser Asp Asn Thr Glu Asn Leu Ile Lys Val Ile Glu Ile Leu Ile Ser Thr Glu Trp Asn Glu Arg Gln Pro Ala Pro Ala Leu Pro Pro Lys Pro Pro Lys Pro Thr Thr Val Ala Asn Asn Gly Met Asn Asn Asn Met Ser Leu Gln Asn Ala Glu Trp Tyr Trp Gly Asp Ile Ser Arg Glu Glu Val Asn Glu Lys Leu Arg Asp Thr Ala Asp Gly Thr Phe Leu Val Arg Asp Ala Ser Thr Lys Met His Gly Asp Tyr Thr Leu Thr Leu Arg Lys Gly Gly Asn Asn Lys Leu Ile Lys Ile Phe His Arg Asp Gly Lys Tyr Gly Phe Ser Asp Pro Leu Thr Phe Ser Ser Val Val Glu Leu Ile Asn His Tyr Arg Asn Glu Ser Leu Ala Gln Tyr Asn Pro Lys Leu Asp Val Lys Leu Leu Tyr Pro Val Ser Lys Tyr Gln Gln Asp Gln Val Val Lys Glu Asp Asn Ile Glu Ala Val Gly Lys Lys Leu His Glu Tyr Asn Thr Gln Phe Gln Glu Lys Ser Arg Glu Tyr Asp Arg Leu Tyr Glu Glu Tyr Thr Arg Thr Ser Gln Glu Ile Gln Met Lys Arg Thr Ala Ile Glu Ala Phe Asn Glu Thr Ile Lys 730 735 Ile Phe Glu Glu Gln Cys Gln Thr Gln Glu Arg Tyr Ser Lys Glu Tyr 740 745 Ile Glu Lys Phe Lys Arg Glu Gly Asn Glu Lys Glu Ile Gln Arg Ile Met His Asn Tyr Asp Lys Leu Lys Ser Arg Ile Ser Glu Ile Ile Asp Ser Arg Arg Arg Leu Glu Glu Asp Leu Lys Lys Gln Ala Ala Glu Tyr Arg Glu Ile Asp Lys Arg Met Asn Ser Ile Lys Pro Asp Leu Ile Gln Leu Arg Lys Thr Arg Asp Gln Tyr Leu Met Trp Leu Thr Gln Lys Gly Val Arg Gln Lys Lys Leu Asn Glu Trp Leu Gly Asn Glu Asn Thr Glu Asp Gln Tyr Ser Leu Val Glu Asp Asp Glu Asp Leu Pro His His Asp Glu Lys Thr Trp Asn Val Gly Ser Ser Asn Arg Asn Lys Ala Glu Asn Leu Leu Arg Gly Lys Arg Asp Gly Thr Phe Leu Val Arg Glu Ser Ser

 Lys
 Gln
 Cys
 Tyr
 Ala
 Cys
 Ser
 Val
 Val
 Val
 Asp
 Gly
 Gly
 Val
 Lys
 Lys
 Hys
 Gly
 Free Ser
 Lys
 Gly
 Tyr
 Gly
 Tyr
 Gly
 Phe
 Ala
 Gly
 Pre

 Tyr
 Asn
 Leu
 Tyr
 Ser
 Leu
 Lys
 Glu
 Leu
 Val
 Leu
 His
 Tyr
 Gln
 His
 Asn
 Yal
 Tyr
 Gln
 His
 Asn
 Asn
 Val
 Thr
 Leu
 Ala
 Tyr
 960

 Pro
 Val
 Tyr
 Ala
 Gln
 Arg
 Arg
 Leu
 Asn
 Val
 Thr
 Leu
 Ala
 Tyr
 960

 Pro
 Val
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 Ala
 Tyr
 960
 </

### (2) INFORMATION FOR SEQ ID NO:50:

### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2160 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

### (ii) MOLECULE TYPE: cDNA

- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2157
  - (D) OTHER INFORMATION:

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:

			CTG Leu					48
			AAC Asn				_	96
			TAC Tyr 40					144
			GTG Val					192
			TTC Phe					240
			GCC Ala					288
			GAC Asp					336
 			CTG Leu					384

115 120 125 ATC GAC TTC AAG GAG GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 432 130 135 140 AAC TAC AAC AGC CAC AAC GTC TAT ATC ATG GCC GAC AAG CAG AAG AAC Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 480 150 155 GGC ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 528 170 GTG CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 576 180 185 CCC GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 624 195 200 AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG GAG TTC Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 672 210 215 GTG ACC GCC GCG GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TCC 720 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 235 GGA CTC AGA TCT CGA GCT CAA GCT TCG AAT TCG ACC ATG TCG TCC ATC Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Thr Met Ser Ser Ile 768 250 TTG CCA TTC ACG CCG CCA GTT GTG AAG AGA CTG CTG GGA TGG AAG AAG Leu Pro Phe Thr Pro Pro Val Val Lys Arg Leu Leu Gly Trp Lys Lys 816 260 265 270 TCA GCT GGT GGG TCT GGA GGA GCA GGC GGA GGA GAG CAG AAT GGG CAG 864 Ser Ala Gly Gly Ser Gly Gly Ala Gly Gly Glu Gln Asn Gly Gln 280 GAA GAA AAG TGG TGT GAG AAA GCA GTG AAA AGT CTG GTG AAG AAG CTA 912 Glu Glu Lys Trp Cys Glu Lys Ala Val Lys Ser Leu Val Lys Lys Leu 295 300 AAG AAA ACA GGA CGA TTA GAT GAG CTT GAG AAA GCC ATC ACC ACT CAA 960 Lys Lys Thr Gly Arg Leu Asp Glu Leu Glu Lys Ala Ile Thr Thr Gln 310 315 AAC TGT AAT ACT AAA TGT GTT ACC ATA CCA AGC ACT TGC TCT GAA ATT 1008 Asn Cys Asn Thr Lys Cys Val Thr Ile Pro Ser Thr Cys Ser Glu Ile 325 330 TGG GGA CTG AGT ACA CCA AAT ACG ATA GAT CAG TGG GAT ACA ACA GGC

Trp Gly Leu Ser Thr Pro Asn Thr Ile Asp Gln Trp Asp Thr Thr Gly

345

		GAA Glu	_				_				1104
		GGA Gly									1152
		CAC His 390									1200
		AAT Asn									1248
		GTT Val									1296
		ATC Ile									1344
		GAA Glu				_	_	_	_		1392
		CCA Pro 470								GAA Glu 480	1440
		GAC Asp								GGC Gly	1488
									His	AGC Ser	1536
 								Trp		TCA Ser	1584
							Thr			GCA Ala	1632
						Asp				TCA Ser 560	1680
					Val					ACG Thr	1728
										TAC Tyr	1776

580 585 590

ATA GGT GGG GAA GTT TTT GCT GAG TGC CTA AGT GAT AGT GCA ATC TTT 1824 Ile Gly Gly Glu Val Phe Ala Glu Cys Leu Ser Asp Ser Ala Ile Phe 600 595 GTG CAG AGC CCC AAT TGT AAT CAG AGA TAT GGC TGG CAC CCT GCA ACA 1872 Val Gln Ser Pro Asn Cys Asn Gln Arg Tyr Gly Trp His Pro Ala Thr 615 620 GTG TGT AAA ATT CCA CCA GGC TGT AAT CTG AAG ATC TTC AAC AAC CAG 1920 Val Cys Lys Ile Pro Pro Gly Cys Asn Leu Lys Ile Phe Asn Asn Gln 630 635 GAA TTT GCT GCT CTG GCT CAG TCT GTT AAT CAG GGT TTT GAA GCC 1968 Glu Phe Ala Ala Leu Leu Ala Gln Ser Val Asn Gln Gly Phe Glu Ala 650 645 GTC TAT CAG CTA ACT AGA ATG TGC ACC ATA AGA ATG AGT TTT GTG AAA 2016 Val Tyr Gln Leu Thr Arg Met Cys Thr Ile Arg Met Ser Phe Val Lys 665 GGG TGG GGA GCA GAA TAC CGA AGG CAG ACG GTA ACA AGT ACT CCT TGC 2064 Gly Trp Gly Ala Glu Tyr Arg Arg Gln Thr Val Thr Ser Thr Pro Cys 675 680 TGG ATT GAA CTT CAT CTG AAT GGA CCT CTA CAG TGG TTG GAC AAA GTA 2112 Trp Ile Glu Leu His Leu Asn Gly Pro Leu Gln Trp Leu Asp Lys Val 690 695 700 TTA ACT CAG ATG GGA TCC CCT TCA GTG CGT TGC TCA AGC ATG TCA TAA 2160 Leu Thr Gln Met Gly Ser Pro Ser Val Arg Cys Ser Ser Met Ser 710 715

### (2) INFORMATION FOR SEQ ID NO:51:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 719 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

 Met
 Val
 Ser
 Lys
 Gly
 Glu
 Glu
 Leu
 Phe
 Thr
 Gly
 Val
 Pro
 Ile
 Leu

 1
 5
 5
 10
 15
 15

 Val
 Glu
 Glu
 Leu
 Asp
 Gly
 Asp
 Val
 Asp
 Gly
 His
 Lys
 Phe
 Ser
 Val
 Ser
 Gly

 Glu
 Gly
 Gly
 Asp
 Ala
 Thr
 Tyr
 Gly
 Lys
 Leu
 Thr
 Leu
 Thr
 Leu
 Thr
 Ile
 Leu
 Thr
 Gly
 Asp
 His
 Met
 Lys
 Leu
 Thr
 Thr
 Leu
 Lys
 Thr
 Thr

Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Thr Met Ser Ser Ile Leu Pro Phe Thr Pro Pro Val Val Lys Arg Leu Leu Gly Trp Lys Lys Ser Ala Gly Gly Ser Gly Gly Ala Gly Gly Glu Gln Asn Gly Gln Glu Glu Lys Trp Cys Glu Lys Ala Val Lys Ser Leu Val Lys Lys Leu Lys Lys Thr Gly Arg Leu Asp Glu Leu Glu Lys Ala Ile Thr Thr Gln Asn Cys Asn Thr Lys Cys Val Thr Ile Pro Ser Thr Cys Ser Glu Ile Trp Gly Leu Ser Thr Pro Asn Thr Ile Asp Gln Trp Asp Thr Thr Gly Leu Tyr Ser Phe Ser Glu Gln Thr Arg Ser Leu Asp Gly Arg Leu Gln Val Ser His Arg Lys Gly Leu Pro His Val Ile Tyr Cys Arg Leu Trp Arg Trp Pro Asp Leu His Ser His His Glu Leu Lys Ala Ile Glu Asn Cys Glu Tyr Ala Phe Asn Leu Lys Lys Asp Glu Val Cys Val Asn Pro Tyr His Tyr Gln Arg Val Glu Thr Pro Val Leu Pro Pro Val Leu Val Pro Arg His Thr Glu Ile Leu Thr Glu Leu Pro Pro Leu Asp Asp Tyr Thr His Ser Ile Pro Glu Asn Thr Asn Phe Pro Ala Gly Ile Glu Pro Gln Ser Asn Tyr Ile Pro Glu Thr Pro Pro Pro Gly Tyr Ile Ser Glu Asp Gly Glu Thr Ser Asp Gln Gln Leu Asn Gln Ser Met Asp Thr Gly Ser Pro Ala Glu Leu Ser Pro Thr Thr Leu Ser Pro Val Asn His Ser Leu Asp Leu Gln Pro Val Thr Tyr Ser Glu Pro Ala Phe Trp Cys Ser Ile Ala Tyr Tyr Glu Leu Asn Gln Arg Val Gly Glu Thr Phe His Ala 

Ser 545	Gln	Pro	Ser	Leu	Thr 550	Val	Asp	Gly	Phe	Thr 555	Asp	Pro	Ser	Asn	Ser 560
				Leu 565					570					575	Thr
			580	Arg				585					590	_	-
		595		Val			600					605			
	610			Asn		615					620				
625				Pro	630					635					640
				Leu 645					650					655	
			660	Thr				665					670		
		675		Glu			680					685			_
	690			His		695					700				Val
Leu 705	Thr	Gln	Met	Gly	Ser 710	Pro	Ser	Val		Cys 715	Ser	Ser	Met	Ser	

# (2) INFORMATION FOR SEQ ID NO:52:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2421 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: Coding Sequence
    - (B) LOCATION: 1...2418
    - (D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

ATG Met 1	GTG Val	AGC Ser	AAG Lys	GGC Gly 5	GAG Glu	GAG Glu	CTG Leu	TTC Phe	ACC Thr 10	GGG Gly	GTG Val	GTG Val	CCC Pro	ATC Ile 15	CTG Leu		48
GTC Val	GAG Glu	CTG Leu	GAC Asp 20	GGC Gly	GAC Asp	GTA Val	AAC Asn	GGC Gly 25	CAC His	AAG Lys	TTC Phe	AGC Ser	GTG Val 30	TCC Ser	GGC		96
GAG Glu	GGC Gly	GAG Glu 35	GGC Gly	GAT Asp	GCC Ala	ACC Thr	TAC Tyr 40	GGC Gly	AAG Lys	CTG Leu	ACC Thr	CTG Leu 45	AAG Lys	TTC Phe	ATC Ile		144
TGC Cys	ACC Thr 50	ACC Thr	GGC Gly	AAG Lys	CTG Leu	CCC Pro 55	GTG Val	CCC Pro	TGG Trp	CCC Pro	ACC Thr 60	CTC Leu	GTG Val	ACC Thr	ACC Thr	:	192
CTG Leu	ACC Thr	TAC Tyr	GGC Gly	GTG Val	CAG Gln	TGC Cys	TTC Phe	AGC Ser	CGC Arg	TAC Tyr	CCC Pro	GAC Asp	CAC His	ATG Met	AAG Lys	:	240

65	70	75	80
	AAG TCC GCC ATG CCC Lys Ser Ala Met Pro 90		_
	AAG GAC GAC GGC AAC Lys Asp Asp Gly Asn 105	_	
	GAC ACC CTG GTG AAC Asp Thr Leu Val Asn 120		
	G GAC GGC AAC ATC CTG A Asp Gly Asn Ile Leu 135		
	C AAC GTC TAT ATC ATG S Asn Val Tyr Ile Met 150		
•	TTC AAG ATC CGC CAC Phe Lys Ile Arg His 170		
	C CAC TAC CAG CAG AAC His Tyr Gln Gln Asn 185		
	C GAC AAC CAC TAC CTG Asp Asn His Tyr Leu 200		
	C GAG AAG CGC GAT CAC n Glu Lys Arg Asp His 215		
	G ATC ACT CTC GGC ATG 7 Ile Thr Leu Gly Met 230		
Gly Leu Arg Ser Arg 245		Ser Asn Ser Thr Met 255	Asp
	G AAT ACA CCA ACA AGT r Asn Thr Pro Thr Ser 265		
	G ATG TGC CAT AGA CAA 1 Met Cys His Arg Gln 280		
	A ATT GAA AGT TTG GTA a Ile Glu Ser Leu Val 295	_	

			TCT Ser 310						960
			GTT Val						1008
			AAA Lys						1056
			CTT Leu						1104
			GAC Asp						1152
			GTT Val 390						1200
			GCT Ala						1248
			GGA Gly						1296
			CAT His						1344
			CTG Leu						1392
			AAC Asn 470						1440
								TCA Ser	1488
		Gly						GCT Ala	1536
	His							GCA Ala	1584
								CAC His	1632

540 530 535 CAC CCG CCT ATG CCG CCC CAT CCC GGA CAT TAC TGG CCT GTT CAC AAT 1680 His Pro Pro Met Pro Pro His Pro Gly His Tyr Trp Pro Val His Asn 550 555 GAG CTT GCA TTC CAG CCT CCC ATT TCC AAT CAT CCT GCT CCT GAG TAT 1728 Glu Leu Ala Phe Gln Pro Pro Ile Ser Asn His Pro Ala Pro Glu Tyr 570 565 TGG TGT TCC ATT GCT TAC TTT GAA ATG GAT GTT CAG GTA GGA GAG ACA 1776 Trp Cys Ser Ile Ala Tyr Phe Glu Met Asp Val Gln Val Gly Glu Thr 585 TTT AAG GTT CCT TCA AGC TGC CCT ATT GTT ACT GTT GAT GGA TAC GTG 1824 Phe Lys Val Pro Ser Ser Cys Pro Ile Val Thr Val Asp Gly Tyr Val 595 600 GAC CCT TCT GGA GGA GAT CGC TTT TGT TTG GGT CAA CTC TCC AAT GTC 1872 Asp Pro Ser Gly Gly Asp Arg Phe Cys Leu Gly Gln Leu Ser Asn Val 615 CAC AGG ACA GAA GCC ATT GAG AGA GCA AGG TTG CAC ATA GGC AAA GGT 1920 His Arg Thr Glu Ala Ile Glu Arg Ala Arg Leu His Ile Gly Lys Gly 635 630 GTG CAG TTG GAA TGT AAA GGT GAA GGT GAT GTT TGG GTC AGG TGC CTT 1968 Val Gln Leu Glu Cys Lys Gly Glu Gly Asp Val Trp Val Arg Cys Leu 650 645 AGT GAC CAC GCG GTC TTT GTA CAG AGT TAC TAC TTA GAC AGA GAA GCT 2016 Ser Asp His Ala Val Phe Val Gln Ser Tyr Tyr Leu Asp Arg Glu Ala 660 GGG CGT GCA CCT GGA GAT GCT GTT CAT AAG ATC TAC CCA AGT GCA TAT 2064 Gly Arg Ala Pro Gly Asp Ala Val His Lys Ile Tyr Pro Ser Ala Tyr 675 680 ATA AAG GTC TTT GAT TTG CGT CAG TGT CAT CGA CAG ATG CAG CAG CAG 2112 Ile Lys Val Phe Asp Leu Arg Gln Cys His Arg Gln Met Gln Gln Gln 695 700 690 GCG GCT ACT GCA CAA GCT GCA GCA GCT GCC CAG GCA GCA GCC GTG GCA 2160 Ala Ala Thr Ala Gln Ala Ala Ala Ala Gln Ala Ala Ala Val Ala 710 715 705 GGA AAC ATC CCT GGC CCA GGA TCA GTA GGT GGA ATA GCT CCA GCT ATC 2208 Gly Asn Ile Pro Gly Pro Gly Ser Val Gly Gly Ile Ala Pro Ala Ile 725 AGT CTG TCA GCT GCT GGA ATT GGT GTT GAT GAC CTT CGT CGC TTA 2256 Ser Leu Ser Ala Ala Ala Gly Ile Gly Val Asp Asp Leu Arg Arg Leu TGC ATA CTC AGG ATG AGT TTT GTG AAA GGC TGG GGA CCG GAT TAC CCA 2304 Cys Ile Leu Arg Met Ser Phe Val Lys Gly Trp Gly Pro Asp Tyr Pro

760

765

AGA CAG AGC ATC AAA GAA ACA CCT TGC TGG ATT GAA ATT CAC TTA CAC 2352 Arg Gln Ser Ile Lys Glu Thr Pro Cys Trp Ile Glu Ile His Leu His 770 775 CGG GCC CTC CAG CTC CTA GAC GAA GTA CTT CAT ACC ATG CCG ATT GCA 2400 Arg Ala Leu Gln Leu Leu Asp Glu Val Leu His Thr Met Pro Ile Ala 785 790 GAC CCA CAA CCT TTA GAC TGA 2421

Asp Pro Gln Pro Leu Asp 805

### (2) INFORMATION FOR SEQ ID NO:53:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 806 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 45 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 65 70 75 80 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 110 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 190 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 230 235 240 Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Asn Ser Thr Met Asp 245 250

Asn Met Ser Ile Thr Asn Thr Pro Thr Ser Asn Asp Ala Cys Leu Ser 260 265 Ile Val His Ser Leu Met Cys His Arg Gln Gly Glu Ser Glu Thr 280 285 275 Phe Ala Lys Arg Ala Ile Glu Ser Leu Val Lys Lys Leu Lys Glu Lys 295 300 Lys Asp Glu Leu Asp Ser Leu Ile Thr Ala Ile Thr Thr Asn Gly Ala 310 315 His Pro Ser Lys Cys Val Thr Ile Gln Arg Thr Leu Asp Gly Arg Leu 325 330 Gln Val Ala Gly Arg Lys Gly Phe Pro His Val Ile Tyr Ala Arg Leu 350 340 345 Trp Arg Trp Pro Asp Leu His Lys Asn Glu Leu Lys His Val Lys Tyr 355 360 Cys Gln Tyr Ala Phe Asp Leu Lys Cys Asp Ser Val Cys Val Asn Pro 375 380 Tyr His Tyr Glu Arg Val Val Ser Pro Gly Ile Asp Leu Ser Gly Leu 390 395 Thr Leu Gln Ser Asn Ala Pro Ser Ser Met Met Val Lys Asp Glu Tyr 405 410 Val His Asp Phe Glu Gly Gln Pro Ser Leu Ser Thr Glu Gly His Ser 420 425 Ile Gln Thr Ile Gln His Pro Pro Ser Asn Arg Ala Ser Thr Glu Thr 440 445 Tyr Ser Thr Pro Ala Leu Leu Ala Pro Ser Glu Ser Asn Ala Thr Ser 455 460 Thr Ala Asn Phe Pro Asn Ile Pro Val Ala Ser Thr Ser Gln Pro Ala 470 475 Ser Ile Leu Gly Gly Ser His Ser Glu Gly Leu Leu Gln Ile Ala Ser 485 490 Gly Pro Gln Pro Gly Gln Gln Gln Asn Gly Phe Thr Gly Gln Pro Ala 500 505 510 Thr Tyr His His Asn Ser Thr Thr Thr Trp Thr Gly Ser Arg Thr Ala 520 515 Pro Tyr Thr Pro Asn Leu Pro His His Gln Asn Gly His Leu Gln His 540 535 His Pro Pro Met Pro Pro His Pro Gly His Tyr Trp Pro Val His Asn 545 550 555 Glu Leu Ala Phe Gln Pro Pro Ile Ser Asn His Pro Ala Pro Glu Tyr 565 570 Trp Cys Ser Ile Ala Tyr Phe Glu Met Asp Val Gln Val Gly Glu Thr 580 585 590 Phe Lys Val Pro Ser Ser Cys Pro Ile Val Thr Val Asp Gly Tyr Val 595 600 605 Asp Pro Ser Gly Gly Asp Arg Phe Cys Leu Gly Gln Leu Ser Asn Val 615 -His Arg Thr Glu Ala Ile Glu Arg Ala Arg Leu His Ile Gly Lys Gly 630 635 Val Gln Leu Glu Cys Lys Gly Glu Gly Asp Val Trp Val Arg Cys Leu 645 650 Ser Asp His Ala Val Phe Val Gln Ser Tyr Tyr Leu Asp Arg Glu Ala 665 Gly Arg Ala Pro Gly Asp Ala Val His Lys Ile Tyr Pro Ser Ala Tyr 680 Ile Lys Val Phe Asp Leu Arg Gln Cys His Arg Gln Met Gln Gln 700 695 Ala Ala Thr Ala Gln Ala Ala Ala Ala Gln Ala Ala Ala Val Ala

715

Gly Asn Ile Pro Gly Pro Gly Ser Val Gly Gly Ile Ala Pro Ala Ile 725 730 735 Ser Leu Ser Ala Ala Ala Gly Ile Gly Val Asp Asp Leu Arg Arg Leu 745 750 740 Cys Ile Leu Arg Met Ser Phe Val Lys Gly Trp Gly Pro Asp Tyr Pro 760 755 765 Arg Gln Ser Ile Lys Glu Thr Pro Cys Trp Ile Glu Ile His Leu His 770 775 780 Arg Ala Leu Gln Leu Leu Asp Glu Val Leu His Thr Met Pro Ile Ala 795 Asp Pro Gln Pro Leu Asp 805

### (2) INFORMATION FOR SEQ ID NO:54:

### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3120 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...3117
  - (D) OTHER INFORMATION:

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

 	 	GAG Glu						48
		GAC Asp						96
 		GCC Ala						144
 	 	CTG Leu						192
		CAG Gln 70				_		240
 		AAG Lys						288
		AAG Lys					_	336

					AAC Asn				384
					CTG Leu				432
					ATG Met				480
					CAC His 170				528
					AAC Asn				576
					CTG Leu				624
					CAC His				672
					ATG Met				720
					ATC Ile 250				768
					CTG Leu				816
_	_				TGG Trp				864
					GAC Asp	Arg			912
					CAG Gln				960
					ATC Ile 330				1008
					TGC Cys				1056

									_				33	U		
TGC Cys	TATY	C CG P Ar 35	g Hi	C AT	T CT e Le	G TA	C AA' r Ası 360	n Gl	A CA	G AG n Ar	G CT g Le	G GT u Va 36	l Ar	A GA g Gl	A GCC u Ala	1104
AAC Asn	AA: Asi 370	з Су	C AG	r Se	T CC r Pr	G GC' o Ala 37!	a Gly	G ATY	C CTV	G GT u Va	T GA 1 As 38	p Ala	C AT a Me	G TC	C CAG r Gln	1152
AAG Lys 385	His	C CT	T CA u Gl	G AT	C AA e As 39	n Glı	G ACA	A TT	r GA0 ∋ Glu	G GA0 1 Gl: 39!	u Lei	G CGA	A CTO	G GTG	C ACG l Thr 400	1200
CAG Gln	GAC Asp	AC.	A GA	G AA' u Ası 40!	ı Glı	G CTC	AAC Lys	AA/ Lys	A CTO Leu 410	ı Glı	G CAC	G ACT	CAC	G GAC n Glu 415	TAC Tyr	1248
TTC Phe	ATC	: ATC	C CAG = Gl: 420	ı Tyı	C CAC	G GAG	AGC Ser	CTC Leu 425	Arg	TATO	C CAA	A GCT	CAC Glr 430	Phe	GCC Ala	1296
CAG Gln	CTG Leu	GCC Ala 435	a Glr	G CTC	AGC Ser	Pro	Gln 440	Glu	CGI Arg	CTC Leu	AGC Ser	CGG Arg 445	Glu	ACC Thr	GCC Ala	1344
CTC Leu	CAG Gln 450	CAG Glr	AAC Lys	G CAG	GTG Val	Ser 455	CTG Leu	GAG Glu	GCC	TGG Trp	Leu 460	Gln	CGI	'GAG	GCA Ala	1392
CAG Gln 465	ACA Thr	CTG Leu	Gln	CAG Gln	TAC Tyr 470	Arg	GTG Val	GAG Glu	CTG Leu	GCC Ala 475	GAG Glu	AAG Lys	CAC	CAG Gln	AAG Lys 480	1440
ACC Thr	CTG Leu	CAG Gln	CTG Leu	CTG Leu 485	CGG Arg	AAG Lys	CAG Gln	CAG Gln	ACC Thr 490	ATC Ile	ATC Ile	CTG Leu	GAT qzA	GAC Asp 495	GAG Glu	1488
CTG Leu	ATC Ile	CAG Gln	TGG Trp 500	AAG Lys	CGG Arg	CGG Arg	CAG Gln	CAG G1n 505	CTG Leu	GCC Ala	GGG Gly	AAC Asn	GGC Gly 510	GGG Gly	CCC Pro	1536
Pro (	GAG Glu	GGC Gly 515	AGC Ser	CTG Leu	GAC Asp	GTG Val	CTA Leu 520	CAG Gln	TCC Ser	TGG Trp	TGT Cys	GAG Glu 525	AAG Lys	TTG Leu	GCC Ala	1584
GAG A	ATC Ile 530	ATC Ile	TGG Trp	CAG Gln	AAC Asn	CGG Arg 535	CAG Gln	CAG Gln	ATC Ile	CGC Arg	AGG Arg 540	GCT Ala	GAG Glu	CAC His	CTC Leu	1632
TGC ( Cys ( 545	CAG Sln	CAG Gln	CTG Leu	CCC Pro	ATC Ile 550	CCC Pro	GGC Gly	CCA Pro	GTG Val	GAG Glu 555	GAG Glu	ATG Met	CTG Leu	GCC Ala	GAG Glu 560	1680
GTC A	AAC ( Asn ,	GCC Ala	ACC Thr	ATC Ile 565	ACG Thr	GAC Asp	ATT . Ile	Ile	TCA Ser 570	GCC Ala	CTG Leu	GTG Val	ACC Thr	AGC Ser 575	ACA Thr	1728

TTC A'											1776
TTT GO	la A										1824
ATG AMET AME				_							1872
AAG TY Lys Se 625											1920
ATC C						_					1968
CTC AG		lla	-								2016
GAC CO	rg A										2064
TTT GA Phe G	_		_								2112
AAG AG Lys Ti 705											2160
CAC A				_							2208
GGC AG		al		_	_	_					2256
TGT GA	lu A										2304
GGC CT Gly Le	eu T										2352
AAC AC Asn Se 785										 	 2400
TCC CA											2448

815 810 805 CAG TGG TTT GAC GGG GTG ATG GAG GTG TTG AAG AAG CAC CAC AAG CCC 2496 Gln Trp Phe Asp Gly Val Met Glu Val Leu Lys Lys His His Lys Pro 825 820 CAC TGG AAT GAT GGG GCC ATC CTA GGT TTT GTG AAT AAG CAA CAG GCC 2544 His Trp Asn Asp Gly Ala Ile Leu Gly Phe Val Asn Lys Gln Gln Ala 840 CAC GAC CTG CTC ATC AAC AAG CCC GAC GGG ACC TTC TTG TTG CGC TTT 2592 His Asp Leu Leu Ile Asn Lys Pro Asp Gly Thr Phe Leu Leu Arg Phe 855 850 AGT GAC TCA GAA ATC GGG GGC ATC ACC ATC GCC TGG AAG TTT GAC TCC 2640 Ser Asp Ser Glu Ile Gly Gly Ile Thr Ile Ala Trp Lys Phe Asp Ser 875 870 CCG GAA CGC AAC CTG TGG AAC CTG AAA CCA TTC ACC ACG CGG GAT TTC Pro Glu Arg Asn Leu Trp Asn Leu Lys Pro Phe Thr Thr Arg Asp Phe 890 885 TCC ATC AGG TCC CTG GCT GAC CGG CTG GGG GAC CTG AGC TAT CTC ATC 2736 Ser Ile Arg Ser Leu Ala Asp Arg Leu Gly Asp Leu Ser Tyr Leu Ile 900 905 TAT GTG TTT CCT GAC CGC CCC AAG GAT GAG GTC TTC TCC AAG TAC TAC Tyr Val Phe Pro Asp Arg Pro Lys Asp Glu Val Phe Ser Lys Tyr Tyr 920 915 2832 ACT CCT GTG CTG GCT AAA GCT GTT GAT GGA TAT GTG AAA CCA CAG ATC Thr Pro Val Leu Ala Lys Ala Val Asp Gly Tyr Val Lys Pro Gln Ile 930 935 940 AAG CAA GTG GTC CCT GAG TTT GTG AAT GCA TCT GCA GAT GCT GGG GGC 2880 Lys Gln Val Val Pro Glu Phe Val Asn Ala Ser Ala Asp Ala Gly Gly 955 945 950 AGC AGC GCC ACG TAC ATG GAC CAG GCC CCC TCC CCA GCT GTG TGC CCC Ser Ser Ala Thr Tyr Met Asp Gln Ala Pro Ser Pro Ala Val Cys Pro 965 CAG GCT CCC TAT AAC ATG TAC CCA CAG AAC CCT GAC CAT GTA CTC GAT 2976 Gln Ala Pro Tyr Asn Met Tyr Pro Gln Asn Pro Asp His Val Leu Asp 980 985 CAG GAT GGA GAA TTC GAC CTG GAT GAG ACC ATG GAT GTG GCC AGG CAC 3024 Gln Asp Gly Glu Phe Asp Leu Asp Glu Thr Met Asp Val Ala Arg His 1000 GTG GAG GAA CTC TTA CGC CGA CCA ATG GAC AGT CTT GAC TCC CGC CTC 3072 Val Glu Glu Leu Leu Arg Arg Pro Met Asp Ser Leu Asp Ser Arg Leu 1015 TCG CCC CCT GCC GGT CTT TTC ACC TCT GCC AGA GGC TCC CTC TCA TGA 3120 Ser Pro Pro Ala Gly Leu Phe Thr Ser Ala Arg Gly Ser Leu Ser

1035

1030

# (2) INFORMATION FOR SEQ ID NO:55:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1039 amino acids
- " (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

Met 1	Val	Ser	Lys	Gly 5	Glu	Glu	Leu	Phe	Thr 10	Gly	Val	Val	Pro	Ile 15	Leu
Val	Glu	Leu	Asp 20	Gly	Asp	Val	Asn	Gly 25	His	Lys	Phe	Ser	Val 30	Ser	Gly
	Gly	35					40					45			
_	Thr 50					55					60				
65	Thr	-			70					75					80
	His			85					90					95	
_	Thr		100					105					110		
	Lys	115					120					125			
	Asp 130					135					140				
145	Tyr				150					155					160
	Ile			165					170					175	
	Gln		180					185					190		
	Val	195					200					205			
	Lys 210					215					220				
225	Thr				230					235					240
	Leu			245					250					255	
	Asp		260					265					270		
	Glu	275					280					285			
-	Ala 290					295					300				
305	Leu			•	310					315					320
	Gly			325					330					335	
Thr	Gln	Leu	Gln 340	Lys	Thr	Tyr	Asp	Arg 345	Cys	Pro	Leu	Glu	Leu 350	Val	Arg

Cy:	s Il	e Ar 35	g Hi 5	s Il	e Lei	л Ту:	r Ası	n Gl	u Gl	n Ar	g Le	u Va:		g Glı	ı Ala
	3/	J				379	a Gly	y Il			380	o Ala	a Mei	t Sei	
38	5				390	)				399	ı Len	ı Arç		ı Val	400
				40	5				410	u Gli	n Glr			1 Glu 415	Туг
			42	0				425	5				430	ı Phe	e Ala
		43	5				440	)				445	g Glu	ı Thr	
	450	)				455	5				460	)		g Glu	
465	•				470	)				475	,			Glņ	480
				485	5				490	)				Asp 495	Glu
			500	)				505	;				510	Gly	·
		515	)				520					525		Leu	
	530					535					540			His	
545					550					555				Ala	560
				565					570					Ser 575	
			580					585					590	Thr	_
		595					600					605		Val	
	610					615					620			Gln	
625					630					635				Gly	640
				645					650					Gly 655	
			660					665					670	Arg	
		675					680					685		Val	
	690					695					700			Gln	
705					710					715				Gln	720
				725					730					Glu 735	
			740					745					750	Gln	
		755					760					765		Asn	
	770					775					780			Phe	
785					790					795				Ser	800
ser	GIN	rne	Asn	Arg 805	Glu .	Asn	Leu		Gly 810	Trp	Asn	Тут	Thr	Phe 815	Trp

Gln Trp Phe Asp Gly Val Met Glu Val Leu Lys Lys His His Lys Pro 825 His Trp Asn Asp Gly Ala Ile Leu Gly Phe Val Asn Lys Gln Gln Ala 845 840 835 His Asp Leu Leu Ile Asn Lys Pro Asp Gly Thr Phe Leu Leu Arg Phe 855 860 Ser Asp Ser Glu Ile Gly Gly Ile Thr Ile Ala Trp Lys Phe Asp Ser 875 870 Pro Glu Arg Asn Leu Trp Asn Leu Lys Pro Phe Thr Thr Arg Asp Phe 885 890 Ser Ile Arg Ser Leu Ala Asp Arg Leu Gly Asp Leu Ser Tyr Leu Ile 910 900 905 Tyr Val Phe Pro Asp Arg Pro Lys Asp Glu Val Phe Ser Lys Tyr Tyr 915 920 925 Thr Pro Val Leu Ala Lys Ala Val Asp Gly Tyr Val Lys Pro Gln Ile 940 930 935 Lys Gln Val Val Pro Glu Phe Val Asn Ala Ser Ala Asp Ala Gly Gly 950 955 Ser Ser Ala Thr Tyr Met Asp Gln Ala Pro Ser Pro Ala Val Cys Pro 965 970 975 Gln Ala Pro Tyr Asn Met Tyr Pro Gln Asn Pro Asp His Val Leu Asp 980 985 Gln Asp Gly Glu Phe Asp Leu Asp Glu Thr Met Asp Val Ala Arg His 995 1000 1005 Val Glu Glu Leu Leu Arg Arg Pro Met Asp Ser Leu Asp Ser Arg Leu 1010 1015 1020 Ser Pro Pro Ala Gly Leu Phe Thr Ser Ala Arg Gly Ser Leu Ser 1030 1035 025

### (2) INFORMATION FOR SEQ ID NO:56:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1875 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...1872
  - (D) OTHER INFORMATION:

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

ATG GCG GCG GCG GCG GCC GCT CCG GGG GGC GGG GGC GGG GAG CCC AGG

Met Ala Ala Ala Ala Ala Ala Pro Gly Gly Gly Gly Gly Glu Pro Arg

1 5 10 15

GGA ACT GCT GGG GTC GTC CCG GTG GTC CCC GGG GAG GTG GAG GTG GTG

Gly Thr Ala Gly Val Val Pro Val Val Pro Gly Glu Val Glu Val Val

20 25 30

AAG GGG CAG CCA TTC GAT GTG GGC CCA CGC TAC ACG CAG CTG CAG TAC

Lys Gly Gln Pro Phe Asp Val Gly Pro Arg Tyr Thr Gln Leu Gln Tyr

40 45

ATC GG Ile Gl									192
CGC AA Arg Ly 65									240
ACC TA Thr Ty		_							288
CGC CA Arg Hi									336
CTG GA Leu Gl									384
GAC CT Asp Le 13	ı Tyr								432
TGC TA Cys Ty 145								-	480
GCC AA									528
ACC AC									576
GAC CC									624
CGC TG Arg Tr	y Tyr								672
AAA TCC Lys Se: 225									720
TCC AAG									768
CAC AT									816
ATC AT									864

275 280 285 ACC AAG GTG GCT TGG GCC AAG CTC TTT CCT AAA TCT GAC TCC AAA GCT 912 Thr Lys Val Ala Trp Ala Lys Leu Phe Pro Lys Ser Asp Ser Lys Ala 290 295 CTT GAC CTG CTG GAC CGG ATG TTA ACC TTC AAC CCA AAC AAG CGC ATC 960 Leu Asp Leu Leu Asp Arg Met Leu Thr Phe Asn Pro Asn Lys Arg Ile 310 ACA GTA GAG GAA GCG CTG GCT CAC CCT TAC CTG GAA CAG TAC TAC GAT 1008 Thr Val Glu Glu Ala Leu Ala His Pro Tyr Leu Glu Gln Tyr Tyr Asp 325 CCG ACA GAT GAG CCA GTG GCC GAG GAG CCA TTC ACC TTC GAC ATG GAG 1056 Pro Thr Asp Glu Pro Val Ala Glu Glu Pro Phe Thr Phe Asp Met Glu 340 345 CTG GAT GAC CTC CCC AAG GAG CGG CTG AAG GAG TTG ATC TTC CAG GAG 1104 Leu Asp Asp Leu Pro Lys Glu Arg Leu Lys Glu Leu Ile Phe Gln Glu 355 360 ACA GCC CGC TTC CAG CCA GGG GCG CCA GAG GGC CCC GGG CGC GCC ATG Thr Ala Arg Phe Gln Pro Gly Ala Pro Glu Gly Pro Gly Arg Ala Met 370 375 AGT AAA GGA GAA GAA CTT TTC ACT GGA GTT GTC CCA ATT CTT GTT GAA 1200 Ser Lys Gly Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu 385 390 395 TTA GAT GGC GAT GTT AAT GGG CAA AAA TTC TCT GTT AGT GGA GAG GGT 1248 Leu Asp Gly Asp Val Asn Gly Gln Lys Phe Ser Val Ser Gly Glu Gly 410 GAA GGT GAT GCA ACA TAC GGA AAA CTT ACC CTT AAA TTT ATT TGC ACT 1296 Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr 425 ACT GGG AAG CTA CCT GTT CCA TGG CCA ACG CTT GTC ACT ACT CTC ACT 1344 Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr 435 440 TAT GGT GTT CAA TGC TTT TCT AGA TAC CCA GAT CAT ATG AAA CAG CAT 1392 Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His 455 GAC TTT TTC AAG AGT GCC ATG CCC GAA GGT TAT GTA CAG GAA AGA ACT 1440 Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr 470 ATA TTT TAC AAA GAT GAC GGG AAC TAC AAG ACA CGT GCT GAA GTC AAG 1488 Ile Phe Tyr Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys 485 490 TTT GAA GGT GAT ACC CTT GTT AAT AGA ATC GAG TTA AAA GGT ATT GAT 1536

Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp

505

500

	AAA Lys			-	 	 	 			1584
	TCA Ser 530									1632
	GTT Val									1680
	GCA Ala									1728
	TTA Leu									1776
	CCC Pro	_								1824
	GCT Ala 610							_	GAG T Glu	1873
AA										1875

- (2) INFORMATION FOR SEQ ID NO:57:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 624 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (v) FRAGMENT TYPE: internal
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

Met Ala Ala Ala Ala Ala Pro Gly Gly Gly Gly Glu Pro Arg 10 Gly Thr Ala Gly Val Val Pro Val Val Pro Gly Glu Val Glu Val Val 20 25 30 Lys Gly Gln Pro Phe Asp Val Gly Pro Arg Tyr Thr Gln Leu Gln Tyr 35 40 45 Ile Gly Glu Gly Ala Tyr Gly Met Val Ser Ser Ala Tyr Asp His Val 55 60 Arg Lys Thr Arg Val Ala Ile Lys Lys Ile Ser Pro Phe Glu His Gln 70 75 Thr Tyr Cys Gln Arg Thr Leu Arg Glu Ile Gln Ile Leu Leu Arg Phe 90 95 85 Arg His Glu Asn Val Ile Gly Ile Arg Asp Ile Leu Arg Ala Pro Thr 100 105 110 Leu Glu Ala Met Arg Asp Val Tyr Ile Val Gln Asp Leu Met Glu Thr

			115					120					125			
2	Asp	Leu 130	Tyr	Lys	Leu	Leu	Lys 135	Ser	Gln	Gln	Leu	Ser 140	Asn	Asp	His	Ile
	Cys 145	Tyr	Phe	Leu	Tyr	Gln 150	Ile	Leu	Arg	Gly	Leu 155	Lys	Tyr	Ile	His	Ser 160
					165		_		_	170					Ile 175	
			_	180				_	185		_			190	Ile	
	-		195		_			200					205		Ala	
	_	210	_				215					220			Tyr	
	225					230					235				Met	240
					245			_	_	250	_		-		Leu 255	
				260					265					270	Asn	
			275		-			280					285		Ser Lys	
		290			_		295					300			Arg	
	305	_				310					315				Tyr	320
					325					330				_	335 Met	_
			_	340					345					350	Gln	
			355					360					365		Ala	
		370	_				375					380			Val	
	385	_	_			390			_		395				Glu	400
		_	_	_	405					410					415 Cys	
				420					425					430	Leu	
			435					440					445		Gln	
	Asp	450 Phe	Phe	Lys	Ser	Ala	455 Met	Pro	Glu	Gly	Tyr	460 Val	Gln	Glu	Arg	Thr
	465					470					475					480 Lys
	Phe	Glu	Gly	Asp	485 Thr	Leu	Val	Asn	Arg	490 Ile	Glu	Leu	Lys	Gly	495 Ile	Asp
	Phe	Lys	Glu	500 Asp	Gly	Asn	Ile	Leu	505 Gly	His	Lys	Met	Glu	510 Tyr	Asn	Tyr
	Asn	Ser	515 His	Asn	Val	Tyr	Ile	520 Met	Ala	Asp	Lys	Pro	525 Lys	Asn	Gly	Ile
	Lys	530 Val	Asn	Phe	Lys	Ile	535 Arg	His	Asn	Ile	Lys	540 Asp	Gly	Ser	Val	Gln
	545 Leu	Ala	Asp	His	Tyr	550 Gln	Gln	Asn	Thr	Pro	555 Ile	Gly	Asp	Gly	Pro	560 Val
	Leu	Leu	Pro	Asp	565 Asn	His	Tyr	Leu	Ser	570 Thr	Gln	Ser	Ala	Leu	575 Ser	Lys

580 585 Asp Pro Asn Glu Lys Arg Asp His Met Ile Leu Leu Glu Phe Val Thr 600 605 Ala Ala Gly Ile Thr His Gly Met Asp Glu Leu Tyr Lys Pro Gln Glu 615 (2) INFORMATION FOR SEQ ID NO:58: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1815 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...1811 (D) OTHER INFORMATION: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:58: ATG GCG GCG GCG GCG GCG GCC CCG GAG ATG GTC CGC GGG CAG GTG 48 Met Ala Ala Ala Ala Ala Gly Pro Glu Met Val Arg Gly Gln Val 10 TTC GAC GTG GGG CCG CGC TAC ACT AAT CTC TCG TAC ATC GGA GAA GGC Phe Asp Val Gly Pro Arg Tyr Thr Asn Leu Ser Tyr Ile Gly Glu Gly 96 25 GCC TAC GGC ATG GTT TGT TCT GCT TAT GAT AAT CTC AAC AAA GTT CGA Ala Tyr Gly Met Val Cys Ser Ala Tyr Asp Asn Leu Asn Lys Val Arg 144 35 40 GTT GCT ATC AAG AAA ATC AGT CCT TTT GAG CAC CAG ACC TAC TGT CAG 192 Val Ala Ile Lys Lys Ile Ser Pro Phe Glu His Gln Thr Tyr Cys Gln 50 55 AGA ACC CTG AGA GAG ATA AAA ATC CTA CTG CGC TTC AGA CAT GAG AAC 240 Arg Thr Leu Arg Glu Ile Lys Ile Leu Leu Arg Phe Arg His Glu Asn 70 ATC ATC GGC ATC AAT GAC ATC ATC CGG GCA CCA ACC ATT GAG CAG ATG 288 Ile Ile Gly Ile Asn Asp Ile Ile Arg Ala Pro Thr Ile Glu Gln Met 85 90 AAA GAT GTA TAT ATA GTA CAG GAC CTC ATG GAG ACA GAT CTT TAC AAG Lys Asp Val Tyr Ile Val Gln Asp Leu Met Glu Thr Asp Leu Tyr Lys 336 100 105 CTC TTG AAG ACA CAG CAC CTC AGC AAT GAT CAT ATC TGC TAT TTT CTT 384 Leu Leu Lys Thr Gln His Leu Ser Asn Asp His Ile Cys Tyr Phe Leu 115 120 125 TAT CAG ATC CTG AGA GGA TTA AAG TAT ATA CAT TCA GCT AAT GTT CTG Tyr Gln Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala Asn Val Leu

130

135

CAC CGT ( His Arg A 145													480
CTC AAG A													528
GAT CAT A				Glu									576
GCT CCA (			Asn :										624
ATT TGG ( Ile Trp ( 210													672
ATC TTC ( Ile Phe 1 225													720
ATT CTT (													768
AAA GCT A				Ser									816
TGG AAC I			Asn .										864
GAT AAA A Asp Lys I 290													912
GCT CTG (Ala Leu 2													960
CCC ATT (													1008
CCT AAG ( Pro Lys (													1056
CAG CCA C			Met 2										1104
AAG GGC (	GAG GAG	CTG TTC	ACC (	GGG	GTG	GTG	CCC	ATC	CTG	GTC	GAG	CTG	1152

Lys Gl	ly ( 70	Glu	Glu	Leu	Phe	Thr 375	Gly	Val	Val	Pro	Ile 380	Leu	Val	Glu	Leu	
GAC GC Asp G: 385						_										1200
GGC GA		_														1248
GGC A																1296
GGC G	al (															1344
TTC T Phe Pl 4																1392
TTC T Phe Pl 465																1440
GAG GG																1488
AAG G																1536
AGC CA	is A															1584
GTG A																1632
GCC GA Ala As 545																1680
CTG CC																1728
CCC A																1776
GCC GG Ala G	ly		_		_							GTAA			·	1815

#### (2) INFORMATION FOR SEQ ID NO:59:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 604 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:

Met Ala Ala Ala Ala Ala Gly Pro Glu Met Val Arg Gly Gln Val -5 10 Phe Asp Val Gly Pro Arg Tyr Thr Asn Leu Ser Tyr Ile Gly Glu Gly 25 20 Ala Tyr Gly Met Val Cys Ser Ala Tyr Asp Asn Leu Asn Lys Val Arg 40 Val Ala Ile Lys Lys Ile Ser Pro Phe Glu His Gln Thr Tyr Cys Gln 55 Arg Thr Leu Arg Glu Ile Lys Ile Leu Leu Arg Phe Arg His Glu Asn 70 75 Ile Ile Gly Ile Asn Asp Ile Ile Arg Ala Pro Thr Ile Glu Gln Met 90 Lys Asp Val Tyr Ile Val Gln Asp Leu Met Glu Thr Asp Leu Tyr Lys 105 110 Leu Leu Lys Thr Gln His Leu Ser Asn Asp His Ile Cys Tyr Phe Leu 120 125 115 Tyr Gln Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala Asn Val Leu 130 135 140 His Arg Asp Leu Lys Pro Ser Asn Leu Leu Leu Asn Thr Thr Cys Asp 150 155 Leu Lys Ile Cys Asp Phe Gly Leu Ala Arg Val Ala Asp Pro Asp His 165 170 Asp His Thr Gly Phe Leu Thr Glu Tyr Val Ala Thr Arg Trp Tyr Arg 185 180 Ala Pro Glu Ile Met Leu Asn Ser Lys Gly Tyr Thr Lys Ser Ile Asp 200 205 Ile Trp Ser Val Gly Cys Ile Leu Ala Glu Met Leu Ser Asn Arg Pro 215 220 Ile Phe Pro Gly Lys His Tyr Leu Asp Gln Leu Asn His Ile Leu Gly 230 235 Ile Leu Gly Ser Pro Ser Gln Glu Asp Leu Asn Cys Ile Ile Asn Leu 245 250 Lys Ala Arg Asn Tyr Leu Leu Ser Leu Pro His Lys Asn Lys Val Pro 265 270 260 Trp Asn Arg Leu Phe Pro Asn Ala Asp Ser Lys Ala Leu Asp Leu Leu 280 285 Asp Lys Met Leu Thr Phe Asn Pro His Lys Arg Ile Glu Val Glu Gln 295 300 Ala Leu Ala His Pro Tyr Leu Glu Gln Tyr Tyr Asp Pro Ser Asp Glu 310 315 Pro Ile Ala Glu Ala Pro Phe Lys Phe Asp Met Glu Leu Asp Asp Leu 325 330 Pro Lys Glu Lys Leu Lys Glu Leu Ile Phe Glu Glu Thr Ala Arg Phe

			340					345					350		
		355	-				360					365		Val	
Lys	Gly 370	Glu	Glu	Leu	Phe	Thr 375	Gly	Val	Val	Pro	Ile 380	Leu	Val	Glu	Leu
Asp 385	Gly	Asp	Val	Asn	Gly 390	His	Lys	Phe	Ser	Val 395	Ser	Gly	Glu	Gly	Glu 400
Gly	Asp	Ala	Thr	Tyr 405	Gly	Lys	Leu	Thr	Leu 410	Lys	Phe	Ile	Cys	Thr 415	Thr
Gly	Lys	Leu	Pro 420	Val	Pro	Trp	Pro	Thr 425	Leu	Val	Thr	Thr	Leu 430	Thr	Tyr
Gly	Val	Gln 435	Cys	Phe	Ser	Arg	Tyr 440	Pro	Asp	His	Met	Lys 445	Gln	His	Asp
Phe	Phe 450	Lys	Ser	Ala	Met	Pro 455	Glu	Gly	Tyr	Val	Gln 460	Glu	Arg	Thr	Ile
Phe 465	Phe	Lys	Asp	Asp	Gly 470	Asn	Tyr	Lys	Thr	Arg 475	Ala	Glu	Val	Lys	Phe 480
Glu	Gly	Asp	Thr	Leu 485	Val	Asn	Arg	Ile	Glu 490	Leu	Lys	Gly	Ile	Asp 495	Phe
Lys	Glu	Asp	Gly 500	Asn	Ile	Leu	Gly	His 505	Lys	Leu	Glu	Tyr	Asn 510	Tyr	Asn
Ser	His	Asn 515	Val	Tyr	Ile	Met	Ala 520	Asp	Lys	Gln	Lys	Asn 525	Gly	Ile	Lys
Val	Asn 530	Phe	Lys	Ile	Arg	His 535	Asn	Ile	Glu	Asp	Gly 540	Ser	Val	Gln	Leu
Ala 545	Asp	His	Tyr	Gln	Gln 550	Asn	Thr	Pro	Ile	Gly 555	_	Gly	Pro	Val	Leu 560
Leu	Pro	Asp	Asn	His 565	Tyr	Leu	Ser	Thr	Gln 570	Ser	Ala	Leu	Ser	Lys 575	Asp
Pro	Asn	Glu	Lys 580	Arg	Asp	His	Met	Val 585	Leu	Leu	Glu	Phe	Val 590	Thr	Ala
Ala	Gly	Ile 595	Thr	Leu	Gly	Met	Asp 600	Glu	Leu	Tyr	Lys				

- (2) INFORMATION FOR SEQ ID NO:60:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2511 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2508
  - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:
- ATG GAG CTG GAA AAC ATC GTG GCC AAC ACG GTC TTG CTG AAA GCC AGG

  Met Glu Leu Glu Asn Ile Val Ala Asn Thr Val Leu Leu Lys Ala Arg

  1 5 10 15

	CTG Leu															144
	GAC Asp 50															192
	CTT Leu															240
	CAG Gln															288
	CTG Leu															336
	TCC Ser															384
	GAG Glu 130															432
	GCA Ala															480
	CTG Leu															528
	AGG Arg															576
	AAA Lys															624
	AAA Lys 210															672
	AAA Lys		_													720
	AAC Asn														Lys	768
GAT	GCA	CTG	TGC	TTG	GTC	CTG	ACC	ATC	ATG	AAT	GGG	GGT	GAC	CTG	AAG	816

Asp	Ala	Leu	Cys 260	Leu	Val	Leu	Thr	11e 265	Met	Asn	Gly	Gly	Asp 270	Leu	Lys	
					ATG Met											864
					GAG Glu											912
					CGA Arg 310											960
				_	AGG Arg							_	_		_	1008
					ATC Ile							_	_			1056
					AAC Asn											1104
					CTC Leu											1152
					GAG Glu 390											1200
					GAG Glu								_	_	_	1248
					ATG Met								_			1296
					GGG Gly	Ala						_			TTC Phe	1344
					AAG Lys											1392
					CGC Arg 470							_			_	1440
					GTG Val							_	_			1488

						TCC Ser										1536
						GAA Glu				_			_	_	_	1584
						CCA Pro 535										1632
				_		CTC Leu	_			_						1680
						CCC Pro				_		_		_		1728
						AGC Ser										1776
						GTG Val					_		_	_	_	1824
						GAG Glu 615										1872
-	Ser					GGC Gly										1920
						ACC Thr						_				1968
						ACC Thr										2016
						CAC His										2064
															TAC Tyr	2112
	Thr														CGC Arg 720	2160
ATC	GAG	CTG	AAG	GGC	ATC	GAC	TTC	AAG	GAG	GAC	GGC	AAC	ATC	CTG	GGG	2208

Ile Glu Leu I	Lys Gly Ile 725	Asp Phe Lys	Glu Asp Gly A	Asn Ile Leu ( 735	Gly
His Lys Leu (			CAC AAC GTC T His Asn Val T		
			AAC TTC AAG A Asn Phe Lys I 7		
			GAC CAC TAC C Asp His Tyr G 780		
			CCC GAC AAC C Pro Asp Asn H 795	lis Tyr Leu S	
			AAC GAG AAG C Asn Glu Lys A 810		
Val Leu Leu (			GGG ATC ACT C	· · ·	
GAG CTG TAC A Glu Leu Tyr I 835					2511

### (2) INFORMATION FOR SEQ ID NO:61:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 836 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:

Met Glu Leu Glu Asn Ile Val Ala Asn Thr Val Leu Leu Lys Ala Arg 5 10 Glu Gly Gly Gly Lys Arg Lys Gly Lys Ser Lys Lys Trp Lys Glu 20 25 30 Ile Leu Lys Phe Pro His Ile Ser Gln Cys Glu Asp Leu Arg Arg Thr 35 40 45 Ile Asp Arg Asp Tyr Cys Ser Leu Cys Asp Lys Gln Pro Ile Gly Arg 55 Leu Leu Phe Arg Gln Phe Cys Glu Thr Arg Pro Gly Leu Glu Cys Tyr 70 75 Ile Gln Phe Leu Asp Ser Val Ala Glu Tyr Glu Val Thr Pro Asp Glu 85 90 Lys Leu Gly Glu Lys Gly Lys Glu Ile Met Thr Lys Tyr Leu Thr Pro

Lys Ser Pro Val Phe Ile Ala Gln Val Gly Gln Asp Leu Val Ser Gln Thr Glu Glu Lys Leu Ceu Gln Lys Pro Cys Lys Glu Leu Phe Ser Ala Cys Ala Gln Ser Val His Glu Tyr Leu Arg Gly Glu Pro Phe His Glu Tyr Leu Asp Ser Met Phe Phe Asp Arg Phe Leu Gln Trp Lys Trp Leu Glu Arg Gln Pro Val Thr Lys Asn Thr Phe Arg Gln Tyr Arg Val Leu Gly Lys Gly Gly Phe Gly Glu Val Cys Ala Cys Gln Val Arg Ala Thr Gly Lys Met Tyr Ala Cys Lys Arg Leu Glu Lys Lys Arg Ile Lys Lys Arg Lys Gly Glu Ser Met Ala Leu Asn Glu Lys Gln Ile Leu Glu Lys Val Asn Ser Gln Phe Val Val Asn Leu Ala Tyr Ala Tyr Glu Thr Lys Asp Ala Leu Cys Leu Val Leu Thr Ile Met Asn Gly Gly Asp Leu Lys Phe His Ile Tyr Asn Met Gly Asn Pro Gly Phe Glu Glu Glu Arg Ala Leu Phe Tyr Ala Ala Glu Ile Leu Cys Gly Leu Glu Asp Leu His Arg 295 300 Glu Asn Thr Val Tyr Arg Asp Leu Lys Pro Glu Asn Ile Leu Leu Asp Asp Tyr Gly His Ile Arg Ile Ser Asp Leu Gly Leu Ala Val Lys Ile Pro Glu Gly Asp Leu Ile Arg Gly Arg Val Gly Thr Val Gly Tyr Met Ala Pro Glu Val Leu Asn Asn Gln Arg Tyr Gly Leu Ser Pro Asp Tyr Trp Gly Leu Gly Cys Leu Ile Tyr Glu Met Ile Glu Gly Gln Ser Pro Phe Arg Gly Arg Lys Glu Lys Val Lys Arg Glu Glu Val Asp Arg Arg Val Leu Glu Thr Glu Glu Val Tyr Ser His Lys Phe Ser Glu Glu Ala Lys Ser Ile Cys Lys Met Leu Leu Thr Lys Asp Ala Lys Gln Arg Leu Gly Cys Gln Glu Glu Gly Ala Ala Glu Val Lys Arg His Pro Phe Phe Arg Asn Met Asn Phe Lys Arg Leu Glu Ala Gly Met Leu Asp Pro Pro Phe Val Pro Asp Pro Arg Ala Val Tyr Cys Lys Asp Val Leu Asp Ile Glu Gln Phe Ser Thr Val Lys Gly Val Asn Leu Asp His Thr Asp Asp Asp Phe Tyr Ser Lys Phe Ser Thr Gly Ser Val Ser Ile Pro Trp Gln Asn Glu Met Ile Glu Thr Glu Cys Phe Lys Glu Leu Asn Val Phe Gly Pro Asn Gly Thr Leu Pro Pro Asp Leu Asn Arg Asn His Pro Pro Glu 530 535 Pro Pro Lys Lys Gly Leu Leu Gln Arg Leu Phe Lys Arg Gln His Gln Asn Asn Ser Lys Ser Ser Pro Ser Ser Lys Thr Ser Phe Asn His His

570 Ile Asn Ser Asn His Val Ser Ser Asn Ser Thr Gly Ser Ser Arg Asp 580 585 Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly 600 605 Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys 610 615 620 Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu 630 635 Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro 650 Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr 660 665 Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu 675 680 685 Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr 695 700 Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg 715 705 710 Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly 725 730 His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala 740 745 750 Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn 760 765 Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr 770 775 780 Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser 785 790 795 Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met 805 810 Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp 820 825 830 Glu Leu Tyr Lys 835

### (2) INFORMATION FOR SEQ ID NO:62:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1893 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence

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- (B) LOCATION: 1...1890
- (D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:

ATG AGC AGA AGC CAT GAC AAC AAT TTT TAT AGT GTA GAG ATT GGA
Met Ser Arg Ser Lys Arg Asp Asn Asn Phe Tyr Ser Val Glu Ile Gly
1 5 10 15

96

GAT TCT ACA TTC ACA GTC CTG AAA CGA TAT CAG AAT TTA AAA CCT ATA

Asp	Ser	Thr	Phe 20	Thr	Val	Leu	Lys	Arg 25	Tyr	Gln	Asn	Leu	Lys 30	Pro	Ile	
	TCA Ser												_	_		144
	A AGA 1 Arg 50	Asn														192
	CAT His															240
	CAC His															288
	C CTA															336
	A AAT a Asn			_												384
	TAC Tyr 130	Leu														432
GCT Ala 145	a Gly													_		480
	r GAT r Asp											_		_	_	528
	A ACG y Thr															576
Ala	A CCC	Glu 195	Val	Ile	Leu	Gly	Met 200	Gly	Tyr	Lys	Glu	Asn 205	Val	Asp	Leu	624
Trī	S TCT Ser 210	Val	Gly	Cys	Ile	Met 215	Gly	Glu	Met	Val	Cys 220	His	Lys	Ile	Leu	672
Phe 225		Gly	Arg	Asp	Tyr 230	Ile	Asp	Gln	Trp	Asn 235	Lys	Val	Ile	Glu	Gln 240	720
	r GGA u Gly														Val	768

									TAT Tyr							816
									GCT Ala							864
									TTA Leu							912
									GAA Glu							960
		_							GCA Ala 330							1008
									GAA Glu							1056
									GAC Asp							1104
									CCT Pro							1152
									AGC Ser							1200
			_						CTG Leu 410							1248
									GAG Glu							1296
									ACC Thr							1344
									TAC Tyr							1392
									GAC Asp							1440
CCC	GAA	GGC	TAC	GTC	CAG	GAG	CGC	ACC	ATC	TTC	TTC	AAG	GAC	GAC	GGC	1488

Pro	Glu	Gly	Tyr	Val 485	Gln	Glu	Arg	Thr	Ile 490	Phe	Phe	Lys	Asp	Asp 495	Gly	
					GCC Ala											1536
					AAG Lys											1584
					GAG Glu											1632
					AAG Lys 550											1680
					GGC Gly											1728
					GAC Asp											1776
					GCC Ala											1824
					GAG Glu											1872
		GAG Glu	-	_	AAG Lys 630	TAA										1893

# (2) INFORMATION FOR SEQ ID NO:63:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 630 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:

 Met
 Ser
 Arg
 Ser
 Lys
 Arg
 Asp
 Asp
 Asp
 Phe
 Tyr
 Ser
 Val
 Glu
 Ile
 Gly

 Asp
 Ser
 Thr
 Phe
 Thr
 Val
 Leu
 Lys
 Arg
 Tyr
 Gln
 Asn
 Leu
 Lys
 Pro
 Ile

 Gly
 Ser
 Gly
 Ala
 Gly
 Ile
 Val
 Cys
 Ala
 Ala
 Tyr
 Asp
 Ala
 Ile
 Leu

	35					40					45			
Glu Ar 50		Val	Ala	Ile	Lys 55	Lys	Leu	Ser	Arg	Pro 60	Phe	Gln	Asn	Gln
Thr Hi 65	s Ala	Lys	Arg	Ala 70	Tyr	Arg	Glu	Leu	Val 75	Leu	Met	Lys	Cys	Val 80
Asn Hi	_		85		_			90					95	-
Ser Le		100					105					110		
Ala As	n Leu 115		Gln	Val	Ile	Gln 120	Met	Glu	Leu	Asp	His 125	Glu	Arg	Met
Ser Ty	0		_		135		_	_		140				
Ala Gl 145	y Ile	Ile	His	Arg 150	Asp	Leu	Lys	Pro	Ser 155	Asn	Ile	Val	Val	Lys 160
Ser As			165	_			_	170	_			_	175	
Gly Th		180					185					190		
Ala Pr	195				_	200	_	-	-		205		_	
Trp Se	.0				215					220				
Phe Pr	_	_	_	230		_			235	_				240
Leu Gl	-		245					250	-				255	
Arg Th	_	260					265					270		
Leu Ly	275					280					285			
Leu Ly 29 Asp Al	0				295					300				
305 Ile As				310					315					320
Ile As			325					330					335	
Lys Gl		340					345					350		
Asn Gl	355					360					365			
37 Trp As	0				375					380				
385 Thr Gl				390					395					400
His Ly			405					410					415	
Lys Le		420					425					430		
Trp Pr	435		_			440					445			
45 Arg Ty	0				455					460				
465 Pro Gl				470					475					480
Asn Ty	_	_	485					490					495	
: 1)							_,_			3				<b>-</b>

			500					202					210			
Asn	Arg	Ile 515	Glu	Leu	Lys	Gly	Ile 520	Asp	Phe	Lys	Glu	Asp 525	Gly	Asn	Ile	
Leu	Gly 530	His	Lys	Leu	Glu	Tyr 535	Asn	Tyr	Asn	Ser	His 540	Asn	Val	Tyr	Ile	
Met 545	Ala	Asp	Lys	Gln	Lys 550	Asn	Gly	Ile	Lys	Val 555	Asn	Phe	Lys	Ile	Arg 560	
	Asn	Ile	Glu	Asp 565	Gly	Ser	Val	Gln	Leu 570		Asp	His	Tyr	Gln 575		
Asn	Thr	Pro	Ile 580		Asp	Gly	Pro	Val 585		Leu	Pro	Asp	Asn 590	_	Tyr	
Leu	Ser	Thr 595		Ser	Ala	Leu	Ser 600		Asp	Pro	Asn	Glu 605		Arg	Asp	
His			Leu	Leu	Glu	Phe 615	Val	Thr	Ala	Ala	Gly 620	Ile	Thr	Leu	Gly	
Met 625	610 Asp	Glu	Leu	Tyr	Lys 630	013					020					
023		(2)	TNIE	-OPM	OJO	ਹ ਜਾ∕ਜ ਹ	772 S	חז ו	NO · 6	: <b>4</b> -						
	(.	(A)	LENC	STH:	1821	l bas	se pa									
					ones			9								
		(D)	TOPO	DLOG	Y: 1:	inear	<u>-</u>									
		ii) 1 ix) 1			TYPI	E: cI	AMC									
	•	·			EY: (	~odir	na 54	2011-01	100							
		(B	LO	CATIO	ON: 3	1:	1818	-que.								
					INFO											
					DES											
					CCC Pro										ACA Thr	48
1				5					10					15		
					GAG											96
He	Trp	GIU	20	PIO	Glu	Arg	Tyr	25	ASII	ьeu	Ser	PIO	30	GIY	Ser	
															TTA	144
Gly	Ala	Tyr 35	Gly	Ser	Val		Ala 40	Ala	Phe	Asp	Thr	Lys 45	Thr	Gly	Leu	
															CAT	192
					AAG Lys											192
Arg	Val 50 AAA	Ala AGA	Val ACC	Lys TAC	Lys AGA	Leu 55 GAA	Ser CTG	Arg CGG	Pro TTA	Phe CTT	Gln 60 AAA	Ser	Ile	Ile AAA	His CAT	192 240
Arg	Val 50 AAA	Ala AGA	Val ACC	Lys TAC	Lys	Leu 55 GAA	Ser CTG	Arg CGG	Pro TTA	Phe CTT	Gln 60 AAA	Ser	Ile	Ile AAA	His CAT	
GCG Ala 65	Val 50 AAA Lys	Ala AGA Arg	Val ACC Thr	Lys TAC Tyr	Lys AGA Arg	Leu 55 GAA Glu	Ser CTG Leu	Arg CGG Arg	Pro TTA Leu	Phe CTT Leu 75	Gln 60 AAA Lys	Ser CAT His	Ile ATG Met	Ile AAA Lys	CAT His 80	

	GAA Glu															336
	AAC Asn															384
	CTT Leu 130															432
	ATA Ile															480
	TGT Cys	_														528
	GAA Glu															576
_	ATG Met															624
	GGA Gly 210															672
_	ACA Thr			_												720
	CCA Pro		_													768
	TAT Tyr															816
	TTT Phe	_	_	_												864
	GTA Val 290															912
	GCC Ala															960
GAT	CCT	TAT	GAT	CAG	TCC	TTT	gaa	AGC	AGG	GAC	CTC	CTT	ATA	GAT	GAG	1008

Asp	Pro	Tyr	Asp	Gln 325	Ser	Phe	Glu	Ser	Arg 330	Asp	Leu	Leu	Ile	Asp 335	Glu	
									ATC Ile							1056
									GAT Asp							1104
									GGG Gly				_		_	1152
									AAG Lys							1200
									CTG Leu 410				_			1248
									CCC Pro							1296
									TAC Tyr							1344
									GAA Glu							1392
									TAC Tyr							1440
									CGC Arg 490							1488
									GGG Gly						AAC Asn	1536
															GGC Gly	1584
									AAC Asn						GTG Val	1632
											Ile				CCC Pro 560	1680

GTG CTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC 1728

Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser 575

AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG GAG TTC GTG 1776

Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val 580

ACC GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA 1821

Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 595

## (2) INFORMATION FOR SEQ ID NO:65:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 606 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

Met Ser Gln Glu Arg Pro Thr Phe Tyr Arg Gln Glu Leu Asn Lys Thr 10 Ile Trp Glu Val Pro Glu Arg Tyr Gln Asn Leu Ser Pro Val Gly Ser 25 Gly Ala Tyr Gly Ser Val Cys Ala Ala Phe Asp Thr Lys Thr Gly Leu 35 40 Arg Val Ala Val Lys Lys Leu Ser Arg Pro Phe Gln Ser Ile Ile His 55 60 Ala Lys Arg Thr Tyr Arg Glu Leu Arg Leu Leu Lys His Met Lys His 70 75 Glu Asn Val Ile Gly Leu Leu Asp Val Phe Thr Pro Ala Arg Ser Leu 85 90 Glu Glu Phe Asn Asp Val Tyr Leu Val Thr His Leu Met Gly Ala Asp 105 110 Leu Asn Asn Ile Val Lys Cys Gln Lys Leu Thr Asp Asp His Val Gln 120 125 Phe Leu Ile Tyr Gln Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala 135 140 Asp Ile Ile His Arg Asp Leu Lys Pro Ser Asn Leu Ala Val Asn Glu 150 155 Asp Cys Glu Leu Lys Ile Leu Asp Phe Gly Leu Ala Arg His Thr Asp 170 Asp Glu Met Thr Gly Tyr Val Ala Thr Arg Trp Tyr Arg Ala Pro Glu 180 185 Ile Met Leu Asn Trp Met His Tyr Asn Gln Thr Val Asp Ile Trp Ser 200 205 Val Gly Cys Ile Met Ala Glu Leu Leu Thr Gly Arg Thr Leu Phe Pro 220 215 Gly Thr Asp His Ile Asp Gln Leu Lys Leu Ile Leu Arg Leu Val Gly 230 235 Thr Pro Gly Ala Glu Leu Leu Lys Lys Ile Ser Ser Glu Ser Ala Arg

245 250 Asn Tyr Ile Gln Ser Leu Thr Gln Met Pro Lys Met Asn Phe Ala Asn 260 265 270 Val Phe Ile Gly Ala Asn Pro Leu Ala Val Asp Leu Leu Glu Lys Met 280 285 Leu Val Leu Asp Ser Asp Lys Arg Ile Thr Ala Ala Gln Ala Leu Ala 295 His Ala Tyr Phe Ala Gln Tyr His Asp Pro Asp Asp Glu Pro Val Ala 310 315 Asp Pro Tyr Asp Gln Ser Phe Glu Ser Arg Asp Leu Leu Ile Asp Glu 325 330 335 Trp Lys Ser Leu Thr Tyr Asp Glu Val Ile Ser Phe Val Pro Pro Pro 340 345 350 Leu Asp Gln Glu Glu Met Glu Ser Glu Asp Pro Pro Val Ala Thr Met 360 365 Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val 375 Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu 390 395 Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys 410 Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu 425 Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln 435 440 445 His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg 455 460 Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val 470 475 Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile 485 490 Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn 500 505 Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly 520 Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val 535 540 Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro 550 555 Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser 565 570 Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val 580 585 Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 600

## (2) INFORMATION FOR SEQ ID NO:66:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2913 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...2910

(D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:

			_	_		CAG Gln			_							48	
						GAC Asp										96	
		_			_	GCT Ala		_	_				_	_	_	144	
			_	_	_	TGG Trp 55			_			_	_	_	_	192	
						GGA Gly						_	_			240	
						CCA Pro										288	
						AAA Lys										336	
						GCA Ala								_	_	384	
						CTC Leu 135								_		432	
						AGA Arg										480	
						TGT Cys					_			_		528	
					_	GCT Ala										576	
						CCA Pro										624 ·	
TTA	GCT	CCA	GAA	GTA	CAA	AGC	TCC	GAA	GAA	TAT	ATT	CAG	CTA	TTG	AAG	672	

Leu	Ala 210	Pro	Glu	Val	Gln	Ser 215	Ser	Glu	Glu	Tyr	Ile 220	Gln	Leu	Leu	Lys	
						AGC Ser										720
-						TTC Phe										768
						GTA Val										816
						AGC Ser										864
						TCA Ser 295										912
						CCA Pro										960
_						TCC Ser		_		_	_				_	1008
						GTG Val										1056
						GAT Asp										1104
						GGG Gly 375										1152
						GGC Gly										1200
						CAC His										1248
						AAA Lys										1296
						GAA Glu										1344

	CAT His 450	_			_	_	_	_	_				_			1392
	TTA Leu															1440
	ACA Thr															1488
	TGC Cys											_	_			1536
	CGT Arg															1584
	AAG Lys 530															1632
	GAA Glu															1680
	CGT Arg															1728
	GAC Asp															1776
	TTG Leu		_			_		_		_	_			_	_	1824
	GTG Val 610															1872
	GTT Val															1920
	CGA Arg												_	_		1968
	GCC Ala											_		_	ATA Ile	2016
AAC	AAA	ACA	GCA	ACT	GGC	TAT	GGC	TTT	GCC	GAG	CCC	TAT	AAC	TTG	TAC	2064

Asn	Lys	Thr 675	Ala	Thr	Gly	Tyr	Gly 680	Phe	Ala	Glu	Pro	Туг 685	Asn	Leu	Tyr	
				_	CTG Leu	_				_	_	_				2112
					CTC Leu 710								_		_	2160
					GAT Asp											2208
_					GGG Gly											2256
					AAG Lys							_	_	_		2304
					CTG Leu											2352
					CCC Pro 790									_	_	2400
					TAC Tyr							_			_	2448
					GAA Glu								_	_	_	2496
					TAC Tyr											2544
					CGC Arg		Glu									2592
					GGG Gly 870											2640
					GCC Ala											2688
					AAC Asn											2736

CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC CCC GTG CTG CCC 2784 His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro 920 GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC AAA GAC CCC AAC 2832 Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn 930 935 GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC GTG ACC GCC GCC GGG 2880 Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly 950 ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA 2913 Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 965 970

### (2) INFORMATION FOR SEQ ID NO:67:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 970 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
  (v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:

Met Ser Ala Glu Gly Tyr Gln Tyr Arg Ala Leu Tyr Asp Tyr Lys Lys 1 5 10 Glu Arg Glu Glu Asp Ile Asp Leu His Leu Gly Asp Ile Leu Thr Val 25 Asn Lys Gly Ser Leu Val Ala Leu Gly Phe Ser Asp Gly Gln Glu Ala 40 45 Arg Pro Glu Glu Ile Gly Trp Leu Asn Gly Tyr Asn Glu Thr Thr Gly 55 60 Glu Arg Gly Asp Phe Pro Gly Thr Tyr Val Glu Tyr Ile Gly Arg Lys 70 75 Lys Ile Ser Pro Pro Thr Pro Lys Pro Arg Pro Pro Arg Pro Leu Pro 85 90 Val Ala Pro Gly Ser Ser Lys Thr Glu Ala Asp Val Glu Gln Gln Ala 105 110 100 Leu Thr Leu Pro Asp Leu Ala Glu Gln Phe Ala Pro Pro Asp Ile Ala 120 125 Pro Pro Leu Leu Ile Lys Leu Val Glu Ala Ile Glu Lys Lys Gly Leu 135 140 Glu Cys Ser Thr Leu Tyr Arg Thr Gln Ser Ser Ser Asn Leu Ala Glu 155 150 Leu Arg Gln Leu Leu Asp Cys Asp Thr Pro Ser Val Asp Leu Glu Met 170 Ile Asp Val His Val Leu Ala Asp Ala Phe Lys Arg Tyr Leu Leu Asp 180 185 190 Leu Pro Asn Pro Val Ile Pro Ala Ala Val Tyr Ser Glu Met Ile Ser 200 205 Leu Ala Pro Glu Val Gln Ser Ser Glu Glu Tyr Ile Gln Leu Leu Lys

215 210 Lys Leu Ile Arg Ser Pro Ser Ile Pro His Gln Tyr Trp Leu Thr Leu 230 235 Gln Tyr Leu Leu Lys His Phe Phe Lys Leu Ser Gln Thr Ser Ser Lys 250 Asn Leu Leu Asn Ala Arg Val Leu Ser Glu Ile Phe Ser Pro Met Leu 260 265 Phe Arg Phe Ser Ala Ala Ser Ser Asp Asn Thr Glu Asn Leu Ile Lys 280 275 Val Ile Glu Ile Leu Ile Ser Thr Glu Trp Asn Glu Arg Gln Pro Ala 295 300 Pro Ala Leu Pro Pro Lys Pro Pro Lys Pro Thr Thr Val Ala Asn Asn 305 310 315 Gly Met Asn Asn Asn Met Ser Leu Gln Asn Ala Glu Trp Tyr Trp Gly 325 330 335 Asp Ile Ser Arg Glu Glu Val Asn Glu Lys Leu Arg Asp Thr Ala Asp 340 345 350 Gly Thr Phe Leu Val Arg Asp Ala Ser Thr Lys Met His Gly Asp Tyr 355 360 Thr Leu Thr Leu Arg Lys Gly Gly Asn Asn Lys Leu Ile Lys Ile Phe 380 375 His Arg Asp Gly Lys Tyr Gly Phe Ser Asp Pro Leu Thr Phe Ser Ser 395 390 Val Val Glu Leu Ile Asn His Tyr Arg Asn Glu Ser Leu Ala Gln Tyr 410 405 Asn Pro Lys Leu Asp Val Lys Leu Leu Tyr Pro Val Ser Lys Tyr Gln 425 430 Gln Asp Gln Val Val Lys Glu Asp Asn Ile Glu Ala Val Gly Lys Lys 445 440 Leu His Glu Tyr Asn Thr Gln Phe Gln Glu Lys Ser Arg Glu Tyr Asp 460 455 Arg Leu Tyr Glu Glu Tyr Thr Arg Thr Ser Gln Glu Ile Gln Met Lys 475 470 Arg Thr Ala Ile Glu Ala Phe Asn Glu Thr Ile Lys Ile Phe Glu Glu 485 490 Gln Cys Gln Thr Gln Glu Arg Tyr Ser Lys Glu Tyr Ile Glu Lys Phe 500 505 Lys Arg Glu Gly Asn Glu Lys Glu Ile Gln Arg Ile Met His Asn Tyr 520 525 Asp Lys Leu Lys Ser Arg Ile Ser Glu Ile Ile Asp Ser Arg Arg Arg 530 535 540 Leu Glu Glu Asp Leu Lys Lys Gln Ala Ala Glu Tyr Arg Glu Ile Asp 545 550 555 Lys Arg Met Asn Ser Ile Lys Pro Asp Leu Ile Gln Leu Arg Lys Thr 570 565 Arg Asp Gln Tyr Leu Met Trp Leu Thr Gln Lys Gly Val Arg Gln Lys 580 585 Lys Leu Asn Glu Trp Leu Gly Asn Glu Asn Thr Glu Asp Gln Tyr Ser 600 Leu Val Glu Asp Asp Glu Asp Leu Pro His His Asp Glu Lys Thr Trp 620 615 Asn Val Gly Ser Ser Asn Arg Asn Lys Ala Glu Asn Leu Leu Arg Gly 630 635 Lys Arg Asp Gly Thr Phe Leu Val Arg Glu Ser Ser Lys Gln Gly Cys 645 650 Tyr Ala Cys Ser Val Val Val Asp Gly Glu Val Lys His Cys Val Ile 665 Asn Lys Thr Ala Thr Gly Tyr Gly Phe Ala Glu Pro Tyr Asn Leu Tyr

680 Ser Ser Leu Lys Glu Leu Val Leu His Tyr Gln His Thr Ser Leu Val 695 700 Gln His Asn Asp Ser Leu Asn Val Thr Leu Ala Tyr Pro Val Tyr Ala 710 715 Gln Gln Arg Arg Gln Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly 725 730 735 Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly 740 745 Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Asp 755 760 765 Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys 770 775 780 Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val 785 790 795 Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe 805 810 Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe 820 825 Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly 840 845 Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu 855 860 Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His 870 875 Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn 885 890 Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp 900 905 910 His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro 915 920 925 Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn 930 935 940 Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly 945 950 Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 965

## (2) INFORMATION FOR SEQ ID NO:68:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1788 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: CDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...1785
  - (D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:

ATG GGC AAC GCC GCC GCC GCC AAG AAG GGC AGC GAG CAG GAG AGC GTG
Met Gly Asn Ala Ala Ala Ala Lys Lys Gly Ser Glu Gln Glu Ser Val

1 5 10 15

	GAG Glu															96
	CCC Pro															144
	CTT Leu 50															192
	AGT Ser															240
	AAG Lys															288
	GCC Ala															336
	AAC Asn															384
	TTC Phe 130															432
	TTC Phe															480
	GAC Asp															528
	CAG Gln															576
	GGC Gly															624
	ATT Ile 210															672
	GGA Gly															720
GCT	GAC	CAG	CCT	ATC	CAG	ATC	TAT	GAG	AAA	ATC	GTC	TCT	GGG	AAG	GTG	768

Ala	Asp	Gln	Pro	Ile 245	Gln	Ile	Tyr	Glu	Lys 250	Ile	Val	Ser	Gly	Lys 255	Val	
					TTC Phe											816
					CTA Leu											864
_			_		AAC Asn											912
					AAG Lys 310											960
_		_		_	AGT Ser											1008
					GAG Glu											1056
					GGA Gly											1104
					GGC Gly											1152
					GAT Asp 390											1200
					AAG Lys											1248
				-	GTT Val											1296
					TTC Phe											1344
					TAC Tyr											1392
					GGT Gly 470											1440

						GAT Asp										1488
						AAT Asn										1536
						TTC Phe										1584
						CAT His 535										1632
GGC Gly 545	CCT Pro	GTC Val	CTT Leu	TTA Leu	CCA Pro 550	GAC Asp	AAC Asn	CAT His	TAC Tyr	CTG Leu 555	TCC Ser	ACG Thr	CAA Gln	TCT Ser	GCC Ala 560	1680
						GAA Glu										1728
						ATT Ile								Tyr	AAA Lys	1776
	CAG Gln	GAG Glu 595	TAA													1788

### (2) INFORMATION FOR SEQ ID NO:69:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 595 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:

 Met
 Gly
 Asn
 Ala
 Ala
 Ala
 Ala
 Lys
 Lys
 Lys
 Gly
 Ser
 Glu
 Glu
 Glu
 Ser
 Val

 Lys
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				85					90					95	
Gln	Ala	Val	Asn 100	Phe	Pro	Phe	Leu	Val 105	Lys	Leu	Glu	Phe	Ser 110	Phe	Lys
Asp	Asn	Ser 115	Asn	Leu	Tyr	Met	Val 120	Met	Glu	Tyr	Val	Ala 125	Gly	Gly	Glu
	130				Arg	135		_	_		140				
Arg 145	Phe	Tyr	Ala	Ala	Gln 150	Ile	Val	Leu	Thr	Phe 155	Glu	Tyr	Leu	His	Ser 160
Leu	Asp	Leu	Ile	Tyr 165	Arg	Asp	Leu	Lys	Pro 170	Glu	Asn	Leu	Leu	Ile 175	Asp
Gln	Gln	Gly	Tyr 180	Ile	Gln	Val	Thr	Asp 185	Phe	Gly	Phe	Ala	Lys 190	Arg	Val
Lys	Gly	Arg 195	Thr	Trp	Thr	Leu	Cys 200	Gly	Thr	Pro	Glu	Tyr 205	Leu	Ala	Pro
Glu	Ile 210	Ile	Leu	Ser	Lys	Gly 215	Tyr	Asn	Lys	Ala	Val 220	Asp	Trp	Trp	Ala
Leu 225	Gly	Val	Leu	Ile	Туr 230	Glu	Met	Ala	Ala	Gly 235	Tyr	Pro	Pro	Phe	Phe 240
	_			245	Gln		_		250				_	255	
_			260		Phe			265					270		
		275		_	Leu		280	_		_		285		_	_
	290	_		_	Asn	295	7	_			300			_	
305		-			Lys 310					315					320
-				325	Ser			_	330					335	
			340		Glu			345					350		
_		355		_	Gly		360				_	365			
	370			_	Gly	375					380				
385		_			390					395					400
				405	Lys				410					415	
			420	_	Val			425					430		
		435			Phe		440					445			_
	450				Tyr	455					460				
465		_			Gly 470	_				475					480
-		-		485	Glu	_	_		490				_	495	
_		_	500		His			505					510		
	_	515	_		Asn		520					525			
	530					535					540		_		Asp
Gly	Pro	Val	Leu	Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gin	ser	Ala

545 550 555 Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Ile Leu Leu Glu 570 Phe Val Thr Ala Ala Gly Ile Thr His Gly Met Asp Glu Leu Tyr Lys 585 Pro Gln Glu 595 (2) INFORMATION FOR SEQ ID NO:70: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2181 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...2178 (D) OTHER INFORMATION: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:70: ATG AGC GAC GTG GCT ATT GTG AAG GAG GGT TGG CTG CAC AAA CGA GGG Met Ser Asp Val Ala Ile Val Lys Glu Gly Trp Leu His Lys Arg Gly 10 GAG TAC ATC AAG ACC TGG CGG CCA CGC TAC TTC CTC CTC AAG AAT GAT 96 Glu Tyr Ile Lys Thr Trp Arg Pro Arg Tyr Phe Leu Leu Lys Asn Asp GGC ACC TTC ATT GGC TAC AAG GAG CGG CCG CAG GAT GTG GAC CAA CGT 144 Gly Thr Phe Ile Gly Tyr Lys Glu Arg Pro Gln Asp Val Asp Gln Arg 40 GAG GCT CCC CTC AAC AAC TTC TCT GTG GCG CAG TGC CAG CTG ATG AAG 192 Glu Ala Pro Leu Asn Asn Phe Ser Val Ala Gln Cys Gln Leu Met Lys 55 ACG GAG CGG CCC CGG CCC AAC ACC TTC ATC ATC CGC TGC CTG CAG TGG 240 Thr Glu Arg Pro Arg Pro Asn Thr Phe Ile Ile Arg Cys Leu Gln Trp ACC ACT GTC ATC GAA CGC ACC TTC CAT GTG GAG ACT CCT GAG GAG CGG 288 Thr Thr Val Ile Glu Arg Thr Phe His Val Glu Thr Pro Glu Glu Arg 85 GAG GAG TGG ACA ACC GCC ATC CAG ACT GTG GCT GAC GGC CTC AAG AAG Glu Glu Trp Thr Thr Ala Ile Gln Thr Val Ala Asp Gly Leu Lys Lys 100 105 CAG GAG GAG GAG ATG GAC TTC CGG TCG GGC TCA CCC AGT GAC AAC 384 Gln Glu Glu Glu Met Asp Phe Arg Ser Gly Ser Pro Ser Asp Asn

120

TCA GGG GCT GAA GAG ATG GAG GTG TCC CTG GCC AAG CCC AAG CAC CGC

432

Ser	Gly 130	Ala	Glu	Glu	Met	Glu 135	Val	Ser	Leu	Ala	Lys 140	Pro	Lys	His	Arg	
	ACC Thr															480
	GGC Gly															528
	ATG Met															576
	CAC His															624
	CTC Leu 210															672
	GTC Val															720
	GAA Glu															768
	GTG Val															816
	GAC Asp															864
	ATC Ile 290															912
	ATG Met															960
	GAG Glu															1008
	GTC Val															1056
	CAT His															1104

									TCC Ser							1152	
									GGG Gly							1200	
									GGT Gly 410							1248	
									AAG Lys							1296	
									TTC Phe							1344	
									ATG Met	_		_			_	1392	
									TAC Tyr							1440	
									AGC Ser 490							1488	
							_		CTG Leu				_		_	1536	
								Gly	GAG Glu							1584	
									ACC Thr							1632	
				_					TAC Tyr	_	_	_		_		1680	
							_		GAC Asp 570							1728	
									ATC Ile							1776	
AAC	TAC	AAG	ACC	CGC	GCC	GAG	GTG	AAG	TTC	GAG	GGC	GAC	ACC	CTG	GTG	1824	

Asn	Tyr	Lys 595	Thr	Arg	Ala	Glu	Val 600	Lys	Phe	Glu	Gly	Asp 605	Thr	Leu	Val	•
					AAG Lys											1872
					GAG Glu 630											1920
		-			AAG Lys						-			_		1968
					GGC Gly											2016
					GAC Asp											2064
					GCC Ala											2112
					GAG Glu 710											2160
	GAC Asp				AAG Lys	TAA										2181

#### (2) INFORMATION FOR SEQ ID NO:71:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 726 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:71:

 Met
 Ser
 Asp
 Val
 Ala
 Ile
 Val
 Lys
 Glu
 Trp
 Leu
 His
 Lys
 Arg
 Gly

 Glu
 Tyr
 Ile
 Lys
 Thr
 Trp
 Arg
 Pro
 Arg
 Tyr
 Phe
 Leu
 Lys
 Asn
 Asp

 Gly
 Thr
 Phe
 Ile
 Gly
 Tyr
 Lys
 Glu
 Arg
 Pro
 Gln
 Asp
 Val
 Asp
 Gln
 Arg

 Glu
 Ala
 Pro
 Leu
 Asn
 Asn
 Phe
 Ser
 Val
 Ala
 Gln
 Leu
 Met
 Lys

 50
 Ile
 Fro
 Asn
 Phe
 Ile
 Ile
 Arg
 Cys
 Leu
 Gln
 Trp

Thr Thr Val Ile Glu Arg Thr Phe His Val Glu Thr Pro Glu Glu Arg Glu Glu Trp Thr Thr Ala Ile Gln Thr Val Ala Asp Gly Leu Lys Lys Gln Glu Glu Glu Met Asp Phe Arg Ser Gly Ser Pro Ser Asp Asn Ser Gly Ala Glu Glu Met Glu Val Ser Leu Ala Lys Pro Lys His Arg Val Thr Met Asn Glu Phe Glu Tyr Leu Lys Leu Leu Gly Lys Gly Thr Phe Gly Lys Val Ile Leu Val Lys Glu Lys Ala Thr Gly Arg Tyr Tyr Ala Met Lys Ile Leu Lys Lys Glu Val Ile Val Ala Lys Asp Glu Val Ala His Thr Leu Thr Glu Asn Arg Val Leu Gln Asn Ser Arg His Pro Phe Leu Thr Ala Leu Lys Tyr Ser Phe Gln Thr His Asp Arg Leu Cys Phe Val Met Glu Tyr Ala Asn Gly Gly Glu Leu Phe Phe His Leu Ser Arg Glu Arg Val Phe Ser Glu Asp Arg Ala Arg Phe Tyr Gly Ala Glu 245 250 Ile Val Ser Ala Leu Asp Tyr Leu His Ser Glu Lys Asn Val Val Tyr Arg Asp Leu Lys Leu Glu Asn Leu Met Leu Asp Lys Asp Gly His Ile 280 285 Lys Ile Thr Asp Phe Gly Leu Cys Lys Glu Gly Ile Lys Asp Gly Ala Thr Met Lys Thr Phe Cys Gly Thr Pro Glu Tyr Leu Ala Pro Glu Val Leu Glu Asp Asn Asp Tyr Gly Arg Ala Val Asp Trp Trp Gly Leu Gly Val Val Met Tyr Glu Met Met Cys Gly Arg Leu Pro Phe Tyr Asn Gln Asp His Glu Lys Leu Phe Glu Leu Ile Leu Met Glu Glu Ile Arg Phe Pro Arg Thr Leu Gly Pro Glu Ala Lys Ser Leu Leu Ser Gly Leu Leu Lys Lys Asp Pro Lys Gln Arg Leu Gly Gly Gly Ser Glu Asp Ala Lys Glu Ile Met Gln His Arg Phe Phe Ala Gly Ile Val Trp Gln His Val Tyr Glu Lys Lys Leu Ser Pro Pro Phe Lys Pro Gln Val Thr Ser Glu Thr Asp Thr Arg Tyr Phe Asp Glu Glu Phe Thr Ala Gln Met Ile Thr Ile Thr Pro Pro Asp Gln Asp Asp Ser Met Glu Cys Val Asp Ser Glu Arg Arg Pro His Phe Pro Gln Phe Ser Tyr Ser Ala Ser Ser Thr Ala Ser Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro

	530					535					540					
Trp 545	Pro	Thr	Leu	Val	Thr 550	Thr	Leu	Thr	Tyr	Gly 555	Val	Gln	Cys	Phe	Ser 560	
Arg	Tyr	Pro	Asp	His 565	Met	Lys	Gln	His	Asp 570	Phe	Phe	Lys	Ser	Ala 575		
Pro	Glu	Gly	Tyr 580	Val	Gln	Glu	Arg	Thr 585	Ile	Phe	Phe	Lys	Asp 590	Asp	Gly	
Asn	Tyr	Lys 595	Thr	Arg	Ala	Glu	Val 600	Lys	Phe	Glu	Gly	Asp 605	Thr	Leu	Val	
Asn	Arg 610	Ile	Glu	Leu	Lys	Gly 615	Ile	Asp	Phe	Lys	Glu 620	Asp	Gly	Asn	Ile	
Leu 625	Gly	His	Lys	Leu	Glu 630	Tyr	Asn	Tyr	Asn	Ser 635	His	Asn	Val	Tyr	Ile 640	
Met	Ala	Asp	Lys	Gln 645	Lys	Asn	Gly	Ile	Lys 650	Val	Asn	Phe	Lys	Ile 655	Arg	
His	Asn	Ile	Glu 660	Asp	Gly	Ser	Val	Gln 665	Leu	Ala	Asp	His	Tyr 670	Gln	Gln	
Asn	Thr	Pro 675	Ile	Gly	Asp	Gly	Pro 680	Val	Leu	Leu	Pro	Asp 685	Asn	His	Tyr	
Leu	Ser 690	Thr	Gln	Ser	Ala	Leu 695	Ser	Lys	Asp	Pro	Asn 700	Glu	Lys	Arg	Asp	
His 705	Met	Val	Leu	Leu	Glu 710	Phe	Val	Thr	Ala	Ala 715	Gly	Ile	Thr	Leu	Gly 720	
Met	Asp	Glu	Leu	Тут 725	Lys											
		(2)	INI	FORM	ATIO	v FOI	RSEX	O ID	NO:	72:						
	( :				CHARA											
	,	(A)	LEN	TH:	275	l bas	se pa									
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				ER I	ENFO											
	()	ci) S	EQUE	ENCE	DESC	RIP	LTON:	: SEÇ	) ID	NO:	72:					
	GCT	GAC	GTT	TAC	CCG	GCC	AAC	GAC	TCC	ACG	GCG			GAC		48
	GCT	GAC	GTT	TAC	CCG	GCC	AAC	GAC	TCC	ACG	GCG			GAC Asp 15		48
Met 1 GCC	GCT Ala AAC	GAC Asp CGC	GTT Val TTC	TAC Tyr 5 GCC	CCG Pro	GCC Ala	AAC Asn GGG	GAC Asp GCG	TCC Ser 10 CTG	ACG Thr AGG	GCG Ala CAG	Ser AAG	Gln AAC	Asp 15 GTG	Val CAT	48 96
Met 1 GCC	GCT Ala AAC	GAC Asp CGC	GTT Val TTC	TAC Tyr 5 GCC	CCG Pro	GCC Ala	AAC Asn GGG	GAC Asp GCG	TCC Ser 10 CTG	ACG Thr AGG	GCG Ala CAG	Ser AAG	Gln AAC	Asp 15	Val CAT	
Met 1 GCC Ala	GCT Ala AAC Asn	GAC Asp CGC Arg	GTT Val TTC Phe 20	TAC Tyr 5 GCC Ala	CCG Pro CGC Arg	GCC Ala AAA Lys	AAC Asn GGG Gly	GAC Asp GCG Ala 25	TCC Ser 10 CTG Leu	ACG Thr AGG Arg	GCG Ala CAG Gln	Ser AAG Lys	Gln AAC Asn 30	Asp 15 GTG	Val CAT His	
Met 1 GCC Ala GAG	GCT Ala AAC Asn	GAC Asp CGC Arg	GTT Val TTC Phe 20 GAC	TAC Tyr 5 GCC Ala	CCG Pro CGC Arg	GCC Ala AAA Lys	AAC Asn GGG Gly	GAC Asp GCG Ala 25 GCC	TCC Ser 10 CTG Leu	ACG Thr AGG Arg	GCG Ala CAG Gln	Ser AAG Lys AAG	AAC Asn 30 CAA	Asp 15 GTG Val	Val CAT His	96

Phe Cys Ser His Cys Thr Asp Phe Ile Trp Gly Phe Gly Lys Gln Gly

TTC CAPPhe Gl:															240
TTC GT Phe Va															288
GAC CC Asp Pr															336
ACC TT		Asp													384
GGG ATV Gly Me 13	Lys														432
ATC AAI Ile Asi 145															480
CGG AT															528
GTA CG. Val Ar															576
GAT CC															624
AAA CAG Lys Gl: 21	ı Lys														672
GAG TCC Glu Se: 225															720
TCT GT															768
GGA TCC			_												816
GGA TGG															864
CCC AT	CCA	GAA	GGA	GAT	GAA	GAA	GGC	AAC	ATG	GAA	CTC	AGG	CAG	AAG	912

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Pro	Ile 290	Pro	Glu	Gly	Asp	Glu 295	Glu	Gly	Asn	Met	Glu 300	Leu	Arg	Gln	Lys	
					CTA Leu 310											960
					CAA Gln								_			1008
					CTC Leu											1056
					AGG Arg											1104
					GTG Val											1152
					GTG Val 390									_		1200
	-				TGC Cys										_	1248
					GGC Gly											1296
					CCA Pro											1344
					CTT Leu										CTG Leu	1392
					ATG Met 470											1440
					AAG Lys											1488
					CCG Pro											1536
												_	_		CTG Leu	1584

					GGG Gly											1632
					ATA Ile 550											1680
					GTC Val											1728
					GGC Gly											1776
					AGG Arg											1824
					TTC Phe											1872
					TTC Phe 630											1920
					GCT Ala											1968
					CAG Gln											2016
					AAA Lys											2064
					GAT Asp											2112
					GGT Gly 710										AAA Lys 720	2160
					GGG Gly										Val	2208
					GGT Gly									Asp	CAT	2256
ATG	AAA	CAG	CAT	GAC	TTT	TTC	AAG	AGT	GCC	ATG	CCC	GAA	GGT	TAT	GTA	2304

Met	Lys	Gln 755	His	Asp	Phe	Phe	Lys 760	Ser	Ala	Met	Pro	Glu 765	Gly	Туг	Val	
									GAC Asp							2352
									CTT Leu							2400
-									AAC Asn 810							2448
									TAC Tyr							2496
									ATT Ile							2544
		_							CAA Gln							2592
									CAT His							2640
									AGA Arg 890							2688
									CAT His							2736
-		CAG Gln 915		TAA												2751

(2) INFORMATION FOR SEQ ID NO:73:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 916 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:

Met Ala Asp Val Tyr Pro Ala Asn Asp Ser Thr Ala Ser Gln Asp Val

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Ala Asn Arg Phe Ala Arg Lys Gly Ala Leu Arg Gln Lys Asn Val His
                            25
Glu Val Lys Asp His Lys Phe Ile Ala Arg Phe Phe Lys Gln Pro Thr
Phe Cys Ser His Cys Thr Asp Phe Ile Trp Gly Phe Gly Lys Gln Gly
            55
Phe Gln Cys Gln Val Cys Cys Phe Val Val His Lys Arg Cys His Glu
                 70
                                   75
Phe Val Thr Phe Ser Cys Pro Gly Ala Asp Lys Gly Pro Asp Thr Asp
              85
                                 90
Asp Pro Arg Ser Lys His Lys Phe Lys Ile His Thr Tyr Gly Ser Pro
                             105
                                               110
Thr Phe Cys Asp His Cys Gly Ser Leu Leu Tyr Gly Leu Ile His Gln
                          120
Gly Met Lys Cys Asp Thr Cys Asp Met Asn Val His Asn Gln Cys Val
                      135
                                         140
Ile Asn Asp Pro Ser Leu Cys Gly Met Asp His Thr Glu Lys Arg Gly
                 150
                                     155
Arg Ile Tyr Leu Lys Ala Glu Val Thr Asp Glu Lys Leu His Val Thr
               165
                                 170
Val Arg Asp Ala Lys Asn Leu Ile Pro Met Asp Pro Asn Gly Leu Ser
           180
                             185
                                                190
Asp Pro Tyr Val Lys Leu Lys Leu Ile Pro Asp Pro Lys Asn Glu Ser
                         200
                                     205
Lys Gln Lys Thr Lys Thr Ile Arg Ser Asn Leu Asn Pro Gln Trp Asn
                     215
                                         220
Glu Ser Phe Thr Phe Lys Leu Lys Pro Ser Asp Lys Asp Arg Arg Leu
                  230
                                      235
Ser Val Glu Ile Trp Asp Trp Asp Arg Thr Thr Arg Asn Asp Phe Met
              245
                                  250
Gly Ser Leu Ser Phe Gly Val Ser Glu Leu Met Lys Met Pro Ala Ser
          260
                             265
Gly Trp Tyr Lys Ala His Asn Gln Glu Glu Glu Glu Tyr Tyr Asn Val
               280
                                          285
Pro Ile Pro Glu Gly Asp Glu Glu Gly Asn Met Glu Leu Arg Gln Lys
                     295
                                        300
Phe Glu Lys Ala Lys Leu Gly Pro Val Gly Asn Lys Val Ile Ser Pro
                 310
                                     315
Ser Glu Asp Arg Lys Gln Pro Ser Asn Asn Leu Asp Arg Val Lys Leu
               325
                                  330
Thr Asp Phe Asn Phe Leu Met Val Leu Gly Lys Gly Ser Phe Gly Lys
           340
                              345
Val Met Leu Ala Asp Arg Lys Gly Thr Glu Glu Leu Tyr Ala Ile Lys
                          360
                                             365
Ile Leu Lys Lys Asp Val Val Ile Gln Asp Asp Val Glu Cys Thr
                      375
                                         380
Met Val Glu Lys Arg Val Leu Ala Leu Leu Asp Lys Pro Pro Phe Leu
                  390
                                      395
Thr Gln Leu His Ser Cys Phe Gln Thr Val Asp Arg Leu Tyr Phe Val
              405
                                 410
Met Glu Tyr Val Asn Gly Gly Asp Leu Met Tyr His Ile Gln Gln Val
           420
                             425
Gly Lys Phe Lys Glu Pro Gln Ala Val Phe Tyr Ala Ala Glu Ile Ser
                          440
                                             445
Ile Gly Leu Phe Phe Leu His Lys Arg Gly Ile Ile Tyr Arg Asp Leu
            455
                                         460
Lys Leu Asn Asn Val Met Leu Asn Ser Glu Gly His Ile Lys Ile Ala
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Asp Phe Gly Met Cys Lys Glu His Met Met Asp Gly Val Thr Thr Arg Thr Phe Cys Gly Thr Pro Asp Tyr Ile Ala Pro Glu Ile Ile Ala Tyr Gln Pro Tyr Gly Lys Ser Val Asp Trp Trp Ala Tyr Gly Val Leu Leu Tyr Glu Met Leu Ala Gly Gln Pro Pro Phe Asp Gly Glu Asp Glu Asp Glu Leu Phe Gln Ser Ile Met Glu His Asn Val Ser Tyr Pro Lys Ser Leu Ser Lys Glu Ala Val Ser Ile Cys Lys Gly Leu Met Thr Lys Gln 565 570 Pro Ala Lys Arg Leu Gly Cys Gly Pro Glu Gly Glu Arg Asp Val Arg 580 585 Glu His Ala Phe Phe Arg Arg Ile Asp Trp Glu Lys Leu Glu Asn Arg Glu Ile Gln Pro Pro Phe Lys Pro Lys Val Cys Gly Lys Gly Ala Glu Asn Phe Asp Lys Phe Phe Thr Arg Gly Gln Pro Val Leu Thr Pro Pro Asp Gln Leu Val Ile Ala Asn Ile Asp Gln Ser Asp Phe Glu Gly Phe Ser Tyr Val Asn Pro Gln Phe Val His Pro Ile Leu Gln Ser Ala Val Gly Arg Ala Met Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly Gln Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His 740 745 Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Tyr Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Met Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Pro Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Lys Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Ile Leu Leu 885 890 Glu Phe Val Thr Ala Ala Gly Ile Thr His Gly Met Asp Glu Leu Tyr Lys Pro Gln Glu 

(2) INFORMATION FOR SEQ ID NO:74:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2157 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2154
  - (D) OTHER INFORMATION:

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:74:

														CTG Leu 15		48
														GGA Gly		96
														AGT Ser		144
														AAA Lys		192
														AGC Ser		240
														CAG Gln 95		288
														CTT Leu		336
														ATA Ile		384
														CTC Leu		432
														GAA Glu		480
TGT	GTA	AAC ·	CCT	TAC	CAC	TAT	CAG	AGA	GTT	GAG	ACA	CCA	GTT	TTG	CCT	528

Cys	Val	Asn	Pro	Tyr 165	His	Tyr	Gln	Arg	Val 170	Glu	Thr	Pro	Val	Leu 175	Pro	
									ATC Ile							576
									GAA Glu							624
									CCA Pro							672
									GAC Asp							720
									TCT Ser 250							768
									GTT Val							816
									TTA Leu				_			864
									ACT Thr							912
									GGT Gly							960
									AGG Arg 330							1008
									TTT Phe							1056
									TGT Cys							1104
									CCA Pro							1152
								-	CTG Leu							1200

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GGT	TTT	GAA	GCC	GTC	TAT	CAG	CTA	ACT	AGA	ATG	TGC	ACC	ATA	AGA	ATG	1248
Gly	Phe	Glu	Ala	Val 405	Tyr	Gln	Leu	Thr	Arg 410	Met	Cys	Thr	Ile	Arg 415	Met	
				#0J					410					417		
				_					TAC			_	_	_	_	1296
Ser	Phe	Val	Lys 420	GIY	Trp	СŢУ	Ala	G1u 425	Tyr	Arg	Arg	Gin	430	vaı	Thr	
			-20													
									CTG							1344
Ser	Thr	Pro 435	Cys	Trp	He	GIu	Leu 440	His	Leu	Asn	GIY	445	Leu	GIN	urp	
									TCC			_				1392
ьeu	450	гÀг	vaı	Leu	Thr	455	Mec	GIĀ	Ser	PIO	460	vaı	ALG	Cys	Ser	
									GAT Asp			_	_			1440
465	nec	SGI	тър	val	470	A. y	viq	wy	പാവ	475	110	VAI	ALG		480	
_		<b>.</b>			<b></b> -		-				-			am-	omc.	1.400
									GGG Gly		_		_		_	1488
		_, _	1	485					490					495	_	
CAC	Curc	GAC	ccc	GAC	ርመአ	אמ	CCC	CAC	AAG	אואט	ልርር	בינוט	ጥርር	GGC	GAG	1536
									Lys			_				1330
			500					505					510			
GGC	GAG	GGC	GAT	GCC	ACC	TAC	GGC	AAG	CTG	ACC	CTG	AAG	TTC	ATC	TGC	1584
									Leu			Lys	_	_		
		515					520					525				
ACC	ACC	GGC	AAG	CTG	CCC	GTG	CCC	TGG	CCC	ACC	CTC	GTG	ACC	ACC	CTG	1632
Thr		Gly	Lys	Leu	Pro		Pro	Trp	Pro	Thr		Val	Thr	Thr	Leu	
	530					535					540					
									TAC							1680
	Tyr	Gly	Val	Gln	Cys 550	Phe	Ser	Arg	Tyr	Pro 555	Asp	His	Met	Lys	Gln 560	
545					230					درر					300	
									GAA							1728
His	Asp	Phe	Phe	Lys 565					Glu 570				GIn	Glu 575	Arg	
				555					2,0							
									TAC					_	_	1776
Thr	TTE	rne	580	ьys	ASP	ASP	стĀ	Asn 585	Tyr	гĀŞ	THE	Arg	590	GIU	vaı	
									CGC Arg					_	_	1824
гÀг	rne	595	GTĀ	wsb	TILL	neu	600	MSII	ALG	116	GIU	605	пуъ	GIY	116	
														<b></b> -		1000
									GGG Gly							1872
קביי	610	<b>-</b> y 5			1	615			3		620			-1-		
m» ~	220	200	CAC	አአጣ	CIT/C	m » m	y m-c	אווער	ccc	CNC	አአጥ	CNC	አአር	א א	ccc	1920
TAC	AAC	AGC	CAC	AAC	GTC	TAT	ATC	ATG	GCC	GAC	AAG	CAG	AAG	AAC	GGC	1320

Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly 625 630 635 ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC GTG Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val 645 650 CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC CCC 2016 Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro 665 660 GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC 2064 Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser 680 AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG GAG TTC GTG 2112 Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val 700 690 695 ACC GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA 2157 Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 715 710

#### (2) INFORMATION FOR SEQ ID NO:75:

- (i) SEQUENCE CHARACTERISTICS:
- . (A) LENGTH: 718 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:75:

Met Ser Ser Ile Leu Pro Phe Thr Pro Pro Val Val Lys Arg Leu Leu 10 Gly Trp Lys Lys Ser Ala Gly Gly Ser Gly Gly Ala Gly Gly Glu 20 25 30 Gln Asn Gly Gln Glu Glu Lys Trp Cys Glu Lys Ala Val Lys Ser Leu 40 Val Lys Lys Leu Lys Lys Thr Gly Arg Leu Asp Glu Leu Glu Lys Ala 55 Ile Thr Thr Gln Asn Cys Asn Thr Lys Cys Val Thr Ile Pro Ser Thr 75 70 Cys Ser Glu Ile Trp Gly Leu Ser Thr Pro Asn Thr Ile Asp Gln Trp 85 90 Asp Thr Thr Gly Leu Tyr Ser Phe Ser Glu Gln Thr Arg Ser Leu Asp 100 105 Gly Arg Leu Gln Val Ser His Arg Lys Gly Leu Pro His Val Ile Tyr 120 125 Cys Arg Leu Trp Arg Trp Pro Asp Leu His Ser His His Glu Leu Lys 135 140 Ala Ile Glu Asn Cys Glu Tyr Ala Phe Asn Leu Lys Lys Asp Glu Val 150 155 Cys Val Asn Pro Tyr His Tyr Gln Arg Val Glu Thr Pro Val Leu Pro the state of the s

Pro Val Leu Val Pro Arg His Thr Glu Ile Leu Thr Glu Leu Pro Pro Leu Asp Asp Tyr Thr His Ser Ile Pro Glu Asn Thr Asn Phe Pro Ala Gly Ile Glu Pro Gln Ser Asn Tyr Ile Pro Glu Thr Pro Pro Pro Gly Tyr Ile Ser Glu Asp Gly Glu Thr Ser Asp Gln Gln Leu Asn Gln Ser Met Asp Thr Gly Ser Pro Ala Glu Leu Ser Pro Thr Thr Leu Ser Pro Val Asn His Ser Leu Asp Leu Gln Pro Val Thr Tyr Ser Glu Pro Ala Phe Trp Cys Ser Ile Ala Tyr Tyr Glu Leu Asn Gln Arg Val Gly Glu Thr Phe His Ala Ser Gln Pro Ser Leu Thr Val Asp Gly Phe Thr Asp Pro Ser Asn Ser Glu Arg Phe Cys Leu Gly Leu Leu Ser Asn Val Asn , 310 Arg Asn Ala Thr Val Glu Met Thr Arg Arg His Ile Gly Arg Gly Val Arg Leu Tyr Tyr Ile Gly Gly Glu Val Phe Ala Glu Cys Leu Ser Asp Ser Ala Ile Phe Val Gln Ser Pro Asn Cys Asn Gln Arg Tyr Gly Trp His Pro Ala Thr Val Cys Lys Ile Pro Pro Gly Cys Asn Leu Lys Ile Phe Asn Asn Gln Glu Phe Ala Ala Leu Leu Ala Gln Ser Val Asn Gln Gly Phe Glu Ala Val Tyr Gln Leu Thr Arg Met Cys Thr Ile Arg Met Ser Phe Val Lys Gly Trp Gly Ala Glu Tyr Arg Arg Gln Thr Val Thr Ser Thr Pro Cys Trp Ile Glu Leu His Leu Asn Gly Pro Leu Gln Trp Leu Asp Lys Val Leu Thr Gln Met Gly Ser Pro Ser Val Arg Cys Ser Ser Met Ser Trp Val Pro Arg Ala Arg Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly

625					630					635					640	
Ile	Lys	Val	Asn	Phe 645	Lys	Ile	Arg	His	Asn 650	Ile	Glu	Asp	Gly	Ser 655	Val	
Gln	Leu	Ala	Asp 660	His	Tyr	Gln	Gln	Asn 665	Thr	Pro	Ile	Gly	Asp 670	Gly	Pro	
Val	Leu	Leu 675	Pro	Asp	Asn	His	Tyr 680	Leu	Ser	Thr	Gln	Ser 685	Ala	Leu	Ser	
Lys	Asp 690	Pro	Asn	Glu	Lys	Arg 695	Asp	His	Met	Val	Leu 700	Leu	Glu	Phe	Val	
Thr 705	Ala	Ala	Gly	Ile	Thr 710	Leu	Gly	Met	Asp	Glu 715	Leu	Tyr	Lys			
		(2)	) INI	FORM	ATION	v FOI	R SEX	Q ID	NO:	76:						
		(A) (B) (C) (D)	LENG TYPI STRA TOPG	ETH: E: nu ANDEI OLOGY	CHARA 2397 IClei ONESS 7: li	7 bas ic ac S: si inear	se pa cid ingle	airs								
	-		TEAT		TYPE	s: C1	JINA									
		(B)	LO	CATIO	EY: C ON: 1	١2	2394	equer	nce							
	()	ci) s	SEQUI	ENCE	DESC	CRIP	rion	: SEX	O ID	NO:	76:					
ATG	GAC	ААТ	ATG	TCT	ATT	ACG	AAT	ACA	CCA	ACA	AGT	ААТ	GAT	GCC	TGT	48
	Asp															
CTG	AGC	ATT	GTG	САТ	AGT	TTG	ATG	TGC	CAT	AGA	CAA	GGT	GGA	GAG	AGT	96
Leu	Ser	Ile	Val 20	His	Ser	Leu	Met	Cys 25	His	Arg	Gln	Gly	Gly 30	Glu	Ser	
GAA	ACA	TTT	GCA	AAA	AGA	GCA	ATT	GAA	AGT	TTG	GTA	AAG	AAG	CTG	AAG	144
Glu	Thr	Phe 35	Ala	Lys	Arg	Ala	Ile 40	Glu	Ser	Leu	Val	Lys 45	Lys	Leu	Lys	
	AAA															192
Glu	Lys 50	Lys	Asp	Glu	Leu	Asp 55	Ser	Leu	Ile	Thr	Ala 60	Ile	Thr	Thr	Asn	
	GCT															240
Gly 65	Ala	His	Pro	Ser	Lys 70	Cys	Val	Thr	Ile	G1n 75	Arg	Thr	Leu	Asp	Gly 80	
	CTT															288
Arg	Leu	Gln	Val	Ala 85	Gly	Arg	Lys	Gly	Phe 90	Pro	His	Val	Ile	Tyr 95	Ala	
	CTC															336
Arg	Leu	Trp	Arg 100	urp	Pro	Asp	Leu	His 105	ьуѕ	Asn	GIU	Leu	Lys 110	HIS	vaı	
AAA	TAT	TGT	CAG	ТАТ	GCG	TTT	GAC	TTA	AAA	TGT	GAT	AGT	GTC	TGT	GTG	384

Ly ·	s Tyr	Суs 115	Gln	Tyr	Ala	Phe	Asp 120	Leu	Lys	Cys	Asp	Ser 125	Val	Cys	Val	
	T CCA n Pro 130	Tyr														432
	A TTA y Leu 5															480
	а тат и Туг												_	_	_	528
	T TCA s Ser						_						_			576
	G ACA u Thr															624
	C AGC ir Ser 210	Thr														672
	T GCC O Ala															720
	CA TCA .a Ser															768
	CA GCT TO Ala															816
	CT GCA															864
	AG CAC In His 290	His					Pro									912
_	AC AA7 is Asr )5															960
	AG TAT Lu Tyr														Gly	1008
	AG ACA lu Thi													Asp		1056

					GGA Gly	_			_							1104
					GAA Glu											1152
					GAA Glu 390											1200
					GCG Ala											1248
					CCT Pro											1296
				-	TTT Phe											1344
					GCA Ala							_	_	_		1392
					CCT Pro 470											1440
					GCT Ala						_			_		1488
					AGG Arg										GAT Asp	1536
					ATC Ile											1584
					CAG Gln										CCG Pro	1632
															ATG Met 560	1680
															GTC Val	1728
GAG	CTG	GAC	GGC	GAC	GTA	AAC	GGC	CAC	AAG	TTC	AGC	GTG	TCC	GGC	GAG	1776

Glu	Leu	Asp	Gly 580	Asp	Val	Asn	Gly	His 585	Lys	Phe	Ser	Val	Ser 590	Gly	Glu	
				_	ACC Thr											1824
					CCC Pro											1872
					TGC Cys 630											1920
					TCC Ser											1968
					GAC Asp											2016
					ACC Thr						_			_	_	2064
					GGC Gly								_			2112
					GTC Val 710											2160
					AAG Lys			_		_	_		_		_	2208
					TAC Tyr											2256
					AAC Asn	His	Tyr									2304
		755					760									
	GAC	ccc			AAG Lys	CGC	GAT					CTG				2352

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 798 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:77:

Met Asp Asn Met Ser Ile Thr Asn Thr Pro Thr Ser Asn Asp Ala Cys 1 5 10 Leu Ser Ile Val His Ser Leu Met Cys His Arg Gln Gly Glu Ser 20 25 Glu Thr Phe Ala Lys Arg Ala Ile Glu Ser Leu Val Lys Lys Leu Lys 40 Glu Lys Lys Asp Glu Leu Asp Ser Leu Ile Thr Ala Ile Thr Thr Asn 55 60 Gly Ala His Pro Ser Lys Cys Val Thr Ile Gln Arg Thr Leu Asp Gly 70 75 Arg Leu Gln Val Ala Gly Arg Lys Gly Phe Pro His Val Ile Tyr Ala 90 Arg Leu Trp Arg Trp Pro Asp Leu His Lys Asn Glu Leu Lys His Val 100 105 110 Lys Tyr Cys Gln Tyr Ala Phe Asp Leu Lys Cys Asp Ser Val Cys Val 115 120 125 Asn Pro Tyr His Tyr Glu Arg Val Val Ser Pro Gly Ile Asp Leu Ser 135 140 Gly Leu Thr Leu Gln Ser Asn Ala Pro Ser Ser Met Met Val Lys Asp 145 150 155 Glu Tyr Val His Asp Phe Glu Gly Gln Pro Ser Leu Ser Thr Glu Gly 165 170 His Ser Ile Gln Thr Ile Gln His Pro Pro Ser Asn Arg Ala Ser Thr 180 185 190 Glu Thr Tyr Ser Thr Pro Ala Leu Leu Ala Pro Ser Glu Ser Asn Ala 195 200 205 Thr Ser Thr Ala Asn Phe Pro Asn Ile Pro Val Ala Ser Thr Ser Gln 210 215 220 Pro Ala Ser Ile Leu Gly Gly Ser His Ser Glu Gly Leu Leu Gln Ile 225 230 235 Ala Ser Gly Pro Gln Pro Gly Gln Gln Asn Gly Phe Thr Gly Gln 245 250 255 Pro Ala Thr Tyr His His Asn Ser Thr Thr Thr Trp Thr Gly Ser Arg 260 265 270 Thr Ala Pro Tyr Thr Pro Asn Leu Pro His His Gln Asn Gly His Leu 280 285 Gln His His Pro Pro Met Pro Pro His Pro Gly His Tyr Trp Pro Val 290 295 300 His Asn Glu Leu Ala Phe Gln Pro Pro Ile Ser Asn His Pro Ala Pro 305 310 315 Glu Tyr Trp Cys Ser Ile Ala Tyr Phe Glu Met Asp Val Gln Val Gly 330 Glu Thr Phe Lys Val Pro Ser Ser Cys Pro Ile Val Thr Val Asp Gly 340 345 350 Tyr Val Asp Pro Ser Gly Gly Asp Arg Phe Cys Leu Gly Gln Leu Ser

360

Asn Val His Arg Thr Glu Ala Ile Glu Arg Ala Arg Leu His Ile Gly

	370					375					380				
Lys 385	Gly	Val	Gln	Leu	Glu 390	Cys	Lys	Gly	Glu	G1y 395	Asp	Val	Trp	Val	Arg 400
Cys	Leu	Ser	Asp	His 405	Ala	Val	Phe	Val	Gln 410	Ser	Tyr	Tyr	Leu	Asp 415	Arg
			420		Pro			425					430		
		435			Phe		440					445			
	450				Ala	455					460				
465					Pro 470					475					480
				485	Ala				490					495	
	•		500		Arg			505					510		
		515			Ile		520					525			
	530				Gln	535					540				
Ile 545	Ala	Asp	Pro	Gln	Pro 550	Leu	Asp	Trp	Asp	Pro 555	Pro	Val	Ala	Thr	Met 560
	Ser	Lys	Gly	Glu 565	Glu	Leu	Phe	Thr	Gly 570		Val	Pro	Ile	Leu 575	
Glu	Leu	Asp	Gly 580	Asp	Val	Asn	Gly	His 585	Lys	Phe	Ser	Val	Ser 590		Glu
Gly	Glu	Gly 595	Asp	Ala	Thr	Tyr	Gly 600	Lys	Leu	Thr	Leu	Lys 605	Phe	Ile	Cys
Thr	Thr 610	Gly	Lys	Leu	Pro	Val 615	Pro	Trp	Pro	Thr	Leu 620	Val	Thr	Thr	Leu
625					Cys 630					635					640
His	Asp	Phe	Phe	Lys 645	Ser	Ala	Met	Pro	Glu 650	Gly	Tyr	Val	Gln	Glu 655	Arg
Thr	Ile	Phe	Phe 660	Lys	Asp	Asp	Gly	Asn 665	Tyr	Lys	Thr	Arg	Ala 670	Glu	Val
		675			Thr		680		_			685	_	_	
	690				Gly	695					700				
705					Val 710					715					720
				725	Lys				730					735	
Gln	Leu	Ala	Asp 740	His	Tyr	Gln	Gln	Asn 745	Thr	Pro	Ile	Gly	Asp 750	Gly	Pro
		755			Asn		760					765			
	770				Lys	775					780			Phe	Val
Thr 785	Ala	Ala	Gly	Ile	Thr 790	Leu	Gly	Met	Asp	Glu 795	Leu	Tyr	Lys		

## (2) INFORMATION FOR SEQ ID NO:78:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3138 base pairs

(B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...3135 (D) OTHER INFORMATION: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:78: ATG GCG GGC TGG ATC CAG GCC CAG CAG CTG CAG GGA GAC GCG CTG CGC 48 Met Ala Gly Trp Ile Gln Ala Gln Gln Leu Gln Gly Asp Ala Leu Arg 10 CAG ATG CAG GTG CTG TAC GGC CAG CAC TTC CCC ATC GAG GTC CGG CAC 96 Gln Met Gln Val Leu Tyr Gly Gln His Phe Pro Ile Glu Val Arg His 25 TAC TTG GCC CAG TGG ATT GAG AGC CAG CCA TGG GAT GCC ATT GAC TTG 144 Tyr Leu Ala Gln Trp Ile Glu Ser Gln Pro Trp Asp Ala Ile Asp Leu 40 GAC AAT CCC CAG GAC AGA GCC CAA GCC ACC CAG CTC CTG GAG GGC CTG 192 Asp Asn Pro Gln Asp Arg Ala Gln Ala Thr Gln Leu Leu Glu Gly Leu 55 GTG CAG GAG CTG CAG AAG AAG GCG GAG CAC CAG GTG GGG GAA GAT GGG 240 Val Gln Glu Leu Gln Lys Lys Ala Glu His Gln Val Gly Glu Asp Gly TTT TTA CTG AAG ATC AAG CTG GGG CAC TAC GCC ACG CAG CTC CAG AAA 288 Phe Leu Leu Lys Ile Lys Leu Gly His Tyr Ala Thr Gln Leu Gln Lys ACA TAT GAC CGC TGC CCC CTG GAG CTG GTC CGC TGC ATC CGG CAC ATT 336 Thr Tyr Asp Arg Cys Pro Leu Glu Leu Val Arg Cys Ile Arg His Ile 100 105 CTG TAC AAT GAA CAG AGG CTG GTC CGA GAA GCC AAC AAT TGC AGC TCT 384 Leu Tyr Asn Glu Gln Arg Leu Val Arg Glu Ala Asn Asn Cys Ser Ser 120 CCG GCT GGG ATC CTG GTT GAC GCC ATG TCC CAG AAG CAC CTT CAG ATC 432 Pro Ala Gly Ile Leu Val Asp Ala Met Ser Gln Lys His Leu Gln Ile 135 AAC CAG ACA TTT GAG GAG CTG CGA CTG GTC ACG CAG GAC ACA GAG AAT 480 Asn Gln Thr Phe Glu Glu Leu Arg Leu Val Thr Gln Asp Thr Glu Asn 150 155

GAG CTG AAG AAA CTG CAG CAG ACT CAG GAG TAC TTC ATC ATC CAG TAC

Glu Leu Lys Lys Leu Gln Gln Thr Gln Glu Tyr Phe Ile Ile Gln Tyr

CAG GAG AGC CTG AGG ATC CAA GCT CAG TTT GCC CAG CTG GCC CAG CTG

165

528

Gln	Glu	Ser	Leu 180	Arg	Ile	Gln	Ala	Gln 185	Phe	Ala	Gln	Leu	Ala 190	Gln	Leu	
					CTG Leu											624
					TGG Trp											672
					GCC Ala 230											720
					ATC Ile											768
					GCC Ala											816
					TGG Trp											864
					CGC Arg											912
					GAG Glu 310											960
					GCC Ala											1008
					CTG Leu											1056
					GGG Gly	Lys										1104
					ATC Ile											1152
					AAC Asn 390											1200
					CAC His											1248

					AAG Lys											1296
					GAG Glu											1344
					GAG Glu											1392
					GTC Val 470											1440
					AAT Asn											1488
					GTG Val											1536
					GAA Glu											1584
					GCG Ala											1632
					GGC Gly 550			_								1680
					TGG Trp											1728
	_				AAG Lys											1776
					GTG Val											1824
					ACC Thr											1872
					GCC Ala 630											1920
TGG	AAC	CTG	AAA	CCA	TTC	ACC	ACG	CGG	GAT	TTC	TCC	ATC	AGG	TCC	CTG	1968

Trp	Asn	Leu	Lys	Pro 645	Phe	Thr	Thr	Arg	Asp 650	Phe	Ser	Ile	Arg	Ser 655	Leu	
					GAC Asp											2016
					GTC Val											2064
					TAT Tyr											2112
					TCT Ser 710											2160
					TCC Ser											2208
					CCT Pro											2256
			_	_	ATG Met											2304
					AGT Ser											2352
					AGA Arg 790											2400
					ACC Thr											2448
					CTG Leu											2496
					GGC Gly											2544
					ATC Ile											2592
					ACC Thr 870											2640

TAC Tyr	CCC	GAC Asp	CAC His	Met 885	Lys	G CAG	CAC His	GAC Asp	Phe 890	Phe	: AAG	TCC Ser	GCC	ATC Met 895		2688
GAA Glu	GGC	TAC	Val	Glr	GAG Glu	CGC	ACC	ATC Ile 905	Ph∈	TTC Phe	AAG Lys	GAC Asp	GAC Asp 910	Gly	AAC Asn	2736
TAC Tyr	AAG Lys	ACC Thr 915	Arg	GCC	GAG Glu	GTG Val	AAG Lys 920	TTC Phe	GAG Glu	GGC Gly	GAC Asp	ACC Thr 925	CTG	GTG Val	AAC Asn	2784
CGC Arg	ATC Ile 930	GAG Glu	CTG Leu	AAG Lys	GGC Gly	ATC Ile 935	GAC Asp	TTC Phe	AAG Lys	GAG Glu	GAC Asp 940	GGC Gly	AAC Asn	ATC	CTG Leu	2832
GGG Gly 945	CAC His	AAG Lys	CTG Leu	GAG Glu	TAC Tyr 950	AAC Asn	TAC Tyr	AAC Asn	AGC Ser	CAC His 955	AAC Asn	GTC Val	тат Туг	ATC	ATG Met 960	2880
GCC Ala	GAC Asp	AAG Lys	CAG Gln	AAG Lys 965	AAC Asn	GGC Gly	ATC Ile	AAG Lys	GTG Val 970	AAC Asn	TTC Phe	AAG Lys	ATC Ile	CGC Arg 975	CAC His	2928
AAC Asn	ATC Ile	GAG Glu	GAC Asp 980	GGC Gly	AGC Ser	GTG Val	CAG Gln	CTC Leu 985	GCC Ala	GAC Asp	CAC His	TAC Tyr	CAG Gln 990	CAG Gln	AAC Asn	2976
ACC (	Pro	ATC Ile 995	GGC Gly	GAC Asp	GGC Gly	Pro	GTG Val 000	CTG Leu	CTG Leu	CCC Pro	Asp	AAC Asn .005	CAC His	TAC Tyr	CTG Leu	3024
AGC A	ACC Thr 010	CAG Gln	TCC Ser	GCC Ala	Leu	AGC Ser 015	AAA Lys	GAC Asp	CCC Pro	Asn	GAG Glu 020	AAG Lys	CGC Arg	GAT Asp	CAC His	3072
ATG ( Met V 1025	GTC /al	CTG ( Leu :	CTG Leu	Glu	TTC Phe 030	GTG . Val	ACC (	GCC Ala	Ala	GGG Gly 035	ATC Ile	ACT Thr	CTC Leu	Gly	ATG Met 040	3120
GAC (			Tyr :		TAA											3138

# (2) INFORMATION FOR SEQ ID NO:79:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1045 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:79:

Met Ala Gly Trp Ile Gln Ala Gln Gln Leu Gln Gly Asp Ala Leu Arg Gln Met Gln Val Leu Tyr Gly Gln His Phe Pro Ile Glu Val Arg His Tyr Leu Ala Gln Trp Ile Glu Ser Gln Pro Trp Asp Ala Ile Asp Leu Asp Asn Pro Gln Asp Arg Ala Gln Ala Thr Gln Leu Leu Glu Gly Leu Val Gln Glu Leu Gln Lys Lys Ala Glu His Gln Val Gly Glu Asp Gly Phe Leu Leu Lys Ile Lys Leu Gly His Tyr Ala Thr Gln Leu Gln Lys Thr Tyr Asp Arg Cys Pro Leu Glu Leu Val Arg Cys Ile Arg His Ile Leu Tyr Asn Glu Gln Arg Leu Val Arg Glu Ala Asn Asn Cys Ser Ser Pro Ala Gly Ile Leu Val Asp Ala Met Ser Gln Lys His Leu Gln Ile Asn Gln Thr Phe Glu Glu Leu Arg Leu Val Thr Gln Asp Thr Glu Asn Glu Leu Lys Lys Leu Gln Gln Thr Gln Glu Tyr Phe Ile Ile Gln Tyr Gln Glu Ser Leu Arg Ile Gln Ala Gln Phe Ala Gln Leu Ala Gln Leu Ser Pro Gln Glu Arg Leu Ser Arg Glu Thr Ala Leu Gln Gln Lys Gln Val Ser Leu Glu Ala Trp Leu Gln Arg Glu Ala Gln Thr Leu Gln Gln Tyr Arg Val Glu Leu Ala Glu Lys His Gln Lys Thr Leu Gln Leu Leu Arg Lys Gln Gln Thr Ile Ile Leu Asp Asp Glu Leu Ile Gln Trp Lys Arg Arg Gln Gln Leu Ala Gly Asn Gly Gly Pro Pro Glu Gly Ser Leu Asp Val Leu Gln Ser Trp Cys Glu Lys Leu Ala Glu Ile Ile Trp Gln Asn Arg Gln Gln Ile Arg Arg Ala Glu His Leu Cys Gln Gln Leu Pro Ile Pro Gly Pro Val Glu Glu Met Leu Ala Glu Val Asn Ala Thr Ile 310 315 Thr Asp Ile Ile Ser Ala Leu Val Thr Ser Thr Phe Ile Ile Glu Lys Gln Pro Pro Gln Val Leu Lys Thr Gln Thr Lys Phe Ala Ala Thr Val Arg Leu Leu Val Gly Gly Lys Leu Asn Val His Met Asn Pro Pro Gln Val Lys Ala Thr Ile Ile Ser Glu Gln Gln Ala Lys Ser Leu Leu Lys Asn Glu Asn Thr Arg Asn Glu Cys Ser Gly Glu Ile Leu Asn Asn Cys Cys Val Met Glu Tyr His Gln Ala Thr Gly Thr Leu Ser Ala His Phe Arg Asn Met Ser Leu Lys Arg Ile Lys Arg Ala Asp Arg Arg Gly Ala Glu Ser Val Thr Glu Glu Lys Phe Thr Val Leu Phe Glu Ser Gln Phe Ser Val Gly Ser Asn Glu Leu Val Phe Gln Val Lys Thr Leu Ser Leu

455 460 Pro Val Val Val Ile Val His Gly Ser Gln Asp His Asn Ala Thr Ala 470 475 Thr Val Leu Trp Asp Asn Ala Phe Ala Glu Pro Gly Arg Val Pro Phe 485 490 Ala Val Pro Asp Lys Val Leu Trp Pro Gln Leu Cys Glu Ala Leu Asn 505 500 510 Met Lys Phe Lys Ala Glu Val Gln Ser Asn Arg Gly Leu Thr Lys Glu 520 525 Asn Leu Val Phe Leu Ala Gln Lys Leu Phe Asn Asn Ser Ser Ser His 535 540 Leu Glu Asp Tyr Ser Gly Leu Ser Val Ser Trp Ser Gln Phe Asn Arg 550 555 Glu Asn Leu Pro Gly Trp Asn Tyr Thr Phe Trp Gln Trp Phe Asp Gly 565 570 Val Met Glu Val Leu Lys Lys His His Lys Pro His Trp Asn Asp Gly 580 585 Ala Ile Leu Gly Phe Val Asn Lys Gln Gln Ala His Asp Leu Leu Ile 600 Asn Lys Pro Asp Gly Thr Phe Leu Leu Arg Phe Ser Asp Ser Glu Ile 615 Gly Gly Ile Thr Ile Ala Trp Lys Phe Asp Ser Pro Glu Arg Asn Leu 630 635 640 Trp Asn Leu Lys Pro Phe Thr Thr Arg Asp Phe Ser Ile Arg Ser Leu 645 650 655 Ala Asp Arg Leu Gly Asp Leu Ser Tyr Leu Ile Tyr Val Phe Pro Asp 665 Arg Pro Lys Asp Glu Val Phe Ser Lys Tyr Tyr Thr Pro Val Leu Ala 675 680 Lys Ala Val Asp Gly Tyr Val Lys Pro Gln Ile Lys Gln Val Val Pro 695 700 Glu Phe Val Asn Ala Ser Ala Asp Ala Gly Gly Ser Ser Ala Thr Tyr 710 715 Met Asp Gln Ala Pro Ser Pro Ala Val Cys Pro Gln Ala Pro Tyr Asn 725 730 Met Tyr Pro Gln Asn Pro Asp His Val Leu Asp Gln Asp Gly Glu Phe 740 745 750 Asp Leu Asp Glu Thr Met Asp Val Ala Arg His Val Glu Glu Leu Leu 760 Arg Arg Pro Met Asp Ser Leu Asp Ser Arg Leu Ser Pro Pro Ala Gly 775 780 Leu Phe Thr Ser Ala Arg Gly Ser Leu Ser Trp Val Pro Arg Ala Arg 790 795 Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr 805 810 Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His 820 825 830 Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys 835 840 845 Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp 855 860 Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg 870 875 Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro 885 890 Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn 905 Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn

915 920	925
Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp 0 930 935 940	Gly Asn Ile Leu
Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn v 945 950 955	Val Tyr Ile Met 960
Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe 1 965 970	
Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His 1980 985	Tyr Gln Gln Asn
Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp	_
Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu I	005 Lys Arg Asp His
1010 1015 1020 Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile 1	Thr Leu Gly Met
025 1030 1035	1040
Asp Glu Leu Tyr Lys	
1045	
(2) INFORMATION FOR SEQ ID NO:80:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 28 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:80:	·
(NE) SEQUENCE PERCENTITION. SEQ 12 NO.00.	
TGGGATCCTC AGGCCGTGCT GCTGGCCG	28
(2) INFORMATION FOR SEQ ID NO:81:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 27 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:81:	
GTCTCGAGGG AGCATGGGCA CCTTGCG	27
(2) INFORMATION FOR SEQ ID NO:82:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 27 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:82:	
	25
TGGGATCCGA GAAGTCTATA TCCCATC	27

(2) INFORMATION FOR SEQ ID NO:83:

28
28
30
30
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30
30
30
30
30
30
30
30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:87:	
GTCTCGAGCC ATGGACGAAC TGTTCCCCCT CATC	34
(2) INFORMATION FOR SEQ ID NO:88:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 31 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:88:	
GTGGATCCAA GGAGCTGATC TGACTCAGCA G	31
(2) INFORMATION FOR SEQ ID NO:89:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 32 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:89:	
GTGGATCCTT AGGAGCTGAT CTGACTCAGC AG	32
(2) INFORMATION FOR SEQ ID NO:90:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 32 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:90:	
CCTCCTAAGC TTATCATGGA CCATTATGAT TC	32
(2) INFORMATION FOR SEQ ID NO:91:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 33 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:91:	
CCTCCTGGAT CCCTGCGCAG GATGATGGTC CAG	33

(2) INFORMATION FOR SEQ ID NO:92:

<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 45 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:92:	
GGATGGAAGC TTCAATGGCT GCCATCCGGA AGAAACTGGT GATTG	45
(2) INFORMATION FOR SEQ ID NO:93:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 45 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:93:	
GGATGGGGAT CCTCACAAGA CAAGGCAACC AGATTTTTTC TTCCC	45
(2) INFORMATION FOR SEQ ID NO:94:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 29 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:94:	
GGGAAGCTTC CATGAGCGAG ACGGTCATC	29
(2) INFORMATION FOR SEQ ID NO:95:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 28 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:95:	
CCCGGATCCT CAGGGAGAAC CCCGCTTC	28
(2) INFORMATION FOR SEQ ID NO:96:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 30 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li></ul>	

(D) TOPOLOGY: linear

GTGAATTCGC TTCCTCTTGA GGGAACC

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:96:	
GTGAATTCGA CCATGGAGCG GCCCCCGGGG	30
(2) INFORMATION FOR SEQ ID NO:97:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 27 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:97:	
GTGGTACCCA TTCTGTTAAC CAACTCC	27
(2) INFORMATION FOR SEQ ID NO:98:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 28 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:98:	
GTGGTACCTC ATTCTGTTAA CCAACTCC	28
(2) INFORMATION FOR SEQ ID NO:99:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 28 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:99:	
GTCTCGAGAG ATGCTGTCCC GTGGGTGG	28
(2) INFORMATION FOR SEQ ID NO:100:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 27 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:100:	

27

(2) INFORMATION FOR SEQ ID NO:101:

<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 27 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:101:	
GTGAATTCAC TTCCTCTTGA GGGAACC	27
(2) INFORMATION FOR SEQ ID NO:102:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 29 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:102:	
GTCTCGAGCC ATGGAGAACT TCCAAAAGG	29
(2) INFORMATION FOR SEQ ID NO:103:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 28 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:103:	
GTGGATCCCA GAGTCGAAGA TGGGGTAC	28
(2) INFORMATION FOR SEQ ID NO:104:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 29 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:104:	
GTGGATCCTC AGAGTCGAAG ATGGGGTAC	29
(2) INFORMATION FOR SEQ ID NO:105:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 30 base pairs</li><li>(B) TYPE: nucleic acid</li></ul>	

(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:105:	
GTGAATTCGG CGATGCCAGA CCCCGCGGCG	30
(2) INFORMATION FOR SEQ ID NO:106:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 32 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:106:	
GTGGATCCCA GGCACAGGCA GCCTCAGCCT TC	32
(2) INFORMATION FOR SEQ ID NO:107:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 33 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:107:	
GTGGATCCTC AGGCACAGGC AGCCTCAGCC TTC	33
(2) INFORMATION FOR SEQ ID NO:108:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 2616 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> <li>(ii) MOLECULE TYPE: cDNA</li> <li>(ix) FEATURE:</li> </ul>	
<ul><li>(A) NAME/KEY: Coding Sequence</li><li>(B) LOCATION: 12613</li><li>(D) OTHER INFORMATION:</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:108:	
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  1 5 10 15	48
GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 30	96

	GGC Gly															144
	ACC Thr 50															192
	ACC Thr															240
	CAC His															288
	ACC Thr															336
	AAG Lys															384
	GAC Asp 130															432
	TAC Tyr															480
	ATC															528
	CAG Gln															576
	GTG Val															624
	AAA Lys 210															672
	ACC Thr															720
	CTC Leu															768
GCG	GCG	CAC	CTG	ccc	TTC	TTC	TAC	GGC	AGC	ATC	TCG	CGT	GCC	GAG	GCC	816

Ala	Ala	His	Leu 260	Pro	Phe	Phe	Tyr	Gly 265	Ser	Ile	Ser	Arg	Ala 270	Glu	Ala	
					CTG Leu											864
					TCG Ser											912
					CAC His 310							_	_			960
			_	_	GGC Gly											1008
					GAC Asp											1056
					TCG Ser											1104
					ATG Met											1152
					CTG Leu 390											1200
					GCT Ala											1248
					CGT Arg											1296
					AAG Lys											1344
					CTC Leu											1392
					GCG Ala 470											1440
					CAG Gln											1488

					CTG Leu											1536
	_		_		GCT Ala	_										1584
	_	_		_	AGA Arg											1632
					CGC Arg 550											1680
			_		GTG Val											1728
					CTC Leu											1776
					TGC Cys											1824
					AAG Lys											1872
					GCA Ala 630											1920
					GAC Asp											1968
					CTC Leu											2016
					CTG Leu											2064
					CTG Leu											2112
					GTG Val 710											2160
CTG	GTT	AAC	CGG	CAC	TAC	GCC	AAG	ATC	AGC	GAC	TTT	GGC	CTC	TCC	AAA	2208

Leu	Val	Asn	Arg	His 725	Tyr	Ala	Lys	Ile	Ser 730	Asp	Phe	Gly	Leu	Ser 735	Lys	
					GAC Asp											2256
					TAC Tyr											2304
					GTC Val											2352
					AAG Lys 790											2400
					CAG Gln											2448
					GCA Ala											2496
					TTC Phe											2544
					AGC Ser											2592
			_	_	TGT Cys 870		TGA									2616

### (2) INFORMATION FOR SEQ ID NO:109:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 871 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:109:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 1 5 15 15 
Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 30 
Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile

Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser Gly Leu Arg Ser Arg Ala Gln Ala Ser Asn Ser Ala Met Pro Asp Pro Ala Ala His Leu Pro Phe Phe Tyr Gly Ser Ile Ser Arg Ala Glu Ala Glu Glu His Leu Lys Leu Ala Gly Met Ala Asp Gly Leu Phe Leu Leu Arg Gln Cys Leu Arg Ser Leu Gly Gly Tyr Val Leu Ser Leu Val His Asp Val Arg Phe His His Phe Pro Ile Glu Arg Gln Leu Asn Gly Thr Tyr Ala Ile Ala Gly Gly Lys Ala His Cys Gly Pro Ala Glu Leu Cys Glu Phe Tyr Ser Arg Asp Pro Asp Gly Leu Pro Cys Asn Leu Arg Lys Pro Cys Asn Arg Pro Ser Gly Leu Glu Pro Gln Pro Gly Val Phe Asp Cys Leu Arg Asp Ala Met Val Arg Asp Tyr Val Arg Gln Thr Trp Lys Leu Glu Gly Glu Ala Leu Glu Gln Ala Ile Ile Ser Gln Ala Pro Gln Val Glu Lys Leu Ile Ala Thr Thr Ala His Glu Arg Met Pro Trp Tyr His Ser Ser Leu Thr Arg Glu Glu Ala Glu Arg Lys Leu Tyr Ser Gly Ala Gln Thr Asp Gly Lys Phe Leu Leu Arg Pro Arg Lys Glu Gln Gly Thr Tyr Ala Leu Ser Leu Ile Tyr Gly Lys Thr Val Tyr His Tyr Leu Ile Ser Gln Asp Lys Ala Gly Lys Tyr Cys Ile Pro Glu Gly Thr Lys Phe Asp Thr Leu Trp Gln Leu Val Glu Tyr Leu Lys Leu Lys Ala Asp Gly Leu Ile Tyr Cys Leu Lys Glu Ala Cys Pro Asn Ser Ser Ala Ser

500 505 Asn Ala Ser Gly Ala Ala Ala Pro Thr Leu Pro Ala His Pro Ser Thr 520 525 Leu Thr His Pro Gln Arg Arg Ile Asp Thr Leu Asn Ser Asp Gly Tyr 535 540 Thr Pro Glu Pro Ala Arg Ile Thr Ser Pro Asp Lys Pro Arg Pro Met 550 555 Pro Met Asp Thr Ser Val Tyr Glu Ser Pro Tyr Ser Asp Pro Glu Glu 565 570 Leu Lys Asp Lys Lys Leu Phe Leu Lys Arg Asp Asn Leu Leu Ile Ala 580 585 Asp Ile Glu Leu Gly Cys Gly Asn Phe Gly Ser Val Arg Gln Gly Val 600 Tyr Arg Met Arg Lys Lys Gln Ile Asp Val Ala Ile Lys Val Leu Lys 615 620 Gln Gly Thr Glu Lys Ala Asp Thr Glu Glu Met Met Arg Glu Ala Gln 630 635 Ile Met His Gln Leu Asp Asn Pro Tyr Ile Val Arg Leu Ile Gly Val 645 650 655 Cys Gln Ala Glu Ala Leu Met Leu Val Met Glu Met Ala Gly Gly Gly 660 665 670 Pro Leu His Lys Phe Leu Val Gly Lys Arg Glu Glu Ile Pro Val Ser 675 680 Asn Val Ala Glu Leu Leu His Gln Val Ser Met Gly Met Lys Tyr Leu 690 695 700 Glu Glu Lys Asn Phe Val His Arg Asp Leu Ala Ala Arg Asn Val Leu 710 715 Leu Val Asn Arg His Tyr Ala Lys Ile Ser Asp Phe Gly Leu Ser Lys 725 730 Ala Leu Gly Ala Asp Asp Ser Tyr Tyr Thr Ala Arg Ser Ala Gly Lys 740 745 Trp Pro Leu Lys Trp Tyr Ala Pro Glu Cys Ile Asn Phe Arg Lys Phe 760 765 Ser Ser Arg Ser Asp Val Trp Ser Tyr Gly Val Thr Met Trp Glu Ala 775 780 Leu Ser Tyr Gly Gln Lys Pro Tyr Lys Lys Met Lys Gly Pro Glu Val 790 795 Met Ala Phe Ile Glu Gln Gly Lys Arg Met Glu Cys Pro Pro Glu Cys 805 810 Pro Pro Glu Leu Tyr Ala Leu Met Ser Asp Cys Trp Ile Tyr Lys Trp 820 825 830 Glu Asp Arg Pro Asp Phe Leu Thr Val Glu Gln Arg Met Arg Ala Cys 835 840 845 Tyr Tyr Ser Leu Ala Ser Lys Val Glu Gly Pro Pro Gly Ser Thr Gln 855 Lys Ala Glu Ala Ala Cys Ala 865 870

#### (2) INFORMATION FOR SEQ ID NO:110:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2598 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:

(A) NAME/KEY: Coding Sequence (B) LOCATION: 1...2595

(D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:110:

			CAC His									48
			CAC His		_	_		_		_		96
			TGC Cys				_		_		-	144
			CGC Arg 55				_				:	192
			ATT Ile								:	240
			TAC Tyr								:	288
			AAC Asn									336
			CGA Arg									384
			GGC Gly 135									432
			AAG Lys			Thr						480
			AGC Ser									528
										CGG Arg		576
	Gly		GCC Ala	Ser				Lys		GTG Val		624

TAC CAC T Tyr His T 210												672
GAG GGC A Glu Gly T 225												720
CTG AAG G Leu Lys A	la Asp		_				_	_				768
AGC AGT G Ser Ser A												816
CAC CCA T His Pro S			_	o Gln			-		_			864
TCA GAT G Ser Asp G 290												912
CCG CGG C Pro Arg F 305												960
GAC CCA G Asp Pro G												1008
CTC CTC A												1056
CGC CAG G Arg Gln G				g Lys		_	_		_	_	_	1104
AAG GTG C Lys Val L 370												1152
CGC GAG G Arg Glu A 385									_	_		1200
CTC ATT G								_		_		1248
GCT GGG G Ala Gly G												1296
ATC CCT G	TG AGC	aat gtg	GCC GA	G CTG	CTG	CAC	CAG	GTG	TCC	ATG	GGG	1344

Ile	Pro	Val 435	Ser	Asn	Val	Ala	Glu 440	Leu	Leu	His	Gln	Val 445	Ser	Met	Gly	
					GAG Glu											1392
					GTT Val 470											1440
					CTG Leu											1488
	_				CCG Pro											1536
			_		AGC Ser											1584
			_		TCC Ser											1632
					GCC Ala 550											1680
					CCC Pro											1728
					GAT Asp											1776
_					TAC Tyr				_	-		_				1824
					GCT Ala											1872
					AAG Lys 630											1920
					GAC Asp											1968
					GGC Gly											2016

				AAG Lys						2064
				GTG Val 695						2112
				TTC Phe						2160
				TTC Phe						2208
				GGC Gly						2256
				GAG Glu						2304
				CAC His 775						2352
				AAC Asn						2400
				GAC Asp						2448
		_		CCC Pro				_	_	2496
				AAC Asn				_		2544
				GGG Gly 855				_		2592
AAG Lys 865	TAA									2598

### (2) INFORMATION FOR SEQ ID NO:111:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 865 amino acids

(B) TYPE: amino acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:111:

Met Pro Asp Pro Ala Ala His Leu Pro Phe Phe Tyr Gly Ser Ile Ser 10 Arg Ala Glu Ala Glu Glu His Leu Lys Leu Ala Gly Met Ala Asp Gly 20 25 Leu Phe Leu Leu Arg Gln Cys Leu Arg Ser Leu Gly Gly Tyr Val Leu 45 40 Ser Leu Val His Asp Val Arg Phe His His Phe Pro Ile Glu Arg Gln Leu Asn Gly Thr Tyr Ala Ile Ala Gly Gly Lys Ala His Cys Gly Pro 65 70 75 Ala Glu Leu Cys Glu Phe Tyr Ser Arg Asp Pro Asp Gly Leu Pro Cys 85 90 Asn Leu Arg Lys Pro Cys Asn Arg Pro Ser Gly Leu Glu Pro Gln Pro 100 105 Gly Val Phe Asp Cys Leu Arg Asp Ala Met Val Arg Asp Tyr Val Arg 120 125 Gln Thr Trp Lys Leu Glu Gly Glu Ala Leu Glu Gln Ala Ile Ile Ser 135 140 Gln Ala Pro Gln Val Glu Lys Leu Ile Ala Thr Thr Ala His Glu Arg 145 150 155 Met Pro Trp Tyr His Ser Ser Leu Thr Arg Glu Glu Ala Glu Arg Lys 170 165 Leu Tyr Ser Gly Ala Gln Thr Asp Gly Lys Phe Leu Leu Arg Pro Arg 180 185 190 Lys Glu Gln Gly Thr Tyr Ala Leu Ser Leu Ile Tyr Gly Lys Thr Val 200 195 Tyr His Tyr Leu Ile Ser Gln Asp Lys Ala Gly Lys Tyr Cys Ile Pro 220 215 Glu Gly Thr Lys Phe Asp Thr Leu Trp Gln Leu Val Glu Tyr Leu Lys 230 235 Leu Lys Ala Asp Gly Leu Ile Tyr Cys Leu Lys Glu Ala Cys Pro Asn 245 250 Ser Ser Ala Ser Asn Ala Ser Gly Ala Ala Ala Pro Thr Leu Pro Ala 260 265 His Pro Ser Thr Leu Thr His Pro Gln Arg Arg Ile Asp Thr Leu Asn 275 280 285 Ser Asp Gly Tyr Thr Pro Glu Pro Ala Arg Ile Thr Ser Pro Asp Lys 290 295 300 Pro Arg Pro Met Pro Met Asp Thr Ser Val Tyr Glu Ser Pro Tyr Ser 310 315 Asp Pro Glu Glu Leu Lys Asp Lys Lys Leu Phe Leu Lys Arg Asp Asn 330 335 325 Leu Leu Ile Ala Asp Ile Glu Leu Gly Cys Gly Asn Phe Gly Ser Val 340 345 350 Arg Gln Gly Val Tyr Arg Met Arg Lys Lys Gln Ile Asp Val Ala Ile 355 360 365 Lys Val Leu Lys Gln Gly Thr Glu Lys Ala Asp Thr Glu Glu Met Met 375 380 Arg Glu Ala Gln Ile Met His Gln Leu Asp Asn Pro Tyr Ile Val Arg

385		_,	,	<b>.</b> .	390		<b>01</b>		•	395				<b>61</b>	400
Leu	Ile	GIÀ	Val	Cys 405	Gin	Ala	GIu	Ala	410		Leu	Vai	Met	415	Met
Ala	Gly	Gly	Gly 420		Leu	His	Lys	Phe 425	Leu		Gly	Lys	Arg 430		Glu
Ile	Pro	Val 435	Ser	Asn	Val	Ala	Glu 440		Leu	His	Gln	Val 445	Ser	Met	Gly
Met	Lys 450	Tyr	Leu	Glu	Glu	Lys 455	Asn	Phe	Val	His	Arg 460	Asp	Leu	Ala	Ala
Arg 465	Asn	Val	Leu	Leu	Val 470		Arg	His	Tyr	Ala 475	Lys	Ile	Ser	Asp	Phe 480
		Ser	Lys	Ala 485		Gly	Ala	Asp	Asp 490		Tyr	Tyr	Thr	Ala 495	Arg
Ser	Ala	Gly	Lys 500		Pro	Leu	Lys	Trp 505		Ala	Pro	Glu	Cys 510		Asn
Phe	Arg	Lys 515	Phe	Ser	Ser	Arg	Ser 520		Val	Trp	Ser	Tyr 525	Gly	Val	Thr
Met	Trp 530	Glu	Ala	Leu	Ser	Tyr 535	_	Gln	Lys	Pro	Tyr 540	Lys	Lys	Met	Lys
545					550					555					560
				565					570		Met			575	
	_	-	580					585			Thr		590		
		595					600	_			Val	605			_
_	610					615					620	_			
625					630	_				635					640
				645					650	)	, His			655	
	_		660					665	,		Lys		670		
		675					680				Trp	685			
	690					695					Arg 700				
705	•				710					715					720
		_		725					730	)	Asn Asn			735	
			740					745	,		Leu		750	)	
		755					760				e Met	765			
	770					775					780 His	ı			
785					790					795					800
				805					810	)	Leu			815	,
			820					825	<b>,</b>		His		830	)	
		835					840				/ Met	845	i		
Giu	THE	vai				~_1			200	- <u>-</u> y				_50	-1-

850 855 860 Lys

ьуs 865

### (2) INFORMATION FOR SEQ ID NO:112:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1635 base pairs(B) TYPE: nucleic acid

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...1632 (D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:112:

					AAG Lys												48
					AGA Arg												96
	-				GAC Asp											:	144
					CTG Leu											:	192
					ATT Ile 70			-								:	240
-		-			GAT Asp											:	288
					CCC Pro											;	336
					TGC Cys											;	384
					CTT Leu												432
GAC	TTT	GGA	СТА	GCC	AGA	GCT	TTT	GGA	GTC	CCT	GTT	CGT	ACT	TAC	ACC		480

Asp 145	Phe	Gly	Leu	Ala	Arg 150	Ala	Phe	Gly	Val	Pro 155	Val	Arg	Thr	Tyr	Thr 160	
					CTG Leu											528
					ACA Thr											576
					ACT Thr							_			_	624
					CGG Arg											672
					GTT Val 230											720
					CAA Gln											768
					TTG Leu											816
					AAG Lys											864
					CCC Pro									_	_	912
					GGC Gly 310										ATC Ile 320	960
					GGC Gly											1008
					GAT Asp										TTC Phe	1056
					AAG Lys							_		_		1104
					GTG Val										ATG Met	1152

					ATG Met					1200
					GGC Gly 410					1248
		 			GTG Val					1296
					ATC Ile					1344
	-				ATC Ile					1392
					CGC Arg					1440
					CAG Gln 490					1488
					TAC Tyr					1536
					GAT Asp					1584
					GGC Gly			AAG Lys	т	1633
AA										1635

### (2) INFORMATION FOR SEQ ID NO:113:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 544 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:113:

Met Glu Asn Phe Gln Lys Val Glu Lys Ile Gly Glu Gly Thr Tyr Gly
1 5 10 15

Val	Val	Tyr	Lys 20	Ala	Arg	Asn	Lys	Leu 25	Thr	Gly	Glu	Val	Val 30	Ala	Leu
Lys	Lys	Ile 35	Arg	Leu	Asp	Thr	Glu 40	Thr	Glu	Gly	Val	Pro 45	Ser	Thr	Ala
Ile	Arg 50	Glu	Ile	Ser	Leu	Leu 55	Lys	Glu	Leu	Asn	His 60	Pro	Asn	Ile	Val
Lys 65	Leu	Leu	Asp	Val	Ile 70	His	Thr	Glu	Asn	Lys 75	Leu	Tyr	Leu	Val	Phe 80
	Phe			85			_	_	90		_			95	
Thr	Gly	Ile	Pro 100	Leu	Pro	Leu	Ile	Lys 105	Ser	Tyr	Leu	Phe	Gln 110	Leu	Leu
Gln	Gly	Leu 115	Ala	Phe	Cys	His	Ser 120	His	Arg	Val	Leu	His 125	Arg	Asp	Leu
-	Pro 130					135					140		_		
Asp 145	Phe	Gly	Leu	Ala	Arg 150	Ala	Phe	Gly	Val	Pro 155	Val	Arg	Thr	Tyr	Thr 160
His	Glu	Va1	Val	Thr 165	Leu	Trp	Tyr	Arg	Ala 170	Pro	Glu	Ile	Leu	Leu 175	Gly
Ser	Lys	Tyr	Tyr 180	Ser	Thr	Ala	Val	Asp 185	Ile	Trp	Ser	Leu	Gly 190	Cys	Ile
	Ala	195				_	200					205	_		
Ile	Asp 210	Gln	Leu	Phe	Arg	Ile 215	Phe	Arg	Thr	Leu	Gly 220	Thr	Pro	Asp	Glu
225	Val	-		_	230					235		_			240
	Lys			245					250					255	
Glu	Asp	Gly	Arg 260	Ser	Leu	Leu	Ser	Gln 265	Met	Leu	His	Tyr	Asp 270	Pro	Asn
Lys	Arg	Ile 275	Ser	Ala	Lys	Ala	Ala 280	Leu	Ala	His	Pro	Phe 285	Phe	Gln	Asp
	Thr 290	_				295		_		_	300				
Thr 305	Met	Val	Ser	Lys	Gly 310	Glu	Glu	Leu	Phe	Thr 315	Gly	Val	Val	Pro	Ile 320
Leu	Val	Glu	Leu	Asp 325	Gly	Asp	Val	Asn	Gly 330	His	Lys	Phe	Ser	Val 335	Ser
-	Glu	_	340	_	-			345	_	_			350	_	
Ile	Cys	Thr 355		Gly			Pro 360		Pro	Trp	Pro	Thr 365		Val	Thr
	370		_	_		375	-				380				Met
Lys 385	Gln	His	Asp	Phe	Phe 390	Lys	Ser	Ala	Met	Pro 395	Glu	Gly	Tyr	Val	Gln 400
Glu	Arg	Thr	Ile	Phe 405	Phe	Lys	Asp	Asp	Gly 410	Asn	Tyr	Lys	Thr	Arg 415	Ala
	Val	-	420					425					430		
Gly	Ile	Asp 435	Phe	Lys	Glu	Asp	Gly 440	Asn	Ile	Leu	Gly	His 445	Lys	Leu	Glu
Tyr	Asn 450	Tyr	Asn	Ser	His	Asn 455	Val	Tyr	Ile	Met	Ala 460	Asp	Lys	Gln	Lys
Asn 465	Gly	Ile	Lys	Val	Asn 470	Phe	Lys	Ile	Arg	His 475	Asn	Ile	Glu	Asp	Gly 480

Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp 485 490 495 Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala 505 500 Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu 520 525 Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys (2) INFORMATION FOR SEQ ID NO:114: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1635 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...1632 (D) OTHER INFORMATION: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:114: ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG 48 Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 1 5 GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC 96 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 35 40 TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 50 CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG 240 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 CGC ACC ATC TTC TTC AAG GAC GAC GGC AAC TAC AAG ACC CGC GCC GAG 336 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 105 GTG AAG TTC GAG GGC GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC 384 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly

120

115

125

ATC GA																432
AAC TY Asn Ty 145																480
GGC A																528
GTG C																576
CCC G	al															624
AGC A Ser L																672
GTG A Val T 225																720
GGA C Gly L																768
GGA G Gly G																816
GGA G Gly G																. 864
GGT G Gly V 2																912
AAC C Asn H 305																960
AAA C Lys L	CTC Leu	TAC Tyr	CTG Leu	GTT Val 325	TTT Phe	GAA Glu	TTT Phe	CTG Leu	CAC His 330	CAA Gln	GAT Asp	CTC	AAG Lys	AAA Lys 335	TTC Phe	1008
ATG G Met A														Lys		1056
TAT C	CTG Leu	TTC Phe	CAG Gln	CTG Leu	CTC Leu	CAG Gln	GGC Gly	CTA Leu	GCT Ala	TTC Phe	TGC Cys	CAT His	TCT Ser	CAT His	CGG Arg	1104

360 355 365 GTC CTC CAC CGA GAC CTT AAA CCT CAG AAT CTG CTT ATT AAC ACA GAG 1152 Val Leu His Arg Asp Leu Lys Pro Gln Asn Leu Leu Ile Asn Thr Glu 375 380 GGG GCC ATC AAG CTA GCA GAC TTT GGA CTA GCC AGA GCT TTT GGA GTC 1200 Gly Ala Ile Lys Leu Ala Asp Phe Gly Leu Ala Arg Ala Phe Gly Val 395 CCT GTT CGT ACT TAC ACC CAT GAG GTG GTG ACC CTG TGG TAC CGA GCT 1248 Pro Val Arg Thr Tyr Thr His Glu Val Val Thr Leu Trp Tyr Arg Ala 405 410 CCT GAA ATC CTC CTG GGC TCG AAA TAT TAT TCC ACA GCT GTG GAC ATC 1296 Pro Glu Ile Leu Leu Gly Ser Lys Tyr Tyr Ser Thr Ala Val Asp Ile 420 425 TGG AGC CTG GGC TGC ATC TTT GCT GAG ATG GTG ACT CGC CGG GCC CTG 1344 Trp Ser Leu Gly Cys Ile Phe Ala Glu Met Val Thr Arg Arg Ala Leu 435 440 TTC CCT GGA GAT TCT GAG ATT GAC CAG CTC TTC CGG ATC TTT CGG ACT 1392 Phe Pro Gly Asp Ser Glu Ile Asp Gln Leu Phe Arg Ile Phe Arg Thr 450 455 CTG GGG ACC CCA GAT GAG GTG GTG TGG CCA GGA GTT ACT TCT ATG CCT 1440 Leu Gly Thr Pro Asp Glu Val Val Trp Pro Gly Val Thr Ser Met Pro 465 470 475 GAT TAC AAG CCA AGT TTC CCC AAG TGG GCC CGG CAA GAT TTT AGT AAA 1488 Asp Tyr Lys Pro Ser Phe Pro Lys Trp Ala Arg Gln Asp Phe Ser Lys 485 GTT GTA CCT CCC CTG GAT GAA GAT GGA CGG AGC TTG TTA TCG CAA ATG 1536 Val Val Pro Pro Leu Asp Glu Asp Gly Arg Ser Leu Leu Ser Gln Met 500 505 CTG CAC TAC GAC CCT AAC AAG CGG ATT TCG GCC AAG GCA GCC CTG GCT 1584 Leu His Tyr Asp Pro Asn Lys Arg Ile Ser Ala Lys Ala Ala Leu Ala 515 520 525 CAC CCT TTC TTC CAG GAT GTG ACC AAG CCA GTA CCC CAT CTT CGA CTC T 1633 His Pro Phe Phe Gln Asp Val Thr Lys Pro Val Pro His Leu Arg Leu 535 540

1635

(2) INFORMATION FOR SEQ ID NO:115:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 544 amino acids
- (B) TYPE: amino acid

GA

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
  (v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:115:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 85 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 155 150 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 170 165 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 190 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 220 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 230 235 Gly Leu Arg Ser Arg Ala Met Glu Asn Phe Gln Lys Val Glu Lys Ile 245 250 Gly Glu Gly Thr Tyr Gly Val Val Tyr Lys Ala Arg Asn Lys Leu Thr 260 265 270 Gly Glu Val Val Ala Leu Lys Lys Ile Arg Leu Asp Thr Glu Thr Glu 280 285 Gly Val Pro Ser Thr Ala Ile Arg Glu Ile Ser Leu Leu Lys Glu Leu 295 300 Asn His Pro Asn Ile Val Lys Leu Leu Asp Val Ile His Thr Glu Asn 315 310 Lys Leu Tyr Leu Val Phe Glu Phe Leu His Gln Asp Leu Lys Lys Phe 330 Met Asp Ala Ser Ala Leu Thr Gly Ile Pro Leu Pro Leu Ile Lys Ser 345 340 350 Tyr Leu Phe Gln Leu Leu Gln Gly Leu Ala Phe Cys His Ser His Arg 365 360 Val Leu His Arg Asp Leu Lys Pro Gln Asn Leu Leu Ile Asn Thr Glu 380 375 Gly Ala Ile Lys Leu Ala Asp Phe Gly Leu Ala Arg Ala Phe Gly Val 390 395 Pro Val Arg Thr Tyr Thr His Glu Val Val Thr Leu Trp Tyr Arg Ala 405 410 Pro Glu Ile Leu Leu Gly Ser Lys Tyr Tyr Ser Thr Ala Val Asp Ile 425 Trp Ser Leu Gly Cys Ile Phe Ala Glu Met Val Thr Arg Arg Ala Leu

		435					440					445				
Phe	Pro 450	_	Asp	Ser	Glu	Ile 455		Gln	Leu	Phe	Arg 460		Phe	Arg	Thr	
	Gly	Thr	Pro	Asp	Glu	Val	Val	Trp	Pro		Val	Thr	Ser	Met		
465 Asp	ጥህተ	Lvs	Pro	Ser	470 Phe	Pro	Lvs	TYTO	Δla	475	Gln	Asn	Phe	Ser	480 Lvs	
	-1-	_,,		485					490	·ug	<b>01</b>	nup	1110	495	בעם	
Val	Val	Pro	Pro 500	Leu	Asp	Glu	Asp	Gly 505	Arg	Ser	Leu	Leu	Ser 510	Gln	Met	•
		515	_		Asn	_	520				_	525				
His	Pro 530	Phe	Phe	Gln	Asp	Val 535	Thr	Lys	Pro	Val	Pro 540	His	Leu	Arg	Leu	
		(2	) INI	FORM	ATIOI	v FOI	R SE	Q ID	NO:	116:						
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	ι.	i i )   ?	MOT.FX	TIT.E	TYPE	e ci	ΔΤΑ									
			FEAT			· ·										
		(A)	NAI	ΜΕ/KI	EY: C	codir	ng Se	eque	ıce							
		(B)	LO	CATIO	ON: 1	2	2529	•								
		(D)	OTI	HER :	INFOF	TAM!	CON:									
	()	ci) S	SEQUI	ENCE	DESC	RIP	rion	: SEX	Q ID	NO:	116:					
ATG					DESC				•			GTG	ccc	ATC	CTG	48
Met	GTG	AGC	AAG	GGC Gly		GAG	CTG	TTC	ACC Thr	GGG	GTG			Ile		48
	GTG	AGC	AAG	GGC	GAG	GAG	CTG	TTC	ACC	GGG	GTG					48
Met 1 GTC	GTG Val GAG	AGC Ser CTG	AAG Lys GAC	GGC Gly 5 GGC	GAG Glu GAC	GAG Glu GTA	CTG Leu AAC	TTC Phe	ACC Thr 10	GGG Gly AAG	GTG Val	Val AGC	Pro GTG	Ile 15 TCC	Leu GGC	<b>4</b> 8 96
Met 1 GTC	GTG Val GAG	AGC Ser CTG	AAG Lys GAC	GGC Gly 5 GGC	GAG Glu	GAG Glu GTA	CTG Leu AAC	TTC Phe	ACC Thr 10	GGG Gly AAG	GTG Val	Val AGC	Pro GTG	Ile 15 TCC	Leu GGC	
Met 1 GTC Val	GTG Val GAG Glu	AGC Ser CTG Leu	AAG Lys GAC Asp 20	GGC Gly 5 GGC Gly	GAG Glu GAC Asp	GAG Glu GTA Val	CTG Leu AAC Asn	TTC Phe GGC Gly 25	ACC Thr 10 CAC His	GGG Gly AAG Lys	GTG Val TTC Phe	Val AGC Ser	Pro GTG Val 30	Ile 15 TCC Ser	Leu GGC Gly	96
Met 1 GTC Val	GTG Val GAG Glu	AGC Ser CTG Leu	AAG Lys GAC Asp 20	GGC Gly 5 GGC Gly	GAG Glu GAC	GAG Glu GTA Val	CTG Leu AAC Asn	TTC Phe GGC Gly 25 GGC	ACC Thr 10 CAC His	GGG Gly AAG Lys	GTG Val TTC Phe	Val AGC Ser	Pro GTG Val 30	Ile 15 TCC Ser	GGC Gly	
Met 1 GTC Val	GTG Val GAG Glu	AGC Ser CTG Leu	AAG Lys GAC Asp 20	GGC Gly 5 GGC Gly	GAG Glu GAC Asp	GAG Glu GTA Val	CTG Leu AAC Asn	TTC Phe GGC Gly 25 GGC	ACC Thr 10 CAC His	GGG Gly AAG Lys	GTG Val TTC Phe	Val AGC Ser	Pro GTG Val 30	Ile 15 TCC Ser	GGC Gly	96
Met 1 GTC Val GAG Glu	GTG Val GAG Glu GGC Gly	AGC Ser CTG Leu GAG Glu 35	AAG Lys GAC Asp 20 GGC Gly	GGC Gly 5 GGC Gly GAT Asp	GAG Glu GAC Asp	GAG Glu GTA Val ACC Thr	CTG Leu AAC Asn TAC Tyr 40	TTC Phe GGC Gly 25 GGC Gly	ACC Thr 10 CAC His	GGG Gly AAG Lys CTG Leu	GTG Val TTC Phe ACC Thr	AGC Ser CTG Leu 45	GTG Val 30 AAG Lys	Ile 15 TCC Ser TTC Phe	GGC Gly ATC	96
Met 1 GTC Val GAG Glu	GTG Val GAG Glu GGC Gly ACC Thr	AGC Ser CTG Leu GAG Glu 35	AAG Lys GAC Asp 20 GGC Gly	GGC Gly 5 GGC Gly GAT Asp	GAG Glu GAC Asp GCC Ala	GAG Glu GTA Val ACC Thr	CTG Leu AAC Asn TAC Tyr .40	TTC Phe  GGC Gly 25  GGC Gly CCC	ACC Thr 10 CAC His AAG Lys	GGG Gly AAG Lys CTG Leu	GTG Val TTC Phe ACC Thr	AGC Ser CTG Leu 45	GTG Val 30 AAG Lys	TCC Ser TTC Phe	GGC Gly ATC Ile	96 144
Met 1 GTC Val GAG Glu	GTG Val GAG Glu GGC Gly	AGC Ser CTG Leu GAG Glu 35	AAG Lys GAC Asp 20 GGC Gly	GGC Gly 5 GGC Gly GAT Asp	GAG Glu GAC Asp GCC Ala	GAG Glu GTA Val ACC Thr	CTG Leu AAC Asn TAC Tyr .40	TTC Phe  GGC Gly 25  GGC Gly CCC	ACC Thr 10 CAC His AAG Lys	GGG Gly AAG Lys CTG Leu	GTG Val TTC Phe ACC Thr	AGC Ser CTG Leu 45	GTG Val 30 AAG Lys	TCC Ser TTC Phe	GGC Gly ATC Ile	96 144
Met 1 GTC Val GAG Glu TGC Cys	GTG Val GAG Glu GGC Gly ACC Thr 50	AGC Ser CTG Leu GAG Glu 35 ACC Thr	AAG Lys GAC Asp 20 GGC Gly GGC	GGC Gly 5 GGC Gly GAT Asp AAG Lys	GAG Glu GAC Asp GCC Ala CTG Leu	GAG Glu GTA Val ACC Thr CCC Pro 55	CTG Leu AAC Asn TAC Tyr 40 GTG Val	TTC Phe GGC Gly 25 GGC Gly CCC Pro	ACC Thr 10 CAC His AAG Lys TGG Trp	GGG Gly  AAG Lys  CTG Leu  CCC Pro	GTG Val TTC Phe ACC Thr ACC Thr 60	AGC Ser CTG Leu 45 CTC Leu	GTG Val 30 AAG Lys GTG Val	TCC Ser TTC Phe ACC Thr	GGC Gly  ATC Ile  ACC Thr	96 144
Met 1 GTC Val GAG Glu TGC Cys	GTG Val GAG Glu GGC Gly ACC Thr 50	AGC Ser CTG Leu GAG Glu 35 ACC Thr	AAG Lys GAC Asp 20 GGC Gly GGC	GGC Gly 5 GGC Gly GAT Asp AAG Lys	GAC Asp GCC Ala CTG Leu	GAG Glu GTA Val ACC Thr CCC Pro 55	CTG Leu AAC Asn TAC Tyr 40 GTG Val	TTC Phe GGC Gly 25 GGC Gly CCC Pro	ACC Thr 10 CAC His AAG Lys TGG Trp	GGG Gly  AAG Lys  CTG Leu  CCC Pro	GTG Val TTC Phe ACC Thr ACC Thr 60	AGC Ser CTG Leu 45 CTC Leu	GTG Val 30 AAG Lys GTG Val	TCC Ser TTC Phe ACC Thr	GGC Gly  ATC Ile  ACC Thr	96 144 192
Met 1 GTC Val GAG Glu TGC Cys CTG Leu 65	GTG Val GAG Glu GGC Gly ACC Thr 50 ACC	AGC Ser CTG Leu GAG Glu 35 ACC Thr	AAG Lys GAC Asp 20 GGC Gly GGC Gly	GGC Gly 5 GGC Gly GAT Asp AAG Lys	GAG Glu GAC Asp GCC Ala CTG Leu CAG Gln 70	GAG Glu GTA Val ACC Thr CCC Pro 55 TGC Cys	CTG Leu AAC Asn TAC Tyr 40 GTG Val	TTC Phe GGC Gly 25 GGC Gly CCC Pro	ACC Thr 10 CAC His AAG Lys TGG Trp	GGG Gly AAG Lys CTG Leu CCC Pro	GTG Val TTC Phe ACC Thr ACC Thr 60 CCC Pro	AGC Ser CTG Leu 45 CTC Leu GAC	GTG Val 30 AAG Lys GTG Val	TTCC Ser TTC Phe ACC Thr ATG Met	GGC Gly  ATC Ile  ACC Thr  AAG Lys	96 144 192
Met 1 GTC Val GAG Glu TGC Cys CTG Leu 65 CAG	GTG Val GAG Glu GGC Gly ACC Thr 50 ACC Thr	AGC Ser CTG Leu GAG Glu 35 ACC Thr TAC Tyr GAC	AAG Lys GAC Asp 20 GGC Gly GGC Gly	GGC Gly 5 GGC Gly GAT Asp AAG Lys GTG Val	GAG Glu GAC Asp GCC Ala CTG Leu CAG Gln	GAG Glu GTA Val ACC Thr CCC Pro 55 TGC Cys	CTG Leu AAC Asn TAC Tyr 40 GTG Val	TTC Phe  GGC Gly 25 GGC Gly CCC Pro  AGC Ser	ACC Thr 10 CAC His AAG Lys TGG Trp CGC Arg	GGG Gly  AAG Lys  CTG Leu  CCC Pro  TAC Tyr 75  GAA	GTG Val TTC Phe ACC Thr ACC Thr 60 CCC Pro	AGC Ser CTG Leu 45 CTC Leu GAC Asp	GTG Val 30 AAG Lys GTG Val CAC His	TTCC Ser TTC Phe ACC Thr ATG Met CAG Gln	GGC Gly  ATC Ile  ACC Thr  AAG Lys 80  GAG	96 144 192 240
Met 1 GTC Val GAG Glu TGC Cys CTG Leu 65 CAG	GTG Val GAG Glu GGC Gly ACC Thr 50 ACC Thr	AGC Ser CTG Leu GAG Glu 35 ACC Thr TAC Tyr GAC	AAG Lys GAC Asp 20 GGC Gly GGC Gly	GGC Gly 5 GGC Gly GAT Asp AAG Lys GTG Val	GAG Glu  GAC Asp  GCC Ala  CTG Leu  CAG Gln 70  AAG	GAG Glu GTA Val ACC Thr CCC Pro 55 TGC Cys	CTG Leu AAC Asn TAC Tyr 40 GTG Val	TTC Phe  GGC Gly 25 GGC Gly CCC Pro  AGC Ser	ACC Thr 10 CAC His AAG Lys TGG Trp	GGG Gly  AAG Lys  CTG Leu  CCC Pro  TAC Tyr 75  GAA	GTG Val TTC Phe ACC Thr ACC Thr 60 CCC Pro	AGC Ser CTG Leu 45 CTC Leu GAC Asp	GTG Val 30 AAG Lys GTG Val CAC His	TTCC Ser TTCC Phe ACC Thr ATG Met CAG	GGC Gly  ATC Ile  ACC Thr  AAG Lys 80  GAG	96 144 192 240
Met 1 GTC Val GAG Glu TGC Cys CTG Leu 65 CAG Gln CGC	GTG Val GAG Glu GGC Gly ACC Thr 50 ACC Thr	AGC Ser CTG Leu GAG Glu 35 ACC Thr TAC Tyr GAC Asp	AAG Lys GAC Asp 20 GGC Gly GGC Gly TTC Phe	GGC Gly 5 GGC Gly GAT Asp AAG Lys GTG Val	GAG Glu  GAC Asp  GCC Ala  CTG Leu  CAG Gln 70  AAG	GAG Glu GTA Val ACC Thr CCC Pro 55 TGC Cys	CTG Leu AAC Asn TAC Tyr 40 GTG Val TTC Phe	TTC Phe  GGC Gly 25 GGC Gly  CCC Pro  AGC Ser  ATG Met	ACC Thr 10 CAC His AAG Lys TGG Trp CGC Arg	GGG Gly  AAG Lys  CTG Leu  CCC Pro  TAC Tyr  75  GAA Glu  TAC	GTG Val TTC Phe ACC Thr 60 CCC Pro GGC Gly	AGC Ser CTG Leu 45 CTC Leu GAC Asp TAC TYr	GTG Val 30 AAG Lys GTG Val CAC His GTC Val CGC	TTCC Serr TTCC Phe ACCC Thr ATG Met CAG Gln 95 GCC	GGC Gly  ATC Ile  ACC Thr  AAG Lys 80  GAG Glu  GAG	96 144 192 240

					GAC Asp											384
	-				GAC Asp					_						432
					AAC Asn 150											480
					TTC Phe											528
					CAC His										_	576
					GAC Asp									_		624
					GAG Glu											672
					ATC Ile 230											720
					GAG Glu											768
					GCA Ala											816
					CGG Arg											864
					GGG Gly					_	_	Arg	_		AAC Asn	912
															CTG Leu 320	960
												_		_	GAC Asp	1008
CGC	GAC	GGC	ACC	ATC	ATC	CAC	CTC	AAG	TAC	CCG	CTG	AAC	TGC	TCC	GAT	1056

	_	_						_	_	_	_	_	_	_	_	_	
	Arg	Asp	GIA		IIe	He	His	Leu		Tyr	Pro	Leu	Asn		Ser	Asp	
				340					345					350			
	CCC	АСТ	AGT	GAG	AGG	TGG	TAC	САТ	GGC	CAC	ATG	тст	GGC	GGG	CAG	GCA	1104
															Gln		
			355			-	-	360	_				365	_			
															GTG		1152
	Glu		Leu	Leu	Gln	Ala	_	Gly	Glu	Pro	Trp		Phe	Leu	Val	Arg	
		370					375					380					
	GAG	AGC	СТС	AGC	CAG	ССТ	GGA	GAC	TTC	GTG	СТТ	тст	GTG	CTC	AGT	GAC	1200
					_		_		_	_			_		Ser		
	385					390	_	_			395					400	
												_	_	_	ATC		1248
	Gln	Pro	Lys	Ala	_	Pro	Gly	Ser	Pro		Arg	val	Thr	His	Ile	Lys	
					405					410					415		
	GTC	ATG	TGC	GAG	GGT	GGA	CGC	TAC	ACA	GTG	GGT	GGT	TTG	GAG	ACC	TTC	1296
															Thr		
			_	420	_	_	_	-	425		_	-		430			
														_	ATT	_	1344
	Asp	Ser		Thr	Asp	Leu	Vai	G1u 440	His	Phe	Lys	Lys	1nr 445	GIY	Ile	GIu	
			435					440					447				
	GAG	GCC	TCA	GGC	GCC	TTT	GTC	TAC	CTG	CGG	CAG	CCG	TAC	TAT	GCC	ACG	1392
											_				Ala	_	
		450					455					460				•	
													_		AAC		1440
	465	vaı	ASII	мта	AIA	470	TIE	GIU	ASII	ALG	475	Leu	GIU	Leu	Asn	480	
	AAG	CAG	GAG	TCC	GAG	GAT	ACA	GCC	AAG	GCT	GGC	TTC	TGG	GAG	GAG	TTT	1488
	Lys	Gln	Glu	Ser		Asp	Thr	Ala	Lys		Gly	Phe	$\operatorname{Trp}$	Glu	Glu	Phe	
					485					490					495		
	CAC	y Can	dan	CNC	አላር	CAC	CAC	Cav	מ מ	מ מ	<b>₩</b>	$C_{N}C$	CAC	CCm	CTG	GAN	1536
															Leu		1930
-	010	JUL	200	500	_y 3	I	CIU		505					510			
															ATT		1584
	Gly	Gln	_	Pro	Glu	Asn	Lys	_	Lys	Asn	Arg	Tyr		Asn	Ile	Leu	
			515					520					525				
	רככ	ДАДАДЬ	GAC	CAC	AGC	CGA	GTC	ንጥል	CITC	CAG	GGA	CGG	GAC	AGT	AAC	ATC	1632
															Asn		
		530	- 1			- 3	535				-	540	•				
														_	CTG		1680
		Gly	Ser	Asp	Tyr		Asn	Ala	Asn	Tyr		Lys	Asn	Gln	Leu		
	545					550					555					560	
	GGC	ССТ	GAT	GAG	AAC	GCT	AAG	ACC	TAC	ATC	GCC	AGC	CAG	GGC	TGT	CTG	1728
													_		Cys		
	_		-		565					570					575		

3

										ATG Met							1776		
										GTG Val							1824		
										ATG Met							1872		
										GAC Asp							1920		
										AAT Asn 650							1968		
										CCC Pro							2016		
										GAC Asp							2064		
										ATC Ile							2112	•	
										ATC Ile							2160		
										ATT Ile 730							2208		
•										ATG Met							2256		
										CAG Gln							2304		
										AAG Lys							2352		
					Tyr					AAG Lys							2400		
	TCC	CGC	ACC	TCG	TCC	AAA	CAC	AAG	GAG	GAT	GTG	TAT	GAG	AAC	CTG	CAC	2448		

.

Ser Arg Thr Ser Ser Lys His Lys Glu Asp Val Tyr Glu Asn Leu His 805 810 815

ACT AAG AAC AAG AGG GAG GAG AAA GTG AAG AAG CAG CGG TCA GCA GAC

Thr Lys Asn Lys Arg Glu Glu Lys Val Lys Lys Gln Arg Ser Ala Asp

820

825

830

AAG GAG AAG AGC AAG GGT TCC CTC AAG AGG AAG TGA Lys Glu Lys Ser Lys Gly Ser Leu Lys Arg Lys 835 840 2532

#### (2) INFORMATION FOR SEQ ID NO:117:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 843 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:117:

 Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu

 1
 5
 10
 15

 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly
 20
 25
 30

 Glu Gly Gly Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile
 10
 15

Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 35 40 45

Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 50 55 60

Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 65 70 75 80 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu

85 90 95
Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu

arg Thr lie Phe Phe Lys Asp Asp Gly Ash Tyr Lys Thr Arg Ala Glu
100 105 110

Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly
115 120 125

Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 130 135 140

Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 145 150 155 160

Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 175

Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 180 185 190

Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205

Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 220

Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 225 230 235 240

Gly Leu Arg Ser Arg Glu Met Leu Ser Arg Gly Trp Phe His Arg Asp 245 250 255

Leu Ser Gly Leu Asp Ala Glu Thr Leu Leu Lys Gly Arg Gly Val His

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260
                             265
Gly Ser Phe Leu Ala Arg Pro Ser Arg Lys Asn Gln Gly Asp Phe Ser
                        280
Leu Ser Val Arg Val Gly Asp Gln Val Thr His Ile Arg Ile Gln Asn
                    295
                                        300
Ser Gly Asp Phe Tyr Asp Leu Tyr Gly Gly Glu Lys Phe Ala Thr Leu
                 310
                                    315
Thr Glu Leu Val Glu Tyr Tyr Thr Gln Gln Gln Gly Val Leu Gln Asp
                                330
             325
Arg Asp Gly Thr Ile Ile His Leu Lys Tyr Pro Leu Asn Cys Ser Asp
          340
                             345
Pro Thr Ser Glu Arg Trp Tyr His Gly His Met Ser Gly Gln Ala
                         360
Glu Thr Leu Leu Gln Ala Lys Gly Glu Pro Trp Thr Phe Leu Val Arg
                     375
                              380
Glu Ser Leu Ser Gln Pro Gly Asp Phe Val Leu Ser Val Leu Ser Asp
                                  395
                390
Gln Pro Lys Ala Gly Pro Gly Ser Pro Leu Arg Val Thr His Ile Lys
             405
                               410
Val Met Cys Glu Gly Gly Arg Tyr Thr Val Gly Gly Leu Glu Thr Phe
        420
                             425
Asp Ser Leu Thr Asp Leu Val Glu His Phe Lys Lys Thr Gly Ile Glu
                        440
Glu Ala Ser Gly Ala Phe Val Tyr Leu Arg Gln Pro Tyr Tyr Ala Thr
                   455
                                        460
Arg Val Asn Ala Ala Asp Ile Glu Asn Arg Val Leu Glu Leu Asn Lys
                  470
                                    475
Lys Gln Glu Ser Glu Asp Thr Ala Lys Ala Gly Phe Trp Glu Glu Phe
                                490
              485
Glu Ser Leu Gln Lys Gln Glu Val Lys Asn Leu His Gln Arg Leu Glu
                             505
Gly Gln Arg Pro Glu Asn Lys Gly Lys Asn Arg Tyr Lys Asn Ile Leu
                         520
                                          525
Pro Phe Asp His Ser Arg Val Ile Leu Gln Gly Arg Asp Ser Asn Ile
                     535
                                       540
Pro Gly Ser Asp Tyr Ile Asn Ala Asn Tyr Ile Lys Asn Gln Leu Leu
        550
                                    555
Gly Pro Asp Glu Asn Ala Lys Thr Tyr Ile Ala Ser Gln Gly Cys Leu
                                 570
Glu Ala Thr Val Asn Asp Phe Trp Gln Met Ala Trp Gln Glu Asn Ser
         580
                            585
Arg Val Ile Val Met Thr Thr Arg Glu Val Glu Lys Gly Arg Asn Lys
                        600
Cys Val Pro Tyr Trp Pro Glu Val Gly Met Gln Arg Ala Tyr Gly Pro
            615
                                        620
Tyr Ser Val Thr Asn Cys Gly Glu His Asp Thr Thr Glu Tyr Lys Leu
        630
                          635
Arg Thr Leu Gln Val Ser Pro Leu Asp Asn Gly Asp Leu Ile Arg Glu
            645
                                650
Ile Trp His Tyr Gln Tyr Leu Ser Trp Pro Asp His Gly Val Pro Ser
                             665
Glu Pro Gly Gly Val Leu Ser Phe Leu Asp Gln Ile Asn Gln Arg Gln
                          680
                                            685
Glu Ser Leu Pro His Ala Gly Pro Ile Ile Val His Cys Ser Ala Gly
                                        700
                     695
Ile Gly Arg Thr Gly Thr Ile Ile Val Ile Asp Met Leu Met Glu Asn
                 710
                                    715
Ile Ser Thr Lys Gly Leu Asp Cys Asp Ile Asp Ile Gln Lys Thr Ile
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				725					730					735		
Gln	Met	Val	Arg 740	Ala	Gln	Arg	Ser	Gly 745	Met	Val	Gln	Thr	Glu 750	Ala	Gln	
Tyr	Lys	Phe 755	Ile	Tyr	Val	Ala	Ile 760	Ala	Gln	Phe	Ile	Glu 765	Thr	Thr	Lys	
Lys	Lys 770	Leu	Glu	Val	Leu	G1n 775	Ser	Gln	Lys	Gly	Gln 780	Glu	Ser	Glu	Tyr	
_	Asn	Ile	Thr	Tyr		Pro	Ala	Met	Lys		Ala	His	Ala	Lys		
785	_	_	_	_	790		_		_	795	_				800	
Ser	Arg	Thr	Ser		Lys	His	гуs	GIu	Asp	Vai	ıyr	GIU	Asn	Leu 815	HIS	
Thr	Lys	Asn	Lys 820	805 Arg	Glu	Glu	Lys	Val 825	810 Lys	Lys	Gln	Arg	Ser 830		Asp	
Lys	Glu	Lys 835	Ser	Lys	Gly	Ser	Leu 840	Lys	Arg	Lys						
		(2)	INE	ORMA	TION	ı FOF	R SEX	O ID	NO:1	L18:						
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		(D)	TOPC	)LCG		inear	•									
			OLEX TEAT		TYPI	E: cI	ONA									
				<b>6</b> 777	70. /		C									
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					INFO											
		(2)	011													
	(:	xi) :	SEQUI	ENCE	DES	CRIP	rion	: SE	Q ID	NO:	118:					
N TO C	CTTC	TCC	CCT	ccc	TYCC:	ינואנואנוי	CAC	CCA	GAC	CTC	እርጥ	ccc	CTC	СУТ	GC A	48
									Asp						_	-10
1	Deu	502		5					10			2		15		
GAG	ACC	CTG	CTC	AAG	GGC	CGA	GGT	GTC	CAC	GGT	AGC	TTC	CTG	GCT	CGG	96
									His							
			20	-	_	_	_	25					30			
									TCG							144
Pro	Ser	_	Lys	Asn	Gln	Gly	_	Phe	Ser	Leu	Ser		Arg	Val	Gly	
		35					40					45				
CAT	CAG	GTG	ACC	САТ	ΑͲͲ	CGG	АТС	CAG	AAC	TCA	GGG	GAT	TTC	TAT	GAC	192
									Asn							
-	50					55					60					
											~~~	ome.				240
									CTG							240
ьеu 65	туг	GIĀ	GIÀ	GIU	ьуs 70	Pne	Ala	THE	Leu	75	GIU	Leu	vaı	Giu	80	
0,5					, 0					, ,					00	
TAC	ACT	CAG	CAG	CAG	GGT	GTC	CTG	CAG	GAC	CGC	GAC	GGC	ACC	ATC	ATC	288
									Asp							
-				85	="				90					95		
										~~~	3.00	3.00	GAG	300		336

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His	Leu	Lys	Tyr 100	Pro	Leu	Asn	Cys	Ser 105	Asp	Pro	Thr	Ser	Glu 110	Arg	Trp	
														CAG Gln		384
														CAG Gln		432
													_	GGC Gly		480
														GGT Gly 175		528
														GAC Asp		576
											_		_	GCC Ala		624
														GCT Ala		672
														GAG Glu		720
														AAG Lys 255		768
														GAG Glu		816
														AGC Ser		864
														TAC Tyr		912
														AAC Asn		960
														AAT Asn 335		1008

TTC TGG CAG ATG GCG TGG CAG GAG AAG Phe Trp Gln Met Ala Trp Gln Glu Ass 340	n Ser Arg Val Ile Val Met Thr
ACC CGA GAG GTG GAG AAA GGC CGG AAC Thr Arg Glu Val Glu Lys Gly Arg Ass 355 360	
GAG GTG GGC ATG CAG CGT GCT TAT GGC Glu Val Gly Met Gln Arg Ala Tyr Gly 370	
GGG GAG CAT GAC ACA ACC GAA TAC AA Gly Glu His Asp Thr Thr Glu Tyr Ly: 385 390	
CCG CTG GAC AAT GGA GAC CTG ATT CCC Pro Leu Asp Asn Gly Asp Leu Ile Arg 405	
CTG AGC TGG CCC GAC CAT GGG GTC CCC Leu Ser Trp Pro Asp His Gly Val Pro 420	o Ser Glu Pro Gly Gly Val Leu
AGC TTC CTG GAC CAG ATC AAC CAG CCC Ser Phe Leu Asp Gln Ile Asn Gln Arc 435	
GGG CCC ATC ATC GTG CAC TGC AGC GCC Gly Pro Ile Ile Val His Cys Ser Ala 450	
ATC ATT GTC ATC GAC ATG CTC ATG GAG  Ile Ile Val Ile Asp Met Leu Met Glu  465	
GAC TGT GAC ATT GAC ATC CAG AAG ACC Asp Cys Asp Ile Asp Ile Gln Lys Tho 485	
CGC TCG GGC ATG GTG CAG ACG GAG GCC Arg Ser Gly Met Val Gln Thr Glu Ala 500	a Gln Tyr Lys Phe Ile Tyr Val
GCC ATC GCC CAG TTC ATT GAA ACC ACT Ala Ile Ala Gln Phe Ile Glu Thr Thr 515 520	
CAG TCG CAG AAG GGC CAG GAG TCG GAG Gln Ser Gln Lys Gly Gln Glu Ser Glu 530	
CCA GCC ATG AAG AAT GCC CAT GCC AAG Pro Ala Met Lys Asn Ala His Ala Lys 545 550	
CAC AAG GAG GAT GTG TAT GAG AAC CTG	G CAC ACT AAG AAC AAG AGG GAG 1728

His Lys	Glu Asp	Val Tyr 565	Glu A	sn Leu	His 570	Thr	Lys	Asn	Lys	Arg 575	Glu	
		AAG CAG Lys Gln										1776
		AAG CGA Lys Arg	Ile L									1824
		GCC ACC										1872
		ATC CTG Ile Leu 630	Val G									1920
		TCC GGC Ser Gly 645										1968
		TTC ATC										2016
		ACC ACC	Leu T									2064
		ATG AAG Met Lys										2112
		CAG GAG Gln Glu 710	Arg T									2160
		GCC GAG Ala Glu 725										2208
		AAG GGC Lys Gly										2256
		GAG TAC	Asn T									2304
		AAG AAC Lys Asn										2352
		GGC AGC Gly Ser 790	Val G									2400

ACC CCC Thr Pro	 		 	 	 	 2448
AGC ACC Ser Thr					 	 2496
ATG GTC Met Val	 	Val T	 	 	 	 2544
GAC GAG Asp Glu 850	 					2562

#### (2) INFORMATION FOR SEQ ID NO:119:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 853 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
  (v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:119:

Met Leu Ser Arg Gly Trp Phe His Arg Asp Leu Ser Gly Leu Asp Ala 5 10 Glu Thr Leu Leu Lys Gly Arg Gly Val His Gly Ser Phe Leu Ala Arg 25 Pro Ser Arg Lys Asn Gln Gly Asp Phe Ser Leu Ser Val Arg Val Gly 40 Asp Gln Val Thr His Ile Arg Ile Gln Asn Ser Gly Asp Phe Tyr Asp 55 60 Leu Tyr Gly Gly Glu Lys Phe Ala Thr Leu Thr Glu Leu Val Glu Tyr 70 75 Tyr Thr Gln Gln Gln Gly Val Leu Gln Asp Arg Asp Gly Thr Ile Ile 85 90 His Leu Lys Tyr Pro Leu Asn Cys Ser Asp Pro Thr Ser Glu Arg Trp 100 105 110 Tyr His Gly His Met Ser Gly Gly Gln Ala Glu Thr Leu Leu Gln Ala 115 120 125 Lys Gly Glu Pro Trp Thr Phe Leu Val Arg Glu Ser Leu Ser Gln Pro 130 135 140 Gly Asp Phe Val Leu Ser Val Leu Ser Asp Gln Pro Lys Ala Gly Pro 145 150 155 Gly Ser Pro Leu Arg Val Thr His Ile Lys Val Met Cys Glu Gly Gly 165 170 175 Arg Tyr Thr Val Gly Gly Leu Glu Thr Phe Asp Ser Leu Thr Asp Leu 180 185 190 Val Glu His Phe Lys Lys Thr Gly Ile Glu Glu Ala Ser Gly Ala Phe 200 205 Val Tyr Leu Arg Gln Pro Tyr Tyr Ala Thr Arg Val Asn Ala Ala Asp

210		215					220				
Ile Glu As 225	sn Arg Val	Leu Glu 230	Leu	Asn	Lys	Lys 235	Gln	Glu	Ser	Glu	Asp 240
Thr Ala Ly	245	i			250					255	
Glu Val Ly	260			265					270		
	75		280					285			
Val Ile Lo 290		295					300				
Asn Ala As 305	_	310				315					320
Lys Thr T	325	•			330					335	
Phe Trp G	340			345					350		
	55	-	360					365			
Glu Val G		375					380				
Gly Glu H	•	390	_	_		395					400
Pro Leu A	405	5			410					415	
Leu Ser T	420			425					430		
	35		440					445			
Gly Pro I		455	,				460			_	
Ile Ile V	_	470				475					480
Asp Cys A	485	5			490					495	
Arg Ser G	500			505					510		
_	15		520					525			
Gln Ser G		535	,				540				
Pro Ala M 545		550				555					560
His Lys G	565	5			570					575	
Glu Lys V	580			585					590		
	95		600					605			
Asp Pro P		615	,				620				
Gly Val V		630				635					640
Lys Phe S	645	5	_		650				_	655	
Leu Thr L	660	_		665	_	-			670		
Pro Thr L	eu Val Thr	Thr Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg

		675					680					685			
Tyr	Pro 690	Asp	His	Met	Lys	Gln 695	His	Ąsp	Phe	Phe	Lys 700	Ser	Ala	Met	Pro
Glu 705	Gly	Tyr	Val	Gln	Glu 710	Arg	Thr	Ile	Phe	Phe 715	Lys	Asp	Asp	Gly	Asn 720
Tyr	Lys	Thr	Arg	Ala 725	Glu	Val	Lys	Phe	Glu 730	Gly	Asp	Thr	Leu	Val 735	Asn
Arg	Ile	Glu	Leu 740	Lys	Gly	Ile	Asp	Phe 745	Lys	Glu	Asp	Gly	Asn 750	Ile	Leu
Gly	His	Lys 755	Leu	Glu	Tyr	Asn	Tyr 760	Asn	Ser	His	Asn	Val 765	Tyr	Ile	Met
Ala	Asp 770	Lys	Gln	Lys	Asn	Gly 775	Ile	Lys	Val	Asn	Phe 780	Lys	Ile	Arg	His
Asn 785	Ile	Glu	Asp	Gly	Ser 790	Val	Gln	Leu	Ala	Asp 795	His	Tyr	Gln	Gln	Asn 800
Thr	Pro	Ile	Gly	Asp 805	Gly	Pro	Val	Leu	Leu 810	Pro	Asp	Asn	His	Tyr 815	Leu
Ser	Thr	Gln	Ser 820	Ala	Leu	Ser	Lys	Asp 825	Pro	Asn	Glu	Lys	Arg 830	Asp	His
Met	Val	Leu 835	Leu	Glu	Phe	Val	Thr 840	Ala	Ala	Gly	Ile	Thr 845	Leu	Gly	Met
Asp	Glu 850	Leu	Tyr	Lys											

## (2) INFORMATION FOR SEQ ID NO:120:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2994 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2991
  - (D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:120:

				GGC Gly 5												48
	-			GGC Gly												96
				GAT Asp												144
				AAG Lys												192
CTG	ACC	TAC	GGC	GTG	CAG	TGC	TTC	AGC	CGC	TAC	CCC	GAC	CAC	ATG	AAG	240

Leu 65	Thr	Tyr	Gly	Val	Gln 70	Cys	Phe	Ser	Arg	Tyr 75	Pro	Asp	His	Met	Lys 80		
				TTC Phe 85												288	
				TTC Phe												336	
				GGC Gly												384	
				GAG Glu												432	
				CAC His												480	
				AAC Asn 165												528	
				GAC Asp												576	
				CCC Pro										_		624	
				AAC Asn												672	
				GGG Gly												720	
				CGA Arg 245												768	
				CCG Pro												816	
				GGC Gly												864	
				ATA Ile												912	

					TGG Trp 310											960
					GTA Val											1008
					GTG Val											10 <u>5</u> 6
					CTG Leu											1104
					CTT Leu											1152
					AAC Asn 390											1200
					GAT Asp											1248
					AAA Lys											1296
Phe	Val	Gly 435	Thr	Leu	CAG Gln	Tyr	Leu 440	Ala	Pro	Glu	Leu	Phe 445	Glu	Asn	Lys	1344
					GTT Val										TTT	1392
	Cys														Phe 480	1440
					Ile					Pro					GCA	1488
									Phe					Pro	CAA Gln	1536
			Leu					Val					Asn		CTA Leu	1584
CAG	TTG	ATG	TTG	TAA	TGG	GAC	CCT	CAG	CAG	AGA	GGA	GGA	CCI	GT1	GAC	1632

Gln	Leu 530	Met	Leu	Asn	Trp	Asp 535	Pro	Gln	Gln	Arg	Gly 540	Gly	Pro	Val	Asp	
	_								GTA Val							1680
						_			ATG Met 570							1728
									CTT Leu	_						1776
									GGT Gly							1824
									AAA Lys							1872
									TAT Tyr		_					1920
									TTT Phe 650							1968
-									AGC Ser							2016
	-								GCA Ala							2064
									CAG Gln							2112
									TTA Leu							2160
									GCT Ala 730							2208
									TAC Tyr							2256
									GCA Ala							2304

						_			GTC Val							2352
					_	_	_	_	ATG Met	_						2400
									GAA Glu 810							2448
									AGA Arg							2496
									ATT Ile							2544
									GGT Gly							2592
									CTC Leu							2640
									ACT Thr 890							2688
									CTT Leu							2736
									AGT Ser		_	_	_	_	_	2784
									ACT Thr			_				2832
									GAT Asp						_	2880
									GGC Gly 970							2928
		_		_	_	_	_		AGT Ser							2976
AGT	TGG	TTA	ACA	GAA	TGA											2994

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Ser Trp Leu Thr Glu 995

# (2) INFORMATION FOR SEQ ID NO:121:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 997 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:121:

	_						_						_		_
1			-	5					10	_			Pro	15	
Val	Glu	Leu	Asp 20	Gly	Asp	Val	Asn	Gly 25	His	Lys	Phe	Ser	Val 30	Ser	Gly
Glu	Gly	Glu 35	Gly	Asp	Ala	Thr	Tyr 40	Gly	Lys	Leu	Thr	Leu 45	Lys	Phe	Ile
Cys	Thr 50	Thr	Gly	Lys	Leu	Pro 55	Val	Pro	Trp	Pro	Thr 60	Leu	Val	Thr	Thr
Leu 65		Tyr	Gly	Val	Gln 70	Cys	Phe	Ser	Arg	Тут 75	Pro	Asp	His	Met	Lys 80
	His	Asp	Phe	Phe 85		Ser	Ala	Met	Pro 90	Glu	Gly	Tyr	Val	Gln 95	Glu
Arg	Thr	Ile	Phe	Phe	Lys	Asp	Asp	Gly 105	Asn	Тут	Lys	Thr	Arg 110	Ala	Glu
Val	Lys	Phe 115		Gly	Asp	Thr	Leu 120		Asn	Arg	Ile	Glu 125	Leu	Lys	Gly
Ile	Asp		Lys	Glu	Asp	Gly 135	Asn	Ile	Leu	Gly	His 140	Lys	Leu	Glu	Tyr
Asn 145	Tyr	Asn	Ser	His	Asn 150	Val	Tyr	Ile	Met	Ala 155	Asp	Lys	Gln	Lys	Asn 160
	Ile	Lys	Val	Asn 165	Phe	Lys	Ile	Arg	His 170	Asn	Ile	Glu	Asp	Gly 175	Ser
Val	Gln	Leu	Ala 180	Asp	His	Tyr	Gln	Gln 185	Asn	Thr	Pro	Ile	Gly 190	Asp	Gly
Pro	Val	Leu 195	Leu	Pro	Asp	Asn	His 200	Tyr	Leu	Ser	Thr	Gln 205	Ser	Ala	Leu
Ser	Lys 210	Asp	Pro	Asn	Glu	Lys 215	Arg	Asp	His	Met	Val 220	Leu	Leu	Glu	Phe
Val 225		Ala	Ala	Gly	Ile 230	Thr	Leu	Gly	Met	Asp 235	Glu	Leu	Tyr	Lys	Ser 240
Gly	Leu	Arg	Ser	Arg 245	Ala	Gln	Ala	Ser	Asn 250	Ser	Thr	Met	Glu	Arg 255	Pro
Pro	Gly	Leu	Arg 260	Pro	Gly	Ala	Gly	Gly 265	Pro	Trp	Glu	Met	Arg 270	Glu	Arg
Leu	Gly	Thr 275	Gly	Gly	Phe	Gly	Asn 280	Val	Cys	Leu	Tyr	Gln 285	His	Arg	Glu
Leu	Asp 290	Leu	Lys	Ile	Ala	Ile 295	Lys	Ser	Cys	Arg	Leu 300	Glu	Leu	Ser	Thr
Lys 305		Arg	Glu	Arg	Trp 310	Cys	His	Glu	Ile	Gln 315	Ile	Met	Lys	Lys	Leu 320
		Ala	Asn	Val		Lys	Ala	Cys	Asp	Val	Pro	Glu	Glu	Leu	Asn

				325					330					335	
Ile	Leu	Ile	His 340	Asp	Val	Pro	Leu	Leu 345	Ala	Met	Glu	Tyr	Cys 350		Gly
Gly	Asp	Leu 355	Arg	Lys	Leu	Leu	Asn 360	Lys	Pro	Glu	Asn	Cys 365	Cys	Gly	Leu
Lys	Glu 370	Ser	Gln	Ile	Leu	Ser 375	Leu	Leu	Ser	Asp	Ile 380	Gly	Ser	Gly	Ile
Arg 385	Tyr	Leu	His	Glu	Asn 390	Lys	Ile	Ile	His	Arg 395	Asp	Leu	Lys	Pro	Glu 400
Asn	Ile	Val	Leu	Gln 405	Asp	Val	Gly	Gly	Lys 410	Ile	Ile	His	Lys	Ile 415	Ile
Asp	Leu	Gly	Tyr 420	Ala	Lys	Asp	Val	Asp 425	Gln	Gly	Ser	Leu	Cys 430	Thr	Ser
		435				-	440		Pro			445			-
Pro	Tyr 450	Thr	Ala	Thr	Val	Asp 455		Trp	Ser	Phe	Gly 460		Met	Val	Phe
465	-			_	470				Leu	475					480
	-			485		_	_		Asp 490					495	
			500					505					510		
		515					520					525			Leu
	530				_	535				_	540	_		_	Asp
545			_		550					555					560
		_		565					Met 570					575	
			580					585	Leu			_	590		
		595					600		Gly		_	605	_	•	
	610	_				615			Lys		620			_	
625	_	_			630				Phe	635					Asp 640 Ser
				645					650					655	Ile
-	-		660	_				665					670		Gly
		675					680					685			Met
	690					695					700				Thr
705				_	710					715	_		-		720
				725				_	730					735	
_			740					745				_	750		Tyr
_		755			_		760					765			Glu
-	770			-		775		_			780	_			Asp
GIN	тте	met	ser	ьeu	HIS	ATA	GIU	тте	mec	GIÀ	ьeu	GIN	ьys	ser	Pro

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785	7	790		795		800
Tyr Gly Arg	Arg Gln G 805	Sly Asp Leu	Met Glu 810	Ser Leu Glu	Gln Arg 815	Ala
Ile Asp Leu	Tyr Lys G 820	In Leu Lys	His Arg 825	Pro Ser Asp	His Ser 830	Tyr
Ser Asp Ser 835	Thr Glu M	1et Val Lys 840		Val His The 845		Ser
Gln Asp Arg 850	Val Leu L	Lys Glu Lev 855	Phe Gly	His Leu Ser 860	Lys Leu	Leu
Gly Cys Lys 865	-	le Ile Asp 370	Leu Leu	Pro Lys Val	Glu Val	Ala 880
Leu Ser Asn	Ile Lys G 885	Glu Ala Asg	Asn Thr 890	Val Met Phe	Met Gln 895	Gly
Lys Arg Gln	Lys Glu I 900	le Trp His	Leu Leu 905	Lys Ile Ala	Cys Thr 910	Gln
Ser Ser Ala 915	Arg Ser L	Leu Val Gly 920		Leu Glu Gly 925		Thr
Pro Gln Thr 930	Ser Ala T	Orp Leu Pro 935	Pro Thr	Ser Ala Glu 940	His Asp	His
Ser Leu Ser 945	-	/al Thr Pro 950	Gln Asp	Gly Glu Thu 955	Ser Ala	Gln 960
Met Ile Glu	Glu Asn L 965	Leu Asn Cys	Leu Gly 970	His Leu Ser	Thr Ile 975	Ile
His Glu Ala	Asn Glu G 980	Glu Gln Gly	/ Asn Ser 985	Met Met Ası	Leu Asp 990	Trp
Ser Trp Leu 995	Thr Glu					

## (2) INFORMATION FOR SEQ ID NO:122:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2991 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2988
  - (D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:122:

										GCG Ala						48
										GGG Gly						96
										ATT Ile						144
GAG	СТА	AGT	ACC	AAA	AAC	AGA	GAA	CGA	TGG	TGC	CAT	GAA	ATC	CAG	TTA	192

Glu	Leu 50	Ser	Thr	Lys	Asn	Arg 55	Glu	Arg	Trp	Cys	His 60	Glu	Ile	Gln	Ile		
					CAT His 70											240	
				_	TTG Leu				_							288	
					GAT Asp											336	
					GAA Glu											384	
					TAT Tyr											432	
					ATA Ile 150											480	
					CTG Leu						_					528	
					GTG Val				_			_				576	
					TAC Tyr				_							624	
					TGT Cys											672	
					TGG Trp 230	_			_							720	J
					GAA Glu											768	į
					AAT Asn									Pro		816	j
															GGA Gly	864	t

									CCA Pro							912
									CAC His							960
									CCT Pro 330							1008
	-								GGA Gly							1056
									CTG Leu							1104
									GGC Gly							1152
									ТАТ Туг							1200
									ATT Ile 410						ATA Ile	1248
									GTG Val						CAC His	1296
															CAA Gln	1344
															AAA Lys	1392
															TTG Leu 480	1440
															GAG Glu	1488
														Trp	AAA Lys	1536
GAA	ATG	GAA	GAA	AAG	GCC	ATC	CAC	TAT	GCT	GAG	GTT	GGT	GTC	TTA	GGA	1584

Glu	Met G 5	lu G 515	lu I	ùys .	Ala	Ile	His 520	Tyr	Ala	Glu	Val	Gly 525	Val	Ile	Gly	
	CTG G Leu G 530				Ile						_					1632
	AAG A Lys S				_			_	_					_	_	1680
	CAG C		la :													1728
	CAC T	Ser T														1776
	GTG C															1824
	AAG 1 Lys I 610															1872
	GAA (															1920
	ATG (		3ly													1968
	TGT A	Thr G														2016
	GCA (															2064
	CAT O His A															2112
	TCA (															2160
	ACT I		Ile							Gln					Met	2208
	CTT (	r qaA							Trp					Arg		2256

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					ACC					_		_		_		_	2304
	Pro	Pro	Val 755	Ala	Thr	Met	Val	<b>Ser</b> 760	Lys	Gly	Glu	Glu	<b>Leu</b> 765	Phe	Thr	Gly	
	GTG	GTG	ccc	ATC	CTG	GTC	GAG	CTG	GAC	GGC	GAC	GTA	AAC	GGC	CAC	AAG	2352
	Val	Val 770	Pro	Ile	Leu	Val	Glu 775	Leu	Asp	Gly	Asp	Val 780	Asn	Gly	His	Lys	
	TTC	AGC	GTG	TCC	GGC	GAG	GGC	GAG	GGC	GAT	GCC	ACC	TAC	GGC	AAG	CTG	2400
	Phe 785	Ser	Val	Ser	Gly	Glu 790	Gly	Glu	Gly	Asp	Ala 795	Thr	Tyr	Gly	Lys	Leu 800	
	ACC	CTG	AAG	TTC	ATC	TGC	ACC	ACC	GGC	AAG	CTG	CCC	GTG	CCC	TGG	CCC	2448
	Thr	Leu	Lys	Phe	Ile 805	Cys	Thr	Thr	Gly	Lys 810	Leu	Pro	Val	Pro	Trp 815	Pro	
	ACC	CTC	GTG	ACC	ACC	CTG	ACC	TAC	GGC	GTG	CAG	TGC	TTC	AGC	CGC	TAC	2496
	Thr	Leu	Val	Thr 820	Thr	Leu	Thr	Tyr	Gly 825	Val	Gln	Cys	Phe	Ser 830	Arg	Tyr	
			-		AAG												2544
	Pro	Asp	His 835	Met	Lys	Gln	His	<b>Asp</b> 840	Phe	Phe	Lys	Ser	Ala 845	Met	Pro	Glu	
	GGC	TAC	GTC	CAG	GAG	CGC	ACC	ATC	TTC	TTC	AAG	GAC	GAC	GGC	AAC	TAC	2592
	Gly	Тут 850	Val	Gln	Glu	Arg	Thr 855	Ile	Phe	Phe	Lys	Asp 860	Asp	Gly	Asn	Tyr	
					GAG												2640
	Lys 865	Thr	Arg	Ala	Glu	Val 870	Lys	Phe	Glu	Gly	875	Thr	Leu	Val	Asn	Arg 880	
					GGC									_		_	2688
	Ile	Glu	Leu	Lys	Gly 885	Ile	Asp	Phe	Lys	Glu 890	Asp	Gly	Asn	Ile	Leu 895	Gly	
					TAC									_		_	2736
	His	Lys	Leu	Glu 900	Tyr	Asn	Tyr	Asn	Ser 905	His	Asn	Val	Tyr	910	Met	Ala	
•					AAC												2784
	Asp	Lys	Gln 915	Lys	Asn	Gly	Ile	Lys 920	Val	Asn	Phe	Lys	925	Arg	His	Asn	
					AGC									_			2832
	Ile	Glu 930	Asp	Gly	Ser	Val	G1n 935	Leu	Ala	Asp	His	Tyr 940	GIn	GIn	Asn	Thr	
					GGC												2880
		Ile	Gly	Asp	Gly		Val	Leu	Leu	Pro	_	Asn	His	Tyr	Leu	Ser 960	
	945					950					955					900	
	ACC	CAG	TCC	GCC	CTG	AGC	AAA	GAC	CCC	AAC	GAG	AAG	CGC	GAT	CAC	ATG	2928
	Thr	Gln	Ser	Ala	Leu 965	Ser	Lys	Asp	Pro	Asn 970	Glu	Lys	Arg	Asp	His 975	Met	
	GTC	CTG	CTG	GAG	TTC	GTG	ACC	GCC	GCC	GGG	ATC	ACT	CTC	GGC	ATG	GAC	2976

Val Leu Clu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp 980 985 990

GAG CTG TAC AAG TAA Glu Leu Tyr Lys 995 2991

### (2) INFORMATION FOR SEQ ID NO:123:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 996 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:123:

Met Glu Arg Pro Pro Gly Leu Arg Pro Gly Ala Gly Gly Pro Trp Glu 1 5 10 Met Arg Glu Arg Leu Gly Thr Gly Gly Phe Gly Asn Val Cys Leu Tyr 25 Gln His Arg Glu Leu Asp Leu Lys Ile Ala Ile Lys Ser Cys Arg Leu 35 40 45 Glu Leu Ser Thr Lys Asn Arg Glu Arg Trp Cys His Glu Ile Gln Ile 50 55 60 Met Lys Lys Leu Asn His Ala Asn Val Val Lys Ala Cys Asp Val Pro 65 70 75 Glu Glu Leu Asn Ile Leu Ile His Asp Val Pro Leu Leu Ala Met Glu 85 90 Tyr Cys Ser Gly Gly Asp Leu Arg Lys Leu Leu Asn Lys Pro Glu Asn 100 105 110 Cys Cys Gly Leu Lys Glu Ser Gln Ile Leu Ser Leu Leu Ser Asp Ile 120 125 Gly Ser Gly Ile Arg Tyr Leu His Glu Asn Lys Ile Ile His Arg Asp 135 140 Leu Lys Pro Glu Asn Ile Val Leu Gln Asp Val Gly Gly Lys Ile Ile 145 150 155 His Lys Ile Ile Asp Leu Gly Tyr Ala Lys Asp Val Asp Gln Gly Ser 165 170 Leu Cys Thr Ser Phe Val Gly Thr Leu Gln Tyr Leu Ala Pro Glu Leu 185 190 Phe Glu Asn Lys Pro Tyr Thr Ala Thr Val Asp Tyr Trp Ser Phe Gly 195 200 205 Thr Met Val Phe Glu Cys Ile Ala Gly Tyr Arg Pro Phe Leu His His 215 220 Leu Gln Pro Phe Thr Trp His Glu Lys Ile Lys Lys Lys Asp Pro Lys 230 235 Cys Ile Phe Ala Cys Glu Glu Met Ser Gly Glu Val Arg Phe Ser Ser 245 250 255 His Leu Pro Gln Pro Asn Ser Leu Cys Ser Leu Ile Val Glu Pro Met 260 265 270 Glu Asn Trp Leu Gln Leu Met Leu Asn Trp Asp Pro Gln Gln Arg Gly 280 285 Gly Pro Val Asp Leu Thr Leu Lys Gln Pro Arg Cys Phe Val Leu Met

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	290					295					300				
Asp 305	His	Ile	Leu	Asn	Leu 310	Lys	Ile	Val	His	Ile 315	Leu	Asn	Met	Thr	Ser 320
Ala	Lys	Ile	Ile	Ser 325	Phe	Leu	Leu	Pro	Pro 330	Asp	Glu	Ser	Leu	His 335	Ser
Leu	Gln	Ser	Arg 340	Ile	Glu	Arg	Glu	Thr 345	Gly	Ile	Asn	Thṛ	Gly 350	Ser	Gln
		355				_	360					365		Pro	
	370					375					380			Met	
385					390					395				Ala	400
				405					410					Lys 415	
			420					425					430	Val	
		435					440					445		Gly	
_	450					455					460			Thr	
Met 465	Lys	Asn	Thr	Leu	11e 470	Ser	Ala	Ser	Gln	G1n 475	Leu	Lys	Ala	Lys	Leu 480
	Phe	Phe	His	Lys 485		Ile	Gln	Leu	Asp 490	-	Glu	Arg	Tyr	Ser 495	
Gln	Met	Thr	Tyr 500	Gly	Ile	Ser	Ser	Glu 505	Lys	Met	Leu	Lys	Ala 510	Trp	Lys
		515		-			520					525		Ile	
_	530					535					540				Leu
545					550					555					Leu 560
				565					570					575	
_			580					585					590		His
		595					600					605			Leu
	610					615					620				Lys
625	GIU	vaı	Ата	Leu	630	ASII	116	гур	Giu	635		ASII	. 1111	vai	Met 640
	Met	Gln	Gly	Lys 645		Gln	Lys	Glu	Ile 650		His	Leu	Leu	Lys 655	Ile
	-		660					665					670		Glu
		675					680					685			Ala
	690					695					700				Glu
705					710					715					720
				725					730					735	
			740					745					750	)	Asp
Pro	Pro	Val	Ala	Thr	Met	Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly

		755					760					765			
Val	Val 770	Pro	Ile	Leu	Val	Glu 775	Leu	Asp	Gly	Asp	Val 780	Asn	Gly	His	Lys
785					790					795				Lys	800
Thr	Leu	Lys	Phe	Ile 805	Cys	Thr	Thr	Gly	Lys 810	Leu	Pro	Val	Pro	Trp 815	Pro
			820					825					830	Arg	
Pro	Asp	His 835	Met	Lys	Gln	His	Asp 840	Phe	Phe	Lys	Ser	Ala 845	Met	Pro	Glu
_	850					855					860			Asn	
865					870					875				Asn	880
				885					890					Leu 895	
	_		900					905					910		Ala
		915					920					925		His	
	930					935					940				Thr
945					950					955					Ser 960
				965					970					His 975	
Val	Leu	Leu	Glu 980	Phe	Val	Thr	Ala	Ala 985	Gly	Ile	Thr	Leu	Gly 990	Met	Asp
Glu	Leu	Tyr 995	Lys												

- (2) INFORMATION FOR SEQ ID NO:124:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1908 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...1905
  - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:124:

														ATC Ile 15		4	18
														TCC Ser		Š	96
GAG	GGC	GAG	GGC	GAT	GCC	ACC	TAC	GGC	AAG	CTG	ACC	CTG	AAG	TTC	ATC	. 14	14

Glu	Gly	Glu 35	Gly	Asp	Ala	Thr	Туг 40	Gly	Lys	Leu	Thr	Leu 45	Lys	Phe	Ile		
					CTG Leu											192	2
-					CAG Gln 70				Arg							240	0
					AAG Lys											28	8
					AAG Lys											33	6
					GAC Asp											38	4
					GAC Asp											43	2
	Tyr				AAC Asn 150											48	10
					TTC Phe											52	8
														Asp	GGC Gly	57	76
			Leu					Tyr					Ser		CTG	62	24
		Asp			GAG Glu							Leu			TTC Phe	67	72
	Thr					Thr					Glu				TCC Ser 240	72	20
					Ala					Ser					ATG Met	7(	68
				Ile					Ala					тут	GAT Asp	8:	16

									GGC Gly							8 <b>64</b>
									ACG Thr							912
									CAG Gln							960
		-							GCC Ala 330							1008
									CTC Leu							1056
									GCC Ala							1104
									GCA Ala							1152
									GAG Glu							1200
									CGG Arg 410							1248
									CCC Pro						CCC Pro	1296
									GGT Gly				Ser		GTC Val	1344
		Ala										Pro			CCT Pro	1392
															GGC Gly 480	1440
															CAG Gln	1488
GAG	GAG	GCC	TCA	GGG	GGG	ccc	ACA	GCC	ccc	AAA	GCT	GAG	AGT	GGT	CGA	1536

Glu	Glu	Ala	Ser 500	Gly	Gly	Pro	Thr	Ala 505	Pro	Lys	Ala	Glu	Ser 510	Gly	Arg	
									ATG Met							1584
									AAA Lys							1632
									GTC Val							1680
									ACA Thr 570							1728
									ACC Thr							1776
									GTG Val							1824
									AAA Lys							1872
	-								TCT Ser							1908

### (2) INFORMATION FOR SEQ ID NO:125:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 635 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:125:

 Met
 Val
 Ser
 Lys
 Gly
 Glu
 Glu
 Leu
 Phe
 Thr
 Gly
 Val
 Pro
 Ile
 Leu

 Val
 Glu
 Glu
 Gly
 Asp
 Gly
 Asp
 Gly
 His
 Lys
 Phe
 Ser
 Val
 Ser
 Gly

 Glu
 Gly
 Gly
 Asp
 Ala
 Thr
 Tyr
 Gly
 Lys
 Leu
 Thr
 Leu
 Leu
 Lys
 Phe
 Ile

 Gly
 Gly
 Asp
 Ala
 Thr
 Tyr
 Gly
 Lys
 Leu
 Thr
 Leu
 Lys
 Leu
 Thr
 Ile
 Lys
 Thr
 Ile
 Lys
 Ile

65					70					75					80
Gln	His	Asp	Phe	Phe 85	Lys	Ser	Ala	Met	Pro 90	Glu	Gly	Тут	Val	Gln 95	Glu
Arg	Thr	Ile	Phe 100	Phe	Lys	Asp	Asp	Gly 105	Asn	Tyr	Lys	Thr	Arg 110	Ala	Glu
Val	Lys	Phe 115	Glu	Gly	Asp	Thr	Leu 120	Val	Asn	Arg	Ile	Glu 125	Leu	Lys	Gly
Ile	Asp 130	Phe	Lys	Glu	Asp	Gly 135	Asn	Ile	Leu	Gly	His 140	Lys	Leu	Glu	Tyr
Asn 145	Tyr	Asn	Ser	His	Asn 150	Val	Tyr	Ile	Met	Ala 155	Asp	Lys	Gln	Lys	Asn 160
Gly	Ile	Lys	Val	Asn 165	Phe	Lys	Ile	Arg	His 170	Asn	Ile	Glu	Asp	Gly 175	Ser
Val	Gln	Leu	Ala 180	Asp	His	Tyr	Gln	Gln 185	Asn	Thr	Pro	Ile	Gly 190	Asp	Gly
Pro	Val	Leu 195	Leu	Pro	Asp	Asn	His 200	Tyr	Leu	Ser	Thr	Gln 205	Ser	Ala	Leu
Ser	Lys 210	Asp	Pro	Asn	Glu	Lys 215	Arg	Asp	His	Met	Val 220	Leu	Leu	Glu	Phe
Val 225	Thr	Ala	Ala	Gly	Ile 230	Thr	Leu	Gly	Met	Asp 235	Glu	Leu	Tyr	Lys	Ser 240
Gly	Leu	Arg	Ser	Arg 245	Ala	Gln	Ala	Ser	Met 250	Ser	Glu	Thr	Val	Ile 255	Met
Ser	Glu	Thr	Val 260	Ile	Cys	Ser	Ser	Arg 265	Ala	Thr	Val	Met	Leu 270	Tyr	Asp
_	Gly	275	_	_			280		_		_	285			
	Arg 290					295					300				
Val 305	Gly	Arg	Lys	Met	Gln 310	Pro	Asp	Gln	Gln	Val 315	Val	Ile	Asn	Cys	Ala 320
	Val		_	325		_			330					335	
_	Arg	_	340				_	345					350		
_	Ala	355					360					365			
	Gly 370	_	_			375					380				
385		_			390					395		_	_		Gln 400
	Gly			405					410					415	
	Pro		420					425					430		
		435	_				440		_			445		_	Val
	450					455					460				Pro
465					470					475					Gly 480
	Ala			485					490					495	
			500	_	_			505		_			510	_	Arg
	_	515		_			520					525			Arg
wrd	ντά	пÃр	VIQ	1111	GIII	vaı	GTĀ	GIU	ъу	1111	FIO	nys	wsb	GIU	Ser

535 540	
530 535 540 Ser State St	
Ala Asn Gin Giu Giu 555 555 550 550 550 Arg Met Lys	
550  545  Val Arg Arg Pro Trp Glu Lys Asn Ser Thr Thr Leu Pro Arg Met Lys 575  576	
565  Ser Ser Ser Ser Val Thr Thr Ser Glu Thr Gln Pro Cys Thr Pro Ser  585  590	
Ser Ser Ser Ser Val Till III 502 590 585 590 580 580 580 580 580 580 580 580 580 58	
580 585  Ser Ser Asp Tyr Ser Asp Leu Gln Arg Val Lys Gln Glu Leu Leu Glu  Ser Ser Asp Tyr Ser Asp Leu Gln Arg Val Lys Gln Glu Leu Leu Glu	
595 600 11e Ile Glu Ala Glu Val Lys Glu Leu Gln Lys Val Lys Glu Glu Ile Ile Glu Ala 615 620	
610 Phe Val Gln Glu Leu Arg Lys Arg Gly Ser Pro 635	
630 635	
(2) INFORMATION FOR SEQ ID NO:126:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 1329 base pairs	
(B) TYPE: nucleic acid	*
(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: CDNA	
(ix) FEATURE:	
(A) NAME/KEY: Coding Sequence	
(B) LOCATION: 11326 (D) OTHER INFORMATION:	
20.126:	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:126:	48
THE THE COL CITY CITY COL ATC CITY	48
THE THE COL CITY CITY COL ATC CITY	48
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  15  10  15	
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  15  10  15  17  18  18  19  10  10  10  10  10  10  10  10  10	48 96
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Clu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly	
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  1 5 10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30	96
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  1 5 10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30	
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  GLU Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile	96
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  35 40 45	96 144
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  40 45	96
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  40 45	96 144
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  1 5 10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  35 40  TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC CTC  Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr  50 55 60	96 144 192
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  35  TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC  Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr  50	96 144
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  35  TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC  Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr  50	96 144 192
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  40 45  TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC  Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr  50 55 60  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC TAC TAC TAC TAC TAC TAC TAC TAC	96 144 192 240
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  35  TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC  Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr  50  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC TAC CCC GAC CAC ATG AGC TAC CCC TAC	96 144 192
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  35  TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC  Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr  50  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC TAC CCC GAC CAC ATG AAG  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC TAC CCC GAC CAC ATG AGC TAC CCC TAC	96 144 192 240
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  35 40 45  TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC  Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr  50 55 60  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys  65 70 70  CAG CAC GAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  CAG CAC GAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAC  CAG CAC GAC GAC TTC TCC GCC ATG CCC GAC GAC TAC GTC CAC CAC ATG CCC GCC ATG CCC ACC CAC ATG CCC GCC ATG CCC CCC ACC CCC GAC GCC TAC CCC GCC ACC CCC ACC CCC	96 144 192 240
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  1	96 144 192 240
ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG  Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu  10 15  GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC  Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly  20 25 30  GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC  Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  35 40 45  TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC  Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr  50 55 60  CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG  Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys  65 70 75 80  CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG  Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu	96 144 192 240

GTG AAG TTC GAG GGC GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 115 120 125	384
ATC GAC TTC AAG GAG GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC  11e Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr  130 135 140	432
AAC TAC AAC AGC CAC AAC GTC TAT ATC ATG GCC GAC AAG CAG AAG AAC ASn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 160	480
GGC ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 175	528
GTG CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 180 185 190	576
CCC GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205	624
AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 220	672
GTG ACC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TCC Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 230 235 240	720
GGA CTC AGA TCT CGA GCT CAA GCT TCA ATG GCT GCC ATC CGG AAG AAA Gly Leu Arg Ser Arg Ala Gln Ala Ser Met Ala Ala Ile Arg Lys Lys 245 250 255	768
CTG GTG ATT GTT GGT GAT GGA GCC TGT GGA AAG ACA TGC TTG CTC ATA Leu Val Ile Val Gly Asp Gly Ala Cys Gly Lys Thr Cys Leu Leu Ile 260 265 270	816
GTC TTC AGC AAG GAC CAG TTC CCA GAG GTG TAT GTG CCC ACA GTG TTT Val Phe Ser Lys Asp Gln Phe Pro Glu Val Tyr Val Pro Thr Val Phe 275 280 285	864
GAG AAC TAT GTG GCA GAT ATC GAG GTG GAT GGA AAG CAG GTA GAG TTG Glu Asn Tyr Val Ala Asp Ile Glu Val Asp Gly Lys Gln Val Glu Leu 290 295 300	912
GCT TTG TGG GAC ACA GCT GGG CAG GAA GAT TAT GAT CGC CTG AGG CCC Ala Leu Trp Asp Thr Ala Gly Gln Glu Asp Tyr Asp Arg Leu Arg Pro 305 310 315 320	960
CTC TCC TAC CCA GAT ACC GAT GTT ATA CTG ATG TGT TTT TCC ATG	008
AGC CCT GAT AGT TTA GAA AAC ATC CCA GAA AAG TGG ACC CCA GAA GTC 10	056

Ser	Pro	Asp	Ser 340	Leu	Glu	Asn	Ile	Pro 345	Glu	Lys	Trp	Thr	Pro 350	Glu	Val	
	-				AAC											1104
Lys	HIS	355	cys	PIO	Asn	Val	360	116	116	Leu	Val	365	ASII	гуъ	Lys	
					GAG											1152
Asp	170	Arg	Asn	Asp	Glu	His 375	Thr	Arg	Arg	GIu	180	Ala	Lys	Met	Lys	
					CCT											1200
Gln 385	Glu	Pro	Val	Lys	Pro 390	Glu	Glu	Gly	Arg	Asp 395	Met	Ala	Asn	Arg	Ile 400	
365					390					333					400	
					ATG											1248
Gly	Ala	Phe	Gly	Тут 405	Met	Glu	Cys	Ser	Ala 410	Lys	Thr	Lys	Asp	Gly 415	Val	
AGA	GAG	GTT	TTT	GAA	ATG	GCT	ACG	AGA	GCT	GCT	CTG	CAA	GCT	AGA	CGT	1296
Arg	Glu	Val		Glu	Met	Ala	Thr		Ala	Ala	Leu	Gln		Arg	Arg	
			420					425					430			
GGG	AAG	AAA	AAA	TCT	GGT	TGC	CTT	GTC	TTG	TGA						1329
Gly	Lys	_	Lys	Ser	Gly	Cys	Leu 440	Val	Leu							
		435					440									

### (2) INFORMATION FOR SEQ ID NO:127:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 442 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:127:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 45 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 110 100 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 125 120 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr

135 130 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 145 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 190 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 225 230 235 Gly Leu Arg Ser Arg Ala Gln Ala Ser Met Ala Ala Ile Arg Lys Lys 245 250 Leu Val Ile Val Gly Asp Gly Ala Cys Gly Lys Thr Cys Leu Leu Ile 260 265 Val Phe Ser Lys Asp Gln Phe Pro Glu Val Tyr Val Pro Thr Val Phe 280 285 275 Glu Asn Tyr Val Ala Asp Ile Glu Val Asp Gly Lys Gln Val Glu Leu 290 295 300 Ala Leu Trp Asp Thr Ala Gly Gln Glu Asp Tyr Asp Arg Leu Arg Pro 305 310 315 Leu Ser Tyr Pro Asp Thr Asp Val Ile Leu Met Cys Phe Ser Ile Asp 330 335 325 Ser Pro Asp Ser Leu Glu Asn Ile Pro Glu Lys Trp Thr Pro Glu Val 340 345 Lys His Phe Cys Pro Asn Val Pro Ile Ile Leu Val Gly Asn Lys Lys 355 360 Asp Leu Arg Asn Asp Glu His Thr Arg Arg Glu Leu Ala Lys Met Lys 380 375 Gln Glu Pro Val Lys Pro Glu Glu Gly Arg Asp Met Ala Asn Arg Ile 385 390 395 Gly Ala Phe Gly Tyr Met Glu Cys Ser Ala Lys Thr Lys Asp Gly Val 405 410 Arg Glu Val Phe Glu Met Ala Thr Arg Ala Ala Leu Gln Ala Arg Arg 420 425 430 Gly Lys Lys Ser Gly Cys Leu Val Leu 440 435

### (2) INFORMATION FOR SEQ ID NO:128:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1140 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...1137
  - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:128:

Met 1	Asp	His	Tyr	Asp 5	Ser	Gln	Gln	Thr	Asn 10	Asp	Tyr	Met	Gln	Pro 15	Glu	
				CGG Arg												96
				TTC Phe												144
				GAG Glu												192
				CTG Leu												240
				ATG Met 85												288
				GCC Ala												336
				GAT Asp											TGG Trp	384
															AAG Lys	432
															GAC Asp 160	480
															GGC	528
														Thr	GGC	576
			Val					Leu					Thr		GGC	624
															TTC Phe	672
	Lys										Glu				TTC Phe 240	720

 	 -	 AAC Asn						768
 		AAC Asn						816
 		CTG Leu						864
 	 	 ATG Met	 	 	 	 	 	912
 		CAC His 310						960
 	 	 AAC Asn	 	 		 -	 	1008
 		CTG Leu						1056
		CAC His						1104
 	 	 ATG Met			TAA			1140

## (2) INFORMATION FOR SEQ ID NO:129:

#### (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 379 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

### (ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE: internal

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:129:

 Leu Met Leu Leu Glu Val Ile Ser Gly Glu Arg Leu Ala Lys Pro 65 70 75 Glu Arg Gly Lys Met Arg Val His Lys Ile Ser Asn Val Asn Lys Ala 85 90 Leu Asp Phe Ile Ala Ser Lys Gly Val Lys Leu Val Ser Ile Gly Ala 100 105 Glu Glu Ile Val Asp Gly Asn Val Lys Met Thr Leu Gly Met Ile Trp 120 125 115 Thr Ile Ile Leu Arg Arg Asp Pro Pro Val Ala Thr Met Val Ser Lys 135 140 Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp 145 150 155 Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly 165 170 175 Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly 180 185 Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly 195 200 205 Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp Phe 210 215 220 Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe 225 230 235 Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu 245 250 Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys 260 265 270 Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser 280 His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val 295 300 Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala 305 310 315 Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu 325 330 Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro 340 345 350 Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala 355 360 365 Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys

### (2) INFORMATION FOR SEQ ID NO:130:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 3516 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...3513
  - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:130:

					GAG Glu											48
	_				GAC Asp											96
					GCC Ala											144
					CTG Leu											192
					CAG Gln 70											240
					AAG Lys											288
					AAG Lys											336
					GAC Asp											384
					GAC Asp											432
					AAC Asn 150											480
					TTC Phe											528
												_	_		GGC	576
															CTG Leu	624
															TTC	672
GTG	ACC	GCC	GCC	GGG	ATC	ACT	CTC	GGC	ATG	GAC	GAG	CTG	TAC	AAG	TCC	720

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Val 225	Thr	Ala	Ala	Gly	Ile 230	Thr	Leu	Gly	Met	Asp 235	Glu	Leu	Tyr	Lys	Ser 240	
										GAG Glu						768
										GGG Gly						816
										GAG Glu						864
										AGC Ser						912
										CTC Leu 315						960
										CGA Arg						1008
										GCC Ala						1056
										CCG Pro						1104
										GCG Ala						1152
										CCG Pro 395						1200
										CTT Leu					Ala	1248
										GAC Asp					TAC Tyr	1296
										GGG Gly					TGT Cys	1344
		Phe													CCA Pro	1392

									GAG Glu							1440
									CGC Arg 490					_		1488
									TTG Leu							1536
									TCG Ser							1584
									GCT Ala							1632
									AAC Asn							1680
									AAG Lys 570							1728
									GGC							1776
									GAG Glu							1824
									CCG Pro						CCG Pro	1872
									ATC Ile							1920
											_				CTG Leu	1968
														_	ACA Thr	2016
												_			GTG Val	2064
GTT	CAG	CTC	CAT	GGC	TAC	ATG	GAA	AAC	AAG	CCT	CTG	GGA	CTT	CAG	ATC	2112

Val	Gln 690	Leu	His	Gly	Tyr	Met 695	Glu	Asn	Lys	Pro	Leu 700	Gly	Leu	Gln	Ile	
									CTT Leu			_				2160
									GTC Val 730							2208
									GAG Glu							2256
									GCG Ala							2304
									GAG Glu							2352
									GTT Val							2400
									TCT Ser 810							2448
									GTT Val							2496
									ATG Met							2544
										_		_	_		GGA Gly	2592
									GTG Val						CAG Gln 880	2640
									GAA Glu 890							2688
									GTC Val			_			AAA Lys	2736
									CAC His						AAG Lys	2784

100	GAG	~~~	200	CAM	C 2 2	መእመ	CAC	ccc	እርጥ	CITC	יעיית	ייניעי	אכיר	CCC	ACC.	2832
	Glu															2032
	930			•		935	•				940	-				-
O1m	001	~~~	OMC.	~~~	200	CAC	OC III	mac.	mac.	ccc	CAC	CAC	ccc	באניע	CITC	2880
	GGA Gly															2000
945	01,	01,			950			-1-	-4-	955					960	
																2022
	GAG Glu															2928
Ald	GIU	ser	PIU	965	Cys	Leu	vai	AIG	970	nec	AIG	110	Cys	975	Gin	
	CGC															2976
Phe	Arg	Thr	980	Leu	Ser	Ser	Pro	985	Ala	Arg	TYL	GIN	990	GIII	ASII	
			200					505								
	GCG															3024
Pro	Ala		Val	Leu	Tyr			Ser	Lys	Ser			Pro	Ser	Leu	
		995				•	1000				-	1005				
	GGC															3072
Leu	Gly	Tyr	Gln	Gln			Leu	Met	Ala			Leu	Ser	Leu	Ala	
	1010					1015					1020					
GAC	GCT	CAC	CGC	TCT	GTG	CTG	GTG	CAC	GCC	GGC	TCC	CAG	GGC	CAG	AGC	3120
	Ala															
1025					1030				;	1035					1040	
σγ. λ	ccc	CITC	CTC	CAC	ccc	ጥርጥ	CCG	ACC	אאר	CAG	CAG	GCC	TCG	ССТ	GTG	3168
	Ala															5100
				1045					1050					1055		
		m> 0	ma.	000	100	220	CNC	CAC	CITY	ccc	mcc.	CCA	NCC	CAC	CAG	3216
															CAG Gln	3210
110		_	1060					1065					1070			
																2064
															ACC Thr	3264
GIU		GIN 1075		me	Mec		ديء 1080		ASII	File		1085		1111	1111	
															GGT	3312
Arg			Pro	Pro		۷aı 1095	Ser	GIn	GIY		Arg 1100	Leu	ser	Pro	Gly	
	1090					1093					1100					
															CCC	3360
	_	Pro	Thr				Gln	Gln				Ser	Gln	Arg	Ala	
1105	•				1110					1115					1120	
GCC	: AAA	AAC	GGA	ccc	CCG	GTC	AGT	GAC	CAA	AAG	GAA	GTA	TTA	CCI	GCG	3408
Ala	Lys	Asn	Gly	Pro	Pro	Val	Ser	Asp	Gln	Lys	Glu	Val	Leu		Ala	
				1125					1130					1135	5	
CCC	: GTYG	ACC	יייים	444	CAG	GAG	CAG	AAC	TTG	GAC	CAG	ACC	TAC	TTC	GAT	3456
															a Asp	
_			1140					1145					1150			
03.0		ייני אל ו		y mv.	, yu.	ארר	יאמ י	י פאר	thin.	תייעה י	מבים .	רריי	י רכיז	י כבר	CAGA	3504
GAT	G11	AA'I'	GAA	ATT	ATC	AGG	MAC	GAC	111	ICA	. GGA		1		, AGA	2204

Asp Val Asn Glu Ile Ile Arg Lys Glu Phe Ser Gly Pro Pro Ala Arg 1155 1160 1165

AAT CAG ACG TAA Asn Gln Thr 1170 3516

### (2) INFORMATION FOR SEQ ID NO:131:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1171 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:131:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 5 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 90 85 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 110 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 125 115 120 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 140 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 155 150 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 170 165 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 185 190 Pro Val Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 205 200 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 220 210 215 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 235 230 Gly Leu Arg Ser Arg Ala Met Asn Ala Pro Glu Arg Gln Pro Gln Pro 245 250 Asp Gly Gly Asp Ala Pro Gly His Glu Pro Gly Gly Ser Pro Gln Asp 260 265 Glu Leu Asp Phe Ser Ile Leu Phe Asp Tyr Glu Tyr Leu Asn Pro Asn 280 Glu Glu Glu Pro Asn Ala His Lys Val Ala Ser Pro Pro Ser Gly Pro

	290					295					300				
Ala 305	Tyr	Pro	Asp	Asp	Val 310	Met	Asp	Tyr	Gly	Leu 315	Lys	Pro	Tyr	Ser	Pro 320
Leu	Ala	Ser	Leu	Ser 325	Gly	Glu	Pro	Pro	Gly 330	Arg	Phe	Gly	Glu	Pro 335	Asp
Arg	Val	Gly	Pro 340	Gln	Lys	Phe	Leu	Ser 345	Ala	Ala	Lys	Pro	Ala 350	Gly	Ala
Ser	Gly	Leu 355	Ser	Pro	Arg	Ile	Glu 360	Ile	Thr	Pro	Ser	His 365	Glu	Leu	Ile
Gln	Ala 370	Val	Gly	Pro	Leu	Arg 375	Met	Arg	Asp	Ala	Gly 380	Leu	Leu	Val	Glu
385	Pro				390					395	_				400
Val	Pro	Gly	Phe	Glu 405	Gly	Tyr	Arg	Glu	Pro 410	Leu	Cys	Leu	Ser	Pro 415	Ala
	Ser	_	420					425		_			430		
	Ser	435					440					445			
Pro	Gln 450	Phe	Gln	Asn	Ile	Pro 455	Ala	His	Tyr	Ser	Pro 460	Arg	Thr	Ser	Pro
465	Met				470					475					480
	Ser			485					490					495	
_	Arg		500					505					510		
	Ser	5 <b>15</b>		_		_	520					525			
	Ala 530					535					540				
545	Ser				550	_				555					560
	Cys	_		565		_			570					575	
			580					585					590		Tyr
		595					600					605			Asn
	610					615					620	_			Pro
625					630					635					Leu 640
				645					650					655	Leu Thr
J			660			-		665		-			670		Val
	-	675	_	_			680				_	685		_	Ile
	690					695					700				
705		_			710		_			715					Tyr 720
				725					730					735	
_			740					745					750		Lys
Asn	Asn	met	Arg	ATA	ınr	тте	ASP	cys	ΑΙά	GTĀ	тте	Leu	ьys	Leu	Arg

760 Asn Ala Asp Ile Glu Leu Arg Lys Gly Glu Thr Asp Ile Gly Arg Lys 770 775 780 Asn Thr Arg Val Arg Leu Val Phe Arg Val His Ile Pro Glu Ser Ser 790 795 Gly Arg Ile Val Ser Leu Gln Thr Ala Ser Asn Pro Ile Glu Cys Ser 805 810 815 Gln Arg Ser Ala His Glu Leu Pro Met Val Glu Arg Gln Asp Thr Asp 820 825 830 Ser Cys Leu Val Tyr Gly Gly Gln Gln Met Ile Leu Thr Gly Gln Asn 840 Phe Thr Ser Glu Ser Lys Val Val Phe Thr Glu Lys Thr Thr Asp Gly 850 855 860 Gln Gln Ile Trp Glu Met Glu Ala Thr Val Asp Lys Asp Lys Ser Gln 865 870 875 880 Pro Asn Met Leu Phe Val Glu Ile Pro Glu Tyr Arg Asn Lys His Ile 885 890 895 Arg Thr Pro Val Lys Val Asn Phe Tyr Val Ile Asn Gly Lys Arg Lys 905 910 Arg Ser Gln Pro Gln His Phe Thr Tyr His Pro Val Pro Ala Ile Lys 925 915 920 Thr Glu Pro Thr Asp Glu Tyr Asp Pro Thr Leu Ile Cys Ser Pro Thr 930 935 940 His Gly Gly Leu Gly Ser Gln Pro Tyr Tyr Pro Gln His Pro Met Val 945 950 955 Ala Glu Ser Pro Ser Cys Leu Val Ala Thr Met Ala Pro Cys Gln Gln 970 975 965 Phe Arg Thr Gly Leu Ser Ser Pro Asp Ala Arg Tyr Gln Gln Asn 980 985 990 Pro Ala Ala Val Leu Tyr Gln Arg Ser Lys Ser Leu Ser Pro Ser Leu 995 1000 1005 Leu Gly Tyr Gln Gln Pro Ala Leu Met Ala Ala Pro Leu Ser Leu Ala 1010 1015 1020 Asp Ala His Arg Ser Val Leu Val His Ala Gly Ser Gln Gly Gln Ser 025 1030 1035 Ser Ala Leu Leu His Pro Ser Pro Thr Asn Gln Gln Ala Ser Pro Val 1045 1050 1055 Ile His Tyr Ser Pro Thr Asn Gln Gln Leu Arg Cys Gly Ser His Gln 1060 1065 1070 Glu Phe Gln His Ile Met Tyr Cys Glu Asn Phe Ala Pro Gly Thr Thr 1075 1080 1085 Arg Pro Gly Pro Pro Pro Val Ser Gln Gly Gln Arg Leu Ser Pro Gly 1090 1095 1100 Ser Tyr Pro Thr Val Ile Gln Gln Gln Asn Ala Thr Ser Gln Arg Ala 105 1110 1115 1120 Ala Lys Asn Gly Pro Pro Val Ser Asp Gln Lys Glu Val Leu Pro Ala 1125 1130 1135 Gly Val Thr Ile Lys Gln Glu Gln Asn Leu Asp Gln Thr Tyr Leu Asp 1140 1145 1150 Asp Val Asn Glu Ile Ile Arg Lys Glu Phe Ser Gly Pro Pro Ala Arg 1160 1165 1155 Asn Gln Thr 1170

- (2) INFORMATION FOR SEQ ID NO:132:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 3546 base pairs

(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...3543 (D) OTHER INFORMATION:

(D) OTHER INFORMATION.

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:132:

											GGC Gly					48
											CTT Leu					96
											GAA Glu					144
		GTC									TAC Tyr 60					192
	GAC					CCA					GCT Ala					240
GAG					TTC					AGG	GTA Val					288
				GCC					GCC		GGC Gly			ССТ		336
		Ile	ACT					CTG			GCA Ala		GGG			384
	Met					Leu	CTG					ccc			GGG Gly	432
Val					Arg					Val	CCC Pro				Gly	480
									Ala					Ser	GCC Ala	528
AGC	TTC	АТТ	TCT	165 GAC	ACC	TTC	TCC	ccc	170 TAC	ACC	TCG	ccc	TGC	175 GTC	TCG	576

Ser	Phe	Ile	Ser 180	Asp	Thr	Phe	Ser	Pro 185	Tyr	Thr	Ser	Pro	Cys 190	Val	Ser	
									TGT Cys							624
									CCA Pro							672
									CGC Arg							720
									GCC Ala 250							768
									GGA Gly							816
									CAC His							864
									GCT Ala							912
									TCG Ser							960
									TCG Ser 330						CCA Pro	1008
															CTG Leu	1056
									AAC Asn						ATC	1104
									CCG Pro						CCC Pro	1152
	Суѕ														CCG Pro 400	1200
															CCC	1248

					GCC Ala											1296
					GGA Gly											1344
					CTG Leu											1392
					CCG Pro 470											1440
					ACC Thr											1488
-					CCC Pro											1536
					ATC Ile											1584
					GAC Asp											1632
					ATC Ile 550										TTA Leu 560	1680
					CCC Pro										Glu	1728
					AGA Arg									Tyr	GGC	1776
			Met												Lys	1824
		Phe										Ile			ATG Met	1872
	Ala										Asn				GTT Val 640	. 1920
GAG	ATC	CCT	GAA	TAT	CGG	AAC	AAG	CAT	ATC	CGC	ACA	CCT	GTA	AAA	GTG	1968

Glu	Ile	Pro	Glu	Tyr 645	Arg	Asn	Lys	His	Ile 650	Arg	Thr	Pro	Val	Lys 655	Val	
								AGA Arg 665							_	2016
								ATC Ile								2064
								CCC Pro								2112
								ATG Met								2160
								CAG Gln								2208
								CAG Gln 745								2256
								AGC Ser							CCG Pro	2304
								CTT Leu			_	_			GTG Val	2352
															CCC Pro 800	2400
															ACC Thr	2448
														Ile	ATG Met	2496
												_			CCG Pro	2544
												Pro			ATT	2592
						Ser									CCG Pro 880	2640

									GCG Ala 890							2688
									GAT Asp							2736
									AGA Arg							2784
									CCA Pro							2832
									GTG Val							2880
									TTC Phe 970							2928
									ACC Thr			_	_		_	2976
						Pro			ACC Thr		Val					3024
Tyr					Phe				CCC Pro	Asp						3072
				Ser					GGC		Val			Arg		3120
			Lys					Tyr	AAG Lys 1050				Glu			3168
		Gly					Asn		Ile			Lys		Ile	GAC Asp	3216
	Lys		Asp			Ile		Gly			Leu				TAC	3264
Asn					Tyr					Lys		Lys			ATC Ile	3312
AAG	GTG	AAC	TTC	AAG	ATC	CGC	CAC	AAC	ATC	GAG	GAC	GGC	AGC	GTG	CAG	3360

Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln 1105 1110 3408 CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC CCC GTG Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val 1125 1130 CTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC AAA 3456 Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys 1145 GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC GTG ACC 3504 Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr 1155 1160 1165 GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA 3546 Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 1170 1175

#### (2) INFORMATION FOR SEQ ID NO:133:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1181 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:133:

Met Asn Ala Pro Glu Arg Gln Pro Gln Pro Asp Gly Gly Asp Ala Pro 10 5 Gly His Glu Pro Gly Gly Ser Pro Gln Asp Glu Leu Asp Phe Ser Ile 25 30 Leu Phe Asp Tyr Glu Tyr Leu Asn Pro Asn Glu Glu Glu Pro Asn Ala 40 45 His Lys Val Ala Ser Pro Pro Ser Gly Pro Ala Tyr Pro Asp Asp Val 60 55 Met Asp Tyr Gly Leu Lys Pro Tyr Ser Pro Leu Ala Ser Leu Ser Gly 70 75 Glu Pro Pro Gly Arg Phe Gly Glu Pro Asp Arg Val Gly Pro Gln Lys 90 Phe Leu Ser Ala Ala Lys Pro Ala Gly Ala Ser Gly Leu Ser Pro Arg 110 100 105 Ile Glu Ile Thr Pro Ser His Glu Leu Ile Gln Ala Val Gly Pro Leu 120. Arg Met Arg Asp Ala Gly Leu Leu Val Glu Gln Pro Pro Leu Ala Gly 135 140 Val Ala Ala Ser Pro Arg Phe Thr Leu Pro Val Pro Gly Phe Glu Gly 150 155 Tyr Arg Glu Pro Leu Cys Leu Ser Pro Ala Ser Ser Gly Ser Ser Ala 165 170 175 Ser Phe Ile Ser Asp Thr Phe Ser Pro Tyr Thr Ser Pro Cys Val Ser 185 Pro Asn Asn Gly Gly Pro Asp Asp Leu Cys Pro Gln Phe Gln Asn Ile

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	450					4	55						460						
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				483	)					490	)					49	5		
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		212	)					520						525					
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	230					53	35						540						
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				565						570	1					575			
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			280						585						590				
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		595					6	00						605					
Val	Val	Phe	Thr	Glu	Lys	Th	r T	hr .	Asp	Gly	G	ln (	Gln	Ile	Trp	Glu	M	let	
	PIO					61	5						520						
Glu	Ala	Thr	Val	Asp	Lys	As	рL	ys :	Ser	Gln	P	ro Z	Asn 1	Met	Leu	Phe	ν	al	
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Glu	Ile	Pro	Glu	Tyr	Arg	As	n L	ys I	His	Ile	A	rg 7	Chr :	Pro	Val	Lvs	v	al	
				645						650						655			
Asn	Phe	Tyr	Val	Ile	Asn	Gl	y L	ys 2	Arg	Lys	Aı	rg S	Ger (	Gln	Pro	Gln	н	is	
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660 665 Phe Thr Tyr His Pro Val Pro Ala Ile Lys Thr Glu Pro Thr Asp Glu 680 Tyr Asp Pro Thr Leu Ile Cys Ser Pro Thr His Gly Gly Leu Gly Ser 700 695 Gln Pro Tyr Tyr Pro Gln His Pro Met Val Ala Glu Ser Pro Ser Cys 705 710 715 Leu Val Ala Thr Met Ala Pro Cys Gln Gln Phe Arg Thr Gly Leu Ser 730 725 Ser Pro Asp Ala Arg Tyr Gln Gln Gln Asn Pro Ala Ala Val Leu Tyr 740 745 Gln Arg Ser Lys Ser Leu Ser Pro Ser Leu Leu Gly Tyr Gln Gln Pro 760 765 Ala Leu Met Ala Ala Pro Leu Ser Leu Ala Asp Ala His Arg Ser Val 780 770 775 Leu Val His Ala Gly Ser Gln Gly Gln Ser Ser Ala Leu Leu His Pro 785 790 795 Ser Pro Thr Asn Gln Gln Ala Ser Pro Val Ile His Tyr Ser Pro Thr 805 810 Asn Gln Gln Leu Arg Cys Gly Ser His Gln Glu Phe Gln His Ile Met 825 830 Tyr Cys Glu Asn Phe Ala Pro Gly Thr Thr Arg Pro Gly Pro Pro 835 840 845 Val Ser Gln Gly Gln Arg Leu Ser Pro Gly Ser Tyr Pro Thr Val Ile 855 Gln Gln Gln Asn Ala Thr Ser Gln Arg Ala Ala Lys Asn Gly Pro Pro 875 . 880 Val Ser Asp Gln Lys Glu Val Leu Pro Ala Gly Val Thr Ile Lys Gln 885 890 Glu Gln Asn Leu Asp Gln Thr Tyr Leu Asp Asp Val Asn Glu Ile Ile 900 905 Arg Lys Glu Phe Ser Gly Pro Pro Ala Arg Asn Gln Thr Arg Ile Leu 920 925 Gln Ser Thr Val Pro Arg Ala Arg Asp Pro Pro Val Ala Thr Met Val 940 935 Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu 955 950 Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly 970 965 Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr 985 990 980 Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr 995 1000 1005 Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His 1010 1015 1020 Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr 025 1030 1035 1040 Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys 1045 1050 Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp 1060 1065 1070 Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr 1080 1085 Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile 1090 1095 1100 Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln 105 1110 1115 Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val

1125 1130 Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys 1140 1145 Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr 1155 1160 Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 1175 (2) INFORMATION FOR SEQ ID NO:134: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2802 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: Coding Sequence (B) LOCATION: 1...2799 (D) OTHER INFORMATION: (xi) SEQUENCE DESCRIPTION: SEQ ID NO:134: 48 ATG GTG AGC AAG GGC GAG GAG CTG TTC ACC GGG GTG GTG CCC ATC CTG Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC 96 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 25 GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC 144 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC 192 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG 240 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG 288 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu CGC ACC ATC TTC TTC AAG GAC GAC GGC AAC TAC AAG ACC CGC GCC GAG 336 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 GTG AAG TTC GAG GGC GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 125

ATC GAC TTC AAG GAG GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC

432

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ARC TAC ARC RCC CAC ARC GRC TAT ATC ATC GCC GRC ARG CAG ARG ARC		Ile	Asp 130	Phe	Lys	Glu	Asp	Gly 135	Asn	Ile	Leu	Gly	His 140	Lys	Leu	Glu	Tyr		
Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 165 170 170 175 175 175 175 175 165 165 177 166 1165 177 176 177 176 176 177 176 176 177 177		Asn	TAC Tyr	AAC Asn	AGC Ser	CAC His	Asn	GTC Val	TAT Tyr	ATC Ile	ATG Met	Ala	GAC Asp	AAG Lys	CAG Gln	AAG Lys	Asn	480	
Val Gin Leu Ala Asp His Tyr Gin Gin Asn Thr Pro Ile Gly Asp Gly 180  CCC GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gin Ser Ala Leu 205  AGC AAA GAC CCC AAC GAG AAG CCC GAT CAC ATG GTC CTG CTG GAG TTC Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 215  GTG ACC GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TCC Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 225  GGA CTC AGA TCT CGA GGG ACC ATG GGC ACC TTG GGG GAT TTA CAG TAC GIly Leu Arg Ser Arg Gly Ser Met Gly Thr Leu Arg Asp Leu Gin Tyr 255  GCG CTC CAG GAG AAG ATC GAG GAG GAG CAG CGG GAT CTC ATC ATC GIly Leu Arg Ser Arg Gly Ser Met Gly Thr Leu Arg Asp Leu Gin Tyr 255  GCG CTC CAG GAG AAG ATC GAG GAG CTG GAG CAG CGG GAT CTC ATC ATC Ala Leu Gin Glu Lys Ile Glu Glu Leu Arg Gin Arg Asp Ala Leu Ile 260  GAC GAG CTG GAG CTG GAG TTG GAT TAG GAG CAG CGG GAT CTC ATC ATC Ala Leu Gin Glu Leu Glu Leu Asp Gin Lys Asp Glu Leu Ile Gin Lys 275  CTG CAG AAC GAG CTG GAC AAG TAG CAG CAG CGG GAA CTG ATC CAG AAG Asp Glu Leu Glu Leu Asp Gin Lys Asp Glu Leu Ile Gin Lys 275  CTG CAG AAC GAG CTG GAC AAG TAC CGC TCG GTG ATC CAG CAC CE Leu Gin Asn Glu Leu Asp Lys Tyr Arg Ser Val Ile Arg Pro Ala Thr 290  CAG CAG CCG CAG CAG AAG CAG GCG GAG CCC CCC						Asn					His					Gly		528	
Pro Val   Leu   Leu   Pro   Asp   Asp   Asp   His   Tyr   Leu   Ser   Thr   Gln   Ser   Ala   Leu   205					Ala					Gln					Gly			576	
Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 CFG ACC CCC GCC GCC ACC ACC CCC CCC GCC ACC A				Leu					His					Gln				624	
Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 240  GGA CTC AGA TCT CGA GGG AGC ATG GGC ACC TTG CGG GAT TTA CAG TAC Gly Leu Arg Ser Arg Gly Ser Met Gly Thr Leu Arg Asp Leu Gln Tyr 255  GCG CTC CAG GAG AAG ATC GAG GAG CTG AGG CAG CAG CAG CAG CAT CTC ATC Ala Leu Gln Glu Lys Ile Glu Glu Leu Arg Gln Arg Asp Ala Leu Ile 260  GAC GAG CTG GAG CTG GAG TTG GAT CAG AAG GAC CTG ATG ATG CAG AAG AAG CTG ATC CAG AAG ASp Glu Leu Glu Leu Glu Leu Asp Gln Lys Asp Gln Lys Arg Gln Arg Asp Ala Leu Ile Gln Lys 275  CTG CAG AAC GAG CTG GAC TTG GAT CAG AAG GAC GAG CAG CAG CAG CAG ACC ACC	ı		Lys					Lys					Val					672	
Gly Leu Arg Ser Arg Gly Ser Met Gly Thr Leu Arg Asp Leu Gln Tyr 255  GCG CTC CAG GAG AAG ATC GAG GAG CTG AGG CAG CGG GAT GCT CTC ATC Ala Leu Gln Glu Lys Ile Glu Glu Leu Arg Gln Arg Asp Ala Leu Ile 260  GAC GAG CTG GAG CTG GAG TTG GAT CAG AAG GAC GAA CTG ATC CAG AAG AGC GAS GU Leu Ile Gln Lys 275  CTG CAG AAC GAG CTG GAG CTG GAC AAG TAC CGC TCG GTG ATC CAG ACC GAA CTG ATC CAG AAG GAC CTG ATC CAG AAG GAS CTG AAC GAC CTG ATC CAG AAC GAC CTG GIN Arg Asp Gln Leu Ile Gln Lys 285  CTG CAG AAC GAG CTG GAC AAG TAC CGC TCG GTG ATC CGA CCA GCC ACC Leu Gln Asn Glu Leu Asp Lys Tyr Arg Ser Val Ile Arg Pro Ala Thr 290  CAG CAG CAG CAG AAG CAG AGC CAG AGC ACC TTG CAG GAC GAG CCG CAC GLO GIN Gln Ala Gln Lys Gln Ser Ala Ser Thr Leu Gln Gly Glu Pro Arg 315  ACC AAG CAG CAG CAG ATC TCC CCC GAC CCC ACC GCC TTC GAC ATC CAG ATC CAG ATC CAG AAG AAG CTG ATC CAG AAG AAC AAG CTG ATC CAG AAG AAC AAG CTG ATC CAG AAG AAG CTG ATC CAG AAG AAG CTG ATC CAG AAG AAC AAG CTG ATC CAG AAG AAC AAG CTG AAG AAC AAG AAC AAG CTG AAG AAC AAC		Val					Ile					Asp					Ser	720	
Ala Leu Gln Glu Lys Ile Glu Glu Leu Arg Gln Arg Asp Ala Leu Ile 260  GAC GAG CTG GAG CTG GAG TTG GAT CAG AAG GAC GAA CTG ATC CAG AAG ASp Glu Leu Glu Leu Glu Leu Glu Leu Asp Gln Lys Asp Glu Leu Ile Gln Lys 280  CTG CAG AAC GAG CTG GAC AAG TAC CGC TCG GTG ATC CGA CCA GCC ACC 285  CTG CAG AAC GAG CTG GAC AAG TAC CGC TCG GTG ATC CGA CCA GCC ACC 2912  Leu Gln Asn Glu Leu Asp Lys Tyr Arg Ser Val Ile Arg Pro Ala Thr 290  CAG CAG GCG CAG AAG CAG AGC GCG AGC ACC TTG CAG GGC GAG CCG CGC GGn Gln Gln Ala Gln Lys Gln Ser Ala Ser Thr Leu Gln Gly Glu Pro Arg 315  ACC AAG CGG CAG GCG ATC TCC GCC GAG CCC ACC GGC GAG CCG CGC GAG CCG CGC ASp Ala Ser Thr Ala Phe Asp Ile Gln 325  GAT CTC AGC CAT GTG ACC CTG CCC TTC TAC CGC AAG AGC CCA GCC CAG TCC ASp Leu Ser His Val Thr Leu Pro Phe Tyr Pro Lys Ser Pro Gln Ser 350  AAG GAT CTT ATA AAG GAA GCT ATC CTT GAC AAT GAC TTT ATG AAG AAC Lys Asp Leu Ile Lys Glu Ala Ile Leu Asp Asn Asp Phe Met Lys Asn		GGA Gly	CTC Leu	AGA Arg	TCT Ser	Arg	GGG Gly	AGC Ser	ATG Met	GGC Gly	Thr	TTG Leu	CGG Arg	GAT Asp	TTA Leu	Gln	TAC Tyr	768	
Asp Glu Leu Glu Leu Glu Leu Asp Gln Lys Asp Gln Lys Asp Glu Leu Tle Gln Lys 280  CTG CAG AAC GAG CTG GAC AAG TAC CGC TCG GTG ATC CGA CCA GCC ACC Leu Gln Asn Glu Leu Asp Lys Tyr Arg Ser Val Ile Arg Pro Ala Thr 290  CAG CAG GCG CAG AAG CAG AGC GCG AGC ACC TTG CAG GGC GAG CCG CGC GCG Gln Gln Ala Gln Lys Gln Ser Ala Ser Thr Leu Gln Gly Glu Pro Arg 315  ACC AAG CGG CAG GCG ATC TCC GCC GAG CCC ACC GCC TTC GAC ATC CAG GCC TTC GAC ATC CAG GAG CCG CAG ATC CAG GAG CCG CAG GCC ACC GCC TTC GAC ATC CAG GAG CCG CAG CA					Glu					Leu					Ala			816	
Leu Gln Asn Glu Leu Asp Lys Tyr Arg Ser Val Ile Arg Pro Ala Thr 290  CAG CAG CAG CAG AAG CAG AGC CCG AGC ACC TTG CAG GGC GAG CCG CGC Gln Gln Ala Gln Lys Gln Ser Ala Ser Thr Leu Gln Gly Glu Pro Arg 305  ACC AAG CGG CAG GCG ATC TCC GCC GAG CCC ACC GCC TTC GAC ATC CAG Thr Lys Arg Gln Ala Ile Ser Ala Glu Pro Thr Ala Phe Asp Ile Gln 325  GAT CTC AGC CAT GTG ACC CTG CCC TTC TAC CCC AAG AGC CCA CAG TCC Asp Leu Ser His Val Thr Leu Pro Phe Tyr Pro Lys Ser Pro Gln Ser 340  AAG GAT CTT ATA AAG GAA GCT ATC CTT GAC AAT GAC TTT ATG AAG AAC Lys Asp Leu Ile Lys Glu Ala Ile Leu Asp Asn Asp Phe Met Lys Asn				Leu					Asp					Leu	Ile			864	
Gln Gln Ala Gln Lys Gln Ser Ala Ser Thr Leu Gln Gly Glu Pro Arg 305 310 310 315 315 320  ACC AAG CGG CAG GCG ATC TCC GCC GAG CCC ACC GCC TTC GAC ATC CAG Thr Lys Arg Gln Ala Ile Ser Ala Glu Pro Thr Ala Phe Asp Ile Gln 325 320  GAT CTC AGC CAT GTG ACC CTG CCC TTC TAC CCC AAG AGC CCA CAG TCC Asp Leu Ser His Val Thr Leu Pro Phe Tyr Pro Lys Ser Pro Gln Ser 340 345 350  AAG GAT CTT ATA AAG GAA GCT ATC CTT GAC AAT GAC TTT ATG AAG AAC Lys Asp Leu Ile Lys Glu Ala Ile Leu Asp Asn Asp Phe Met Lys Asn			Gln					Lys					Ile	Arg				912	
Thr Lys Arg Gln Ala Ile Ser Ala Glu Pro Thr Ala Phe Asp Ile Gln 325  GAT CTC AGC CAT GTG ACC CTG CCC TTC TAC CCC AAG AGC CCA CAG TCC 1056  Asp Leu Ser His Val Thr Leu Pro Phe Tyr Pro Lys Ser Pro Gln Ser 340  AAG GAT CTT ATA AAG GAA GCT ATC CTT GAC AAT GAC TTT ATG AAG AAC 1104  Lys Asp Leu Ile Lys Glu Ala Ile Leu Asp Asn Asp Phe Met Lys Asn		Gln	Gln				Gln					Leu	Gln				Arg	960	
Asp Leu Ser His Val Thr Leu Pro Phe Tyr Pro Lys Ser Pro Gln Ser 340 345 350  AAG GAT CTT ATA AAG GAA GCT ATC CTT GAC AAT GAC TTT ATG AAG AAC Lys Asp Leu Ile Lys Glu Ala Ile Leu Asp Asn Asp Phe Met Lys Asn		ACC Thr	AAG Lys	CGG Arg	CAG Gln	Ala	ATC Ile	TCC Ser	GCC Ala	GAG Glu	Pro	Thr	GCC Ala	TTC Phe	GAC Asp	Ile	Gln	1008	
Lys Asp Leu Ile Lys Glu Ala Ile Leu Asp Asn Asp Phe Met Lys Asn		GAT Asp	CTC Leu	AGC Ser	His	GTG Val	ACC Thr	CTG Leu	CCC Pro	Phe	Туг	Pro	: AAG Lys	AGC Ser	Pro	Gln	TCC Ser	1056	
		AAG Lys	GAT Asp	Leu	Ile	AAG Lys	GAA Glu	GCT	Ile	CTT	GAC Asp	AAT Asn	GAC Asp	Phe	Met	AAC Lys	AAC Asn	1104	

						CAG Gln 375										1152
						TGC Cys										1200
						GAT Asp										1248
						GGT Gly										1296
						CGG Arg										1344
						GAT Asp 455										1392
						CAT His										1440
						CTT Leu										1488
															AGG Arg	1536
															GTA Val	1584
		Thr													AGA Arg	1632
	Leu					Trp					Ala				GAA Glu 560	1680
															CTT Leu	1728
														Asp	GAT Asp	1776
GTI	TCT	TAA '	' AAA	GCA	TAT	GAA	GAT	GCA	GAA	GCT	' AAA	GCA	AAA .	TAT	GAA	1824

Val	Ser	Asn 595	Lys	Ala	Tyr	Glu	Asp 600	Ala	Glu	Ala	Lys	Ala 605	Lys	Tyr	Glu	
					TTC Phe											1872
					GTT Val 630											1920
					TCC Ser											1968
					ACA Thr											2016
					GCT Ala											2064
					AAA Lys											2112
					ACC Thr 710											2160
					TAC Tyr											2208
					ATC Ile										CTC Leu	2256
															GCA Ala	2304
		Ile			GGA Gly							Cys			CCA Pro	2352
	Tyr										Gly				TCA Ser 800	2400
GCC Ala	GAC Asp	TAC Tyr	TGG Trp	TCA Ser 805	CTG Leu	GGA Gly	ATC Ile	CTA Leu	ATG Met 810	Tyr	GAA Glu	. CTC . Leu	CTG	ACT Thr 815	GGC	2448
									Met					Ile	ATA Ile	2496

 		_	ATG Met						2544
 	 -		AAA Lys						2592
			AAT Asn 870						2640
			TGG Trp						2688
			GTT Val					_	2736
			AAC Asn						2784
 GAT Asp 930		TTC Phe	TAA						2802

### (2) INFORMATION FOR SEQ ID NO:135:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 933 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:135:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 30 20 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 75 70 80 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly

			115					120					125			
11			Phe	Lys	Glu	Asp			Ile	Leu	Gly			Leu	Glu	Tyr
3.0		130	3	Com	772	<b>.</b>	135		<b>-</b> 1 -			140		<b>~</b> 3 .	_	_
14:		ıyı	ASII	Ser	пте	150		TYE	тте	Met	155		гÀг	GIN	гÀг	
		Ile	Lvs	Val	Asn			Tle	Ara	His			Glu	Aen	Glv	160
	•		-2-		165		-,-		9	170					175	001
Va.	1 (	Gln	Leu	Ala	Asp	His	Tyr	Gln	Gln	Asn		Pro	Ile	Gly		Gly
				180					185					190		_
Pro	0 '	Val	Leu	Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Ala	Leu
_		_	195		_		_	200	_				205		_	
Se		Lys 210	Asp	Pro	Asn	Glu	Lys 215	Arg	Asp	His	Met		Leu	Leu	Glu	Phe
Va			Δla	Δla	Glv	Tle		T.eu	Glv	Met	Aen	220 Glu	Leu	There	Tuc	Ser
22					0	230		LCu	Cry	1100	235		Deu	171	nys	240
Gl	y I	Leu	Arg	Ser	Arg	Gly	Ser	Met	Gly	Thr			Asp	Leu	Gln	
					245					250					255	_
Ala	a i	Leu	Gln		Lys	Ile	Glu	Glu		Arg	Gln	Arg	Asp			Ile
<b>3</b>		<b>~1</b>	T	260	<b>T</b>	01.	<b>T</b>		265	•	_	۵.	_	270		_
AS	p (	GIU	275		Leu	GIU	Leu	280	Gin	Lys	Asp	GIU		He	GIn	Lys
Lei	u (	Gln	_		Leu	Asp	Lvs		Ara	Ser	Val	Ile	285 Ara	Pro	Ala	ጥከተ
		290					295	-3-	3			300	3			
Glı	n (	Gln	Ala	Gln	Lys	Gln	Ser	Ala	Ser	Thr	Leu	Gln	Gly	Glu	Pro	Arg
309						310					315					320
Thi	r J	Lys	Arg	Gln		Ile	Ser	Ala	Glu	Pro	Thr	Ala	Phe	Asp		Gln
λα	<b>.</b> 1	f.e	Ser	Hic	325	ጥኩዮ	Leu	Pro	Pho	330 Tyr	Dro	Tuc	502	Dro	335	Cox
		Jeu	<b>D</b> C1	340	vai	****	Deu	110	345	ıyı	110	Lys	261	350	GIII	261
Lys	s i	Asp	Leu	Ile	Lys	Glu	Ala	Ile		Asp	Asn	Asp	Phe		Lys	Asn
			355					360					365			
Lei			Leu	Ser	Gln	Ile		Glu	Ile	Val	qzA		Met	Tyr	Pro	Val
G1v		370 Tv~	Clv	Lvc	V c.D	Sor	375	T10	T1.	Lys	C1	380	7~~	1703	<b>~1</b>	C
385		-y-	GIY	БуЗ	qen	390	Cys	116	116	Lys	395	GIY	ASD	vaı	GIY	400
		Val	Tyr	Val	Met		Asp	Gly	Lys	Val		Val	Thr	Lys	Glu	
					405			_	_	410				-	415	-
Va]	l I	Lys	Leu		Thr	Met	Gly	Pro	Gly	Lys	Val	Phe	Gly	Glu	Leu	Ala
*1		٠	m	420	<b>~</b>	m	<b>3</b>	m¹-	425	m¹.			m³-	430		_
116	= 1	Leu	1yr 435	ASR	суѕ	ınr	Arg	Thr 440	ΑΙΑ	Thr	vaı	гуѕ	Thr 445	Leu	val	Asn
Va]	LI	Lys		Trp	Ala	Ile	azA		Gln	Cys	Phe	Gln		Ile	Met	Met
		150		•			455	. 5				460				
		Thr	Gly	Leu	Ile		His	Thr	Glu	Tyr	Met	Glu	Phe	Leu	Lys	Ser
465		D-4 -	ent.	D)	<b>63</b>	470	<b>.</b> .	_	~1	~1	475	_	_	_	_	480
Val	L	rro	ınr	rne	Gln 485	ser	Leu	Pro	GIu	Glu 490	Пе	Leu	Ser	Lys		Ala
Asr	<i>,</i>	/al	Leu	Glu		Thr	His	ጥህተ	Glu	Asn	Glv	Glu	ጥረጉ	Tle	495	Δra
				500				-1-	505		013		-7-	510	110	'mg
Glr	1 (	Sly	Ala	Arg	Gly	Asp	Thr	Phe	Phe	Ile	Ile	Ser	Lys		Thr	Val
			515					520					525			
Asn			Thr	Arg	Glu	Asp		Pro	Ser	Glu	Asp		Val	Phe	Leu	Arg
mh x		530	Clar	Tuc	Clv	) co	535	Dho	C1	C1.,	T	540	T 0	C1-	<b>~1</b>	<b>~1</b>
545		Jeu	GTĀ	пХэ	GTĀ	550	тър	FILE	стУ	Glu	ьуs 555	WIG	ьeu	GIU	стλ	560
		/al	Arg	Thr	Ala		Val	Ile	Ala	Ala		Ala	Val	Thr	Cys	
_			-		565					570					575	
Val	. I	le	Asp	Arg	Asp	Ser	Phe	Lys	His	Leu	Ile	Gly	Gly	Leu	Asp	Asp

585 580 Val Ser Asn Lys Ala Tyr Glu Asp Ala Glu Ala Lys Ala Lys Tyr Glu 600 Ala Glu Ala Ala Phe Phe Ala Asn Leu Lys Leu Ser Asp Phe Asn Ile 615 620 Ile Asp Thr Leu Gly Val Gly Gly Phe Gly Arg Val Glu Leu Val Gln 630 635 Leu Lys Ser Glu Glu Ser Lys Thr Phe Ala Met Lys Ile Leu Lys Lys 645 650 Arg His Ile Val Asp Thr Arg Gln Gln Glu His Ile Arg Ser Glu Lys 665 Gln Ile Met Gln Gly Ala His Ser Asp Phe Ile Val Arg Leu Tyr Arg 675 680 685 Thr Phe Lys Asp Ser Lys Tyr Leu Tyr Met Leu Met Glu Ala Cys Leu 690 695 700 Gly Gly Glu Leu Trp Thr Ile Leu Arg Asp Arg Gly Ser Phe Glu Asp 710 715 Ser Thr Thr Arg Phe Tyr Thr Ala Cys Val Val Glu Ala Phe Ala Tyr 730 Leu His Ser Lys Gly Ile Ile Tyr Arg Asp Leu Lys Pro Glu Asn Leu 740 745 Ile Leu Asp His Arg Gly Tyr Ala Lys Leu Val Asp Phe Gly Phe Ala 760 765 Lys Lys Ile Gly Phe Gly Lys Lys Thr Trp Thr Phe Cys Gly Thr Pro 775 780 Glu Tyr Val Ala Pro Glu Ile Ile Leu Asn Lys Gly His Asp Ile Ser 790 795 Ala Asp Tyr Trp Ser Leu Gly Ile Leu Met Tyr Glu Leu Leu Thr Gly 805 810 Ser Pro Pro Phe Ser Gly Pro Asp Pro Met Lys Thr Tyr Asn Ile Ile 820 825 830 Leu Arg Gly Ile Asp Met Ile Glu Phe Pro Lys Lys Ile Ala Lys Asn 840 Ala Ala Asn Leu Ile Lys Lys Leu Cys Arg Asp Asn Pro Ser Glu Arg 850 855 860 Leu Gly Asn Leu Lys Asn Gly Val Lys Asp Ile Gln Lys His Lys Trp 870 875 Phe Glu Gly Phe Asn Trp Glu Gly Leu Arg Lys Gly Thr Leu Thr Pro 885 890 Pro Ile Ile Pro Ser Val Ala Ser Pro Thr Asp Thr Ser Asn Phe Asp 900 905 910 Ser Phe Pro Glu Asp Asn Asp Glu Pro Pro Pro Asp Asp Asn Ser Gly 915 920 925 Trp Asp Ile Asp Phe 930

### (2) INFORMATION FOR SEQ ID NO:136:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2799 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...2795

(D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:136:

	_								GCG Ala 10							48
									GAC Asp							96
									CTG Leu							144
									CAG Gln							192
									ACC Thr							240
	_		_	_			_		GAT Asp 90							288
									AAG Lys							336
									TTG Leu							384
									GAG Glu							432
									CTG Leu							480
									GTG Val 170							528
									ATT Ile							576
									GTA Val							624
CGA	CAA	TGT	TTT	CAA	ACA	ATA	ATG	ATG	AGG	ACA	GGA	CTC	ATC	AAG	CAT	672

Arg	Gln 210	Cys	Phe	Gln	Thr	Ile 215	Met	Met	Arg	Thr	Gly 220	Leu	Ile	Lys	His	
		-							GTT Val							720
									GAT Asp 250							768
									CAA Gln							816
_									AAT Asn							864
									ACT Thr		_					912
									GAT Asp							960
									GTG Val 330							1008
									GTT Val				_		GAA Glu	1056
									GCT Ala							1104
															GGA Gly	1152
									TTG Leu						AAA Lys 400	1200
											_	_		_	AGA Arg	1248
															CAT His	1296
															TAT Tyr	1344

									GGT Gly							1392
				_		_	_		TCT Ser							1440
									CTG Leu 490							1488
									ATC Ile			_				1536
									AAG Lys							1584
									GAG Glu							1632
				_	_		_		GCC Ala						_	1680
				Glu 565					AGC Ser 570			_		_		1728
				ACC	TAT		-		TTG Leu		_	_			_	1776
						_			GCT Ala	_			_			1824
									TTA Leu							1872
									TTT Phe							1920
									CCT Pro 650							1968
									AGT Ser							2016
GAA	CCA	CCA	CCT	GAT	GAC	AAC	TCA	GGA	TGG	GAT	ATA	GAC	TTC	TCG	GAT	2064

Glu	Pro	Pro 675	Pro	Asp	Asp	Asn	Ser 680	Gly	Trp	Asp	Ile	Asp 685	Phe	Ser	Asp	
					ATG Met											2112
					GTC Val 710											2160
					GAG Glu											2208
					TGC Cys											2256
					CTG Leu											2304
					CAG Gln											2352
					CGC Arg 790											2400
			_		GTG Val											2448
					ATC Ile										GGG Gly	2496
					AAC Asn											2544
			_		GGC Gly											2592
					GTG Val 870											2640
					CCC Pro											2688
_	_		_		AGC Ser											2736

GTC CTG CTG GAG TTC GTG ACC GCC GCG GTG ATC ACT CTC GGC ATG GAC 2784
Val Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp
915 920 925

GAG CTG TAC AA GTAA Glu Leu Tyr Lys 930 2799

#### (2) INFORMATION FOR SEQ ID NO:137:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 932 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
  (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:137:

Met Gly Thr Leu Arg Asp Leu Gln Tyr Ala Leu Gln Glu Lys Ile Glu 1 5 10 Glu Leu Arg Gln Arg Asp Ala Leu Ile Asp Glu Leu Glu Leu Glu Leu 25 Asp Gln Lys Asp Glu Leu Ile Gln Lys Leu Gln Asn Glu Leu Asp Lys 40 45 Tyr Arg Ser Val Ile Arg Pro Ala Thr Gln Gln Ala Gln Lys Gln Ser 50 55 60 Ala Ser Thr Leu Gln Gly Glu Pro Arg Thr Lys Arg Gln Ala Ile Ser 70 75 Ala Glu Pro Thr Ala Phe Asp Ile Gln Asp Leu Ser His Val Thr Leu 85 90 Pro Phe Tyr Pro Lys Ser Pro Gln Ser Lys Asp Leu Ile Lys Glu Ala 100 105 110 Ile Leu Asp Asn Asp Phe Met Lys Asn Leu Glu Leu Ser Gln Ile Gln 115 120 125 Glu Ile Val Asp Cys Met Tyr Pro Val Glu Tyr Gly Lys Asp Ser Cys 130 135 140 Ile Ile Lys Glu Gly Asp Val Gly Ser Leu Val Tyr Val Met Glu Asp 145 150 155 Gly Lys Val Glu Val Thr Lys Glu Gly Val Lys Leu Cys Thr Met Gly 165 170 Pro Gly Lys Val Phe Gly Glu Leu Ala Ile Leu Tyr Asn Cys Thr Arg 180 185 190 Thr Ala Thr Val Lys Thr Leu Val Asn Val Lys Leu Trp Ala Ile Asp 195 200 205 Arg Gln Cys Phe Gln Thr Ile Met Met Arg Thr Gly Leu Ile Lys His 210 215 220 Thr Glu Tyr Met Glu Phe Leu Lys Ser Val Pro Thr Phe Gln Ser Leu 230 235 Pro Glu Glu Ile Leu Ser Lys Leu Ala Asp Val Leu Glu Glu Thr His 245 250 255 Tyr Glu Asn Gly Glu Tyr Ile Ile Arg Gln Gly Ala Arg Gly Asp Thr 270 260 265 Phe Phe Ile Ile Ser Lys Gly Thr Val Asn Val Thr Arg Glu Asp Ser

		275					280					285			
Pro	Ser 290		Asp	Pro	Val	Phe 295		Arg	Thr	Leu	Gly 300		Gly	Asp	Trp
Phe 305	Gly	Glu	Lys	Ala	Leu 310	Gln	Gly	Glu	Asp	Val 315	Arg	Thr	Ala	Asn	Val 320
Ile	Ala	Ala	Glu	Ala 325	Val	Thr	Суѕ	Leu	Val 330	Ile	Asp	Arg	Asp	Ser 335	Phe
Lys	His	Leu	Ile 340	Gly	Gly	Leu	Asp	Asp 345	Val	Ser	Asn	Lys	Ala 350	Tyr	Glu
Asp	Ala	Glu 355	Ala	Lys	Ala	Lys	Тут 360	Glu	Ala	Glu	Ala	Ala 365	Phe	Phe	Ala
Asn	Leu 370	Lys	Leu	Ser	Asp	Phe 375	Asn	Ile	Ile	Asp	Thr 380	Leu	Gly	Val	Gly
385		_	_		390					395			Glu		400
				405				_	410				Asp	415	
			420		_			425					Gly 430		
	-	435			•		440	_			-	445	Ser	-	_
	450					455					460		Trp		
465	_	_	_	_	470			_		475			Phe Gly		480
	_			485				-	490				Arg	495	
_	_	_	500	_				505					510 Phe		
		515		_			520					525	Pro		
-	530	_				535				_	540		Ser		_
545			-	_	550	_				555			Ser		560
			_	565					570				Asp	575	
Glu	Phe	Pro	580 Lys	Lys	Ile	Ala	Lys	585 Asn	Ala	Ala	Asn	Leu	590 Ile	Lys	Lys
Leu	Cys	595 Arg	Asp	Asn	Pro	Ser	600 Glu	Arg	Leu	Gly	Asn	605 Leu	Lys	Asn	Gly
Val	610 Lys	Asp	Ile	Gln	Lys	615 His	Lys	Trp	Phe	Glu	620 Gly	Phe	Asn	Trp	Glu
625 Gly		Arg	Lys		630 Thr	Leu	Thr	Pro		635 Ile	Ile	Pro	Ser		640 Ala
Ser	Pro	Thr		645 Thr	Ser	Asn	Phe		650 Ser	Phe	Pro	Glu	Asp	655 Asn	Asp
Glu	Pro		660 Pro	Asp	Asp	Asn		665 Gly	Trp	Asp	Ile		670 Phe	Ser	Asp
Pro		675 Val	Ala	Thr	Met		680 Ser	Lys	Gly	Glu		685 Leu	Phe	Thr	Gly
		Pro	Ile	Leu		695 Glu	Leu	Asp	Gly	Asp 715	700 Val	Asn	Gly	His	Lys 720
705 Phe		Val	Ser		710 Glu	Gly	Glu	Gly	'Asp		Thr	Tyr	Gly	Lys 735	Leu
Thr	Leu	Lys	Phe	725 Ile	Cys	Thr	Thr	Gly		Leu	Pro	Val	Pro		

			740					745					750		
Thr	Leu	Val 755	Thr	Thr	Leu	Thr	Tyr 760	Gly	Val	Gln	Cys	Phe 765	Ser	Arg	Tyr
Pro	Asp 770	His	Met	Lys	Gln	His 775	Asp	Phe	Phe	Lys	Ser 780	Ala	Met	Pro	Glu
Gly 785	Tyr	Val	Gln	Glu	Arg 790	Thr	Ile	Phe	Phe	Lys 795	Asp	Asp	Gly	Asn	Tyr 800
Lys	Thr	Arg	Ala	Glu 805	Val	Lys	Phe	Glu	Gly 810	Asp	Thr	Leu	Val	Asn 815	Arg
Ile	Glu	Leu	Lys 820	Gly	Ile	Asp	Phe	Lys 825	Glu	Asp	Gly	Asn	Ile 830	Leu	Gly
His	Lys	Leu 835	Glu	Tyr	Asn	Tyr	Asn 840	Ser	His	Asn	Val	Tyr 845	Ile	Met	Ala
Asp	Lys 850	Gln	Lys	Asn	Gly	Ile 855	Lys	Val	Asn	Phe	Lys 860	Ile	Arg	His	Asn
Ile 865	Glu	Asp	Gly	Ser	Val 870	Gln	Leu	Ala	Asp	His 875	Tyr	Gln	Gln	Asn	Thr 880
Pro	Ile	Gly	Asp	Gly 885	Pro	Val	Leu	Leu	Pro 890	Asp	Asn	His	Tyr	Leu 895	Ser
Thr	Gln	Ser	Ala 900	Leu	Ser	Lys	Asp	Pro 905	Asn	Glu	Lys	Arg	Asp 910	His	Met
Val	Leu	Leu 915	Glu	Phe	Val	Thr	Ala 920	Ala	Gly	Ile	Thr	Leu 925	Gly	Met	Asp
Glu	Leu 930	Tyr	Lys												

# (2) INFORMATION FOR SEQ ID NO:138:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2184 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2181
  - (D) OTHER INFORMATION:
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:138:

 		GGC Gly 5						48
 	 	GGC Gly	 					96
 	 	GAT Asp	 					144
		AAG Lys						192

CTG ACC TAC GGC GTG CAG TGC TTC AGC CGC TAC CCC GAC CAC ATG AAG Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 65 70 75 80	240
CAG CAC GAC TTC TTC AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 95	288
CGC ACC ATC TTC TTC AAG GAC GAC GGC AAC TAC AAG ACC CGC GCC GAG Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 110	336
GTG AAG TTC GAG GGC GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 115 120 125	384
ATC GAC TTC AAG GAG GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 130 135 140	432
AAC TAC AAC AGC CAC AAC GTC TAT ATC ATG GCC GAC AAG CAG AAG AAC Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 160	480
GGC ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 175	528
GTG CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 180 185 190	576
CCC GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205	624
AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG GAG TTC Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 210 215 220	672
GTG ACC GCC GCC GGG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TCC Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 225 230 235 240	720
GGA CTC AGA TCT CGA GGC ACC ATG AGC GAC GTG GCT ATT GTG AAG GAG Gly Leu Arg Ser Arg Gly Thr Met Ser Asp Val Ala Ile Val Lys Glu 245 250 255	768
GGT TGG CTG CAC AAA CGA GGG GAG TAC ATC AAG ACC TGG CGG CCA CGC Gly Trp Leu His Lys Arg Gly Glu Tyr Ile Lys Thr Trp Arg Pro Arg 260 265 270	816
TAC TTC CTC CTC AAG AAT GAT GGC ACC TTC ATT GGC TAC AAG GAG CGG Tyr Phe Leu Leu Lys Asn Asp Gly Thr Phe Ile Gly Tyr Lys Glu Arg 275 280 285	864
CCG CAG GAT GTG GAC CAA CGT GAG GCT CCC CTC AAC AAC TTC TCT GTG	912

Pro G	ln <i>1</i> 90	Asp	Val	Asp	Gln	Arg 295	Glu	Ala	Pro	Leu	Asn 300	Asn	Phe	Ser	Val	
GCG CA Ala GI 305																960
ATC AT																1008
GTG G																1056
GTG GG Val A	la A													_		1104
TCG GG																1152
CTG G Leu A 385													_			1200
AAG C																1248
AAG G										_				_	_	1296
ATC G	al z											_			_	1344
CTG C. Leu G. 4																1392
CAG AGGIN TI																1440
GAG C																1488
GCC CO																1536
TCG G	lu 1															1584

					CAC His						1632
					GGT Gly 550						1680
_			_		GAG Glu						1728
					CTG Leu						1776
			_		AAC Asn						1824
					CGC Arg						1872
				_	CTG Leu 630						1920
_	_		_		GCC Ala						1968
_	_	_			CAC His						2016
					TCG Ser						2064
					ATC Ile						2112
					AGC Ser 710						2160
					ACG Thr	TGA					2184

### (2) INFORMATION FOR SEQ ID NO:139:

# (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 727 amino acids

- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:139:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 40 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys .70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 100 105 110 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 130 135 140 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 145 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 170 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 180 185 190 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 195 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 225 230 235 Gly Leu Arg Ser Arg Gly Thr Met Ser Asp Val Ala Ile Val Lys Glu 245 250 Gly Trp Leu His Lys Arg Gly Glu Tyr Ile Lys Thr Trp Arg Pro Arg 260 265 Tyr Phe Leu Leu Lys Asn Asp Gly Thr Phe Ile Gly Tyr Lys Glu Arg 275 280 285 Pro Gln Asp Val Asp Gln Arg Glu Ala Pro Leu Asn Asn Phe Ser Val 300 290 295 Ala Gln Cys Gln Leu Met Lys Thr Glu Arg Pro Arg Pro Asn Thr Phe 310 315 Ile Ile Arg Cys Leu Gln Trp Thr Thr Val Ile Glu Arg Thr Phe His 330 325 Val Glu Thr Pro Glu Glu Arg Glu Glu Trp Thr Thr Ala Ile Gln Thr 340 350 345 Val Ala Asp Gly Leu Lys Lys Gln Glu Glu Glu Met Asp Phe Arg 355 360 365 Ser Gly Ser Pro Ser Asp Asn Ser Gly Ala Glu Glu Met Glu Val Ser 375 380

Leu Ala Lys Pro Lys His Arg Val Thr Met Asn Glu Phe Glu Tyr Leu

385					390					395					400
Lys	Leu	Leu	Gly	Lys 405	Gly	Thr	Phe	Gly	Lys 410	Val	Ile	Leu	Val	Lys 415	Glu
Lys	Ala	Thr	Gly 420	Arg	Tyr	Tyr	Ala	Met 425	Lys	Ile	Leu	Lys	Lys 430	Glu	Val
Ile	Val	Ala 435	Lys	Asp	Glu	Val	Ala 440	His	Thr	Leu	Thr	Glu 445	Asn	Arg	Val
Leu	Gln 450	Asn	Ser	Arg	His	Pro 455	Phe	Leu	Thr	Ala	Leu 460	Lys	Tyr	Ser	Phe
Gln	Thr	His	Asp	Arg	Leu	Cys	Phe	Val	Met	Glu	Tyr	Ala	Asn	Gly	Gly
465					470					475					480
				485	Leu				490					495	
Ala	Arg	Phe	Tyr 500	Gly	Ala	Glu	Ile	Val 505	Ser	Ala	Leu	Asp	Туг 510	Leu	His
Ser	Glu	Lys 515	Asn	Val	Val	Tyr	Arg 520	Asp	Leu	Lys	Leu	Glu 525	Asn	Leu	Met
Leu	Asp 530	Lys	Asp	Gly	His	Ile 535	Lys	Ile	Thr	Asp	Phe 540	Gly	Leu	Cys	Lys
Glu	Gly	Ile	Lys	Asp	Gly	Ala	Thr	Met	Lys	Thr	Phe	Cys	Gly	Thr	Pro
545					550					555					560
Glu	Tyr	Leu	Ala	Pro 565	Glu	Val	Leu	Glu	Asp 570	Asn	Asp	Tyr	Gly	Arg 575	Ala
Val	Asp	Trp	Trp 580	Gly	Leu	Gly	Val	Val 585	Met	Tyr	Glu	Met	Met 590	Cys	Gly
Arg	Leu	Pro 595	Phe	Tyr	Asn	Gln	Asp 600	His	Glu	Lys	Leu	Phe 605	Glu	Leu	Ile
Leu	Met 610	Glu	Glu	Ile	Arg	Phe 615	Pro	Arg	Thr	Leu	Gly 620	Pro	Glu	Ala	Lys
Ser 625	Leu	Leu	Ser	Gly	Leu 630	Leu	Lys	Lys	Asp	Pro 635	Lys	Gln	Arg	Leu	Gly 640
Gly	Gly	Ser	Glu	Asp 645	Ala	Lys	Glu	Ile	Met 650	Gln	His	Arg	Phe	Phe 655	Ala
Gly	Ile	Val	Trp 660	Gln	His	Val	Tyr	Glu 665	Lys	Lys	Leu	Ser	Pro 670	Pro	Phe
Lys	Pro	Gln 675	Val	Thr	Ser	Glu	Thr 680	Asp	Thr	Arg	Tyr	Phe 685	Asp	Glu	Glu
Phe	Thr 690	Ala	Gln	Met	Ile	Thr 695	Ile	Thr	Pro	Pro	Asp 700	Gln	Asp	Asp	Ser
Met 705	Glu	Cys	Val	Asp	Ser 710	Glu	Arg	Arg	Pro	His 715	Phe	Pro	Gln	Phe	Ser 720
Tyr	Ser	Ala	Ser	Ser 725	Thr	Ala									

# (2) INFORMATION FOR SEQ ID NO:140:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2394 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2391
  - (D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:140:

		CCC Pro										48
		GAG Glu										96
		TGC Cys										144
		ACC Thr										192
		GGG Gly 70		_			_	_			:	240
		CAC His										288
		GCT Ala						_				336
		ATC Ile										384
	_	ATC Ile	_	_			_	_	_			432
		GGG Gly 150										480
		CGG Arg										528
		CCC Pro								GCC Ala		576
										GGT Gly		624
 	 	 				-				ATT Ile		672

GAG GTG TAT TTC Glu Val Tyr Phe 225					720
CAA GCT GAT GTG Gln Ala Asp Val				_	768
			CGT GTC TCC ATC Arg Val Ser Med 270	Gln Leu	816
CGG CGG CCT TCC Arg Arg Pro Ser 275				_	864
CTG CCA GAT ACA Leu Pro Asp Thr 290					912
			AAG AGT CCT TTV Lys Ser Pro Pho 315		960
			ATT GCT GTG CC		1008
			CAG CCC TAT CC Gln Pro Tyr Pro 35	Phe Thr	1056
			TTT CCC ACC ATC Phe Pro Thr Me 365		1104
		Ala Ser Ala	TTG GCC CCG GC Leu Ala Pro Al 380		1152
			CCT GCT CCA GC Pro Ala Pro Al 395		1200
			CCA GTC CTA GC Pro Val Leu Al		1248
	Val Ala Pro		: AAG CCC ACC CA : Lys Pro Thr Gl 43	n Ala Gly	1296
			CTG CAG TTT GA Leu Gln Phe As 445	_	1344
GAC CTG GGG GCC	TTG CTT GGC	AAC AGC ACA	GAC CCA GCT GT	G TTC ACA	1392

Ası	Leu 450	Gly	Ala	Leu	Leu	Gly 455	Asn	Ser	Thr	Asp	Pro 460	Ala	Val	Phe	Thr	
	CTG Leu									-	_				_	1440
	C ATA / Ile															1488
	r GAG o Glu		_						_	_	_					1536
	A GCT o Ala												_			1584
	A GGA r Gly 530					_										1632
= =	G CTG 1 Leu 5		_								_					1680
	C AAG r Lys															1728
	G GAC u Asp															1776
	G GGC u Gly															1824
	c GGC r Gly 610								_		_	_	_			1872
	c GGC r Gly 5															1920
	C TTC p Phe															1968
	C TTC e Phe															2016
	C GAG e Glu															2064

		AAC Asn						2112
		тат туг 710						2160
		ATC Ile						2208
		CAG Gln					 	2256
		CĄC His						2304
		CGC Arg						2352
		CTC Leu 790				TAA		2394

#### (2) INFORMATION FOR SEQ ID NO:141:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 797 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:141:

Met Asp Glu Leu Phe Pro Leu Ile Phe Pro Ala Glu Pro Ala Gln Ala 1 5 10 15 Ser Gly Pro Tyr Val Glu Ile Ile Glu Gln Pro Lys Gln Arg Gly Met 20 25 30 Arg Phe Arg Tyr Lys Cys Glu Gly Arg Ser Ala Gly Ser Ile Pro Gly 35 40 45 Glu Arg Ser Thr Asp Thr Thr Lys Thr His Pro Thr Ile Lys Ile Asn 55 60 Gly Tyr Thr Gly Pro Gly Thr Val Arg Ile Ser Leu Val Thr Lys Asp 70 75 Pro Pro His Arg Pro His Pro His Glu Leu Val Gly Lys Asp Cys Arg 85 90 95 Asp Gly Phe Tyr Glu Ala Glu Leu Cys Pro Asp Arg Cys Ile His Ser 100 105 110 Phe Gln Asn Leu Gly Ile Gln Cys Val Lys Lys Arg Asp Leu Glu Gln

Ala Ile Ser Gln Arg Ile Gln Thr Asn Asn Asn Pro Phe Gln Val Pro Ile Glu Glu Gln Arg Gly Asp Tyr Asp Leu Asn Ala Val Arg Leu Cys Phe Gln Val Thr Val Arg Asp Pro Ser Gly Arg Pro Leu Arg Leu Pro Pro Val Leu Pro His Pro Ile Phe Asp Asn Arg Ala Pro Asn Thr Ala Glu Leu Lys Ile Cys Arg Val Asn Arg Asn Ser Gly Ser Cys Leu Gly Gly Asp Glu Ile Phe Leu Cys Asp Lys Val Gln Lys Glu Asp Ile Glu Val Tyr Phe Thr Gly Pro Gly Trp Glu Ala Arg Gly Ser Phe Ser Gln Ala Asp Val His Arg Gln Val Ala Ile Val Phe Arg Thr Pro Pro Tyr Ala Asp Pro Ser Leu Gln Ala Pro Val Arg Val Ser Met Gln Leu Arg Arg Pro Ser Asp Arg Glu Leu Ser Glu Pro Met Glu Phe Gln Tyr Leu Pro Asp Thr Asp Asp Arg His Arg Ile Glu Glu Lys Arg Lys Arg Thr Tyr Glu Thr Phe Lys Ser Ile Met Lys Lys Ser Pro Phe Ser Gly Pro Thr Asp Pro Arg Pro Pro Pro Arg Arg Ile Ala Val Pro Ser Arg Ser Ser Ala Ser Val Pro Lys Pro Ala Pro Gln Pro Tyr Pro Phe Thr Ser Ser Leu Ser Thr Ile Asn Tyr Asp Glu Phe Pro Thr Met Val Phe Pro Ser Gly Gln Ile Ser Gln Ala Ser Ala Leu Ala Pro Ala Pro Pro Gln Val Leu Pro Gln Ala Pro Ala Pro Ala Pro Ala Pro Ala Met Val Ser Ala Leu Ala Gln Ala Pro Ala Pro Val Pro Val Leu Ala Pro Gly Pro Pro Gln Ala Val Ala Pro Pro Ala Pro Lys Pro Thr Gln Ala Gly Glu Gly Thr Leu Ser Glu Ala Leu Leu Gln Leu Gln Phe Asp Asp Glu Asp Leu Gly Ala Leu Leu Gly Asn Ser Thr Asp Pro Ala Val Phe Thr Asp Leu Ala Ser Val Asp Asn Ser Glu Phe Gln Gln Leu Leu Asn Gln Gly Ile Pro Val Ala Pro His Thr Thr Glu Pro Met Leu Met Glu Tyr Pro Glu Ala Ile Thr Arg Leu Val Thr Gly Ala Gln Arg Pro Pro Asp Pro Ala Pro Ala Pro Leu Gly Ala Pro Gly Leu Pro Asn Gly Leu Leu Ser Gly Asp Glu Asp Phe Ser Ser Ile Ala Asp Met Asp Phe Ser Ala Leu Leu Ser Gln Ile Ser Ser Leu Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly



			580					585					590				
Glu	Gly	Asp 595	Ala	Thr	Tyr	Gly	Lys 600	Leu	Thr	Leu	Lys	Phe 605	Ile	Cys	Thr		
Thr	Gly 610	Lys	Leu	Pro	Val	Pro 615	Trp	Pro	Thr	Leu	Val 620	Thr	Thr	Leu	Thr		
Tyr 625	Gly	Val	Gln	Cys	Phe 630	Ser	Arg	Tyr	Pro	Asp 635	His	Met	Lys	Gln	His 640		
Asp	Phe	Phe	Lys	Ser 645	Ala	Met	Pro	Glu	Gly 650	Tyr	Val	Gln	Glu	Arg 655	Thr		
Ile	Phe	Phe	Lys 660	Asp	Asp	Gly	Asn	Tyr 665	Lys	Thr	Arg	Ala	Glu 670	Val	Lys		
Phe	Glu	Gly 675	Asp	Thr	Leu	Val	Asn 680	Arg	Ile	Glu	Leu	Lys 685	Gly	Ile	Asp		
Phe	Lys 690	Glu	Asp	Gly	Asn	Ile 695	Leu	Gly	His	Lys	Leu 700	Glu	Tyr	Asn	Tyr		
Asn 705	Ser	His	Asn	Val	Tyr 710	Ile	Met	Ala	Asp	Lys 715	Gln	Lys	Asn	Gly	Ile 720		
Lys	Val	Asn	Phe	Lys 725	Ile	Arg	His	Asn	Ile 730	Glu	Asp	Gly	Ser	Val 735	Gln		
Leu	Ala	Asp	His 740	Tyr	Gln	Gln	Asn	Thr 745	Pro	Ile	Gly	Asp	Gly 750	Pro	Val		
Leu	Leu	Pro 755	Asp	Asn	His	Tyr	Leu 760	Ser	Thr	Gln	Ser	Ala 765	Leu	Ser	Lys		
Asp	Pro 770	Asn	Glu	Lys	Arg	Asp 775	His	Met	Val	Leu	Leu 780	Glu	Phe	Val	Thr		
Ala 785	Ala	Gly	Ile	Thr	Leu 790	Gly	Met	Asp	Glu	Leu 795	Tyr	Lys					
		(2)	IN	FORM	OITA	v FOI	R SEX	Q ID	NO:	142:							
	( :				CHAR												
					2394		_	airs									
					ole: ONES			_									
					7: 1:		_	<del>.</del>									
		ii) N ix) I			TYPI	E: cI	ONA										
	, -	., .															
		(A)	NAI	Æ/KI	Y: (	Codi	ng Se	eque	nce								
					ON:												
		(D)	OTE	IER .	INFO	MAT.	LON:										
	(3	ki) S	SEQUI	ENCE	DESC	CRIP	rion	: SE	Q ID	NO:	142:						
									ACC							4	8
Met 1	Val	Ser	гуѕ	Gly 5	Glu	Glu	Leu		Thr 10	_				Ile 15	Leu		

GTC GAG CTG GAC GGC GAC GTA AAC GGC CAC AAG TTC AGC GTG TCC GGC Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly

25

GAG GGC GAG GGC GAT GCC ACC TAC GGC AAG CTG ACC CTG AAG TTC ATC

Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile  $35 \hspace{1.5cm} 40 \hspace{1.5cm} 45$ 

TGC ACC ACC GGC AAG CTG CCC GTG CCC TGG CCC ACC CTC GTG ACC ACC

144

192

20

Cys	Thr 50	Thr	Gly	Lys	Leu	Pro 55	Val	Pro	Trp	Pro	Thr 60	Leu	Val	Thr	Thr	
					CAG Gln 70											240
	•				AAG Lys											288
					AAG Lys											336
					GAC Asp											384
					GAC Asp											432
					AAC Asn 150											480
					TTC Phe											528
					CAC His											576
					GAC Asp											624
					GAG Glu											672
					ATC Ile 230											720
					GCC Ala											768
					GCC Ala											816
					ATG Met											864

GCG GGC AGC ATC Ala Gly Ser Ile 290					912
CCC ACC ATC AAG Pro Thr Ile Lys 305					960
TCC CTG GTC ACC Ser Leu Val Thr			Pro His Pro		800
GTA GGA AAG GAC Val Gly Lys Asp 340	Cys Arg Asp				056
GAC CGC TGC ATC Asp Arg Cys Ile 355					104
AAG CGG GAC CTG Lys Arg Asp Leu 370					152
AAC CCC TTC CAA Asn Pro Phe Gln 385			_		200
AAT GCT GTG CGG Asn Ala Val Arg			Val Arg Asp		248
AGG CCC CTC CGC Arg Pro Leu Arg 420	Leu Pro Pro				296
CGT GCC CCC AAC Arg Ala Pro Asn 435					.344
TCT GGC AGC TGC Ser Gly Ser Cys 450					.392
GTG CAG AAA GAG Val Gln Lys Glu 465					440
GCC CGA GGC TCC Ala Arg Gly Ser			His Arg Gln		488
GTG TTC CGG ACC Val Phe Arg Thr 500	Pro Pro Tyr				1536
CGT GTC TCC ATG	CAG CTG CGG	CGG CCT TCC	GAC CGG GAG	CTC AGT GAG 1	1584

Arg Val Ser 515	Met Gln Leu	Arg Arg Pr 520	o Ser Asp	Arg Glu Le 525	u Ser Glu	
	TTC CAG TAC Phe Gln Tyr					1632
	CGT AAA AGG Arg Lys Arg 550	Thr Tyr Gl				1680
	TTC AGC GGA Phe Ser Gly 565					1728
	CCT TCC CGC Pro Ser Arg 580		a Ser Val		o Ala Pro	1776
	CCC TTT ACG					1824
	ATG GTG TTT Met Val Phe				_	1872
	GCC CCT CCC Ala Pro Pro 630	Gln Val Le				1920
	GCC ATG GTA Ala Met Val 645			_	_	1968
	GCC CCA GGC Ala Pro Gly 660		ln Ala Val		o Ala Pro	2016
	CAG GCT GGG				_	
	GAT GAT GAA Asp Asp Glu			_	_	2112
	GTG TTC ACA Val Phe Thr 710	Asp Leu Al				
	CTG AAC CAG Leu Asn Gln 725					
	ATG GAG TAC Met Glu Tyr 740		a Ile Thr		al Thr Gly	

GCC CAG AGG CCC CCC GAC CCA GCT CCT GCT CCA CTG GGG GCC CCG GGG
Ala Gln Arg Pro Pro Asp Pro Ala Pro Ala Pro Leu Gly Ala Pro Gly
755 760 765

CTC CCC AAT GGC CTC CTT TCA GGA GAT GAA GAC TTC TCC TCC ATT GCG 2352

CTC CCC AAT GGC CTC CTT TCA GGA GAT GAA GAC TTC TCC TCC ATT GCG 2352
Leu Pro Asn Gly Leu Leu Ser Gly Asp Glu Asp Phe Ser Ser Ile Ala
770 780

GAC ATG GAC TTC TCA GCC CTG CTG AGT CAG ATC AGC TCC TAA 2394
Asp Met Asp Phe Ser Ala Leu Leu Ser Gln Ile Ser Ser
785 790 795

#### (2) INFORMATION FOR SEQ ID NO:143:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 797 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
  (v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:143:

Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu 10 Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly 20 25 Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile 35 40 45 Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr 55 60 Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys 70 75 Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu 85 90 Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu 105 Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly 120 125 Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr 140 135 Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn 150 155 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 165 170 175 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 180 185 190 Pro Val Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 200 205 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 215 220 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys Ser 230 235 Gly Leu Arg Ser Arg Ala Met Asp Glu Leu Phe Pro Leu Ile Phe Pro

Ala Glu Pro Ala Gln Ala Ser Gly Pro Tyr Val Glu Ile Ile Glu Gln Pro Lys Gln Arg Gly Met Arg Phe Arg Tyr Lys Cys Glu Gly Arg Ser Ala Gly Ser Ile Pro Gly Glu Arg Ser Thr Asp Thr Thr Lys Thr His Pro Thr Ile Lys Ile Asn Gly Tyr Thr Gly Pro Gly Thr Val Arg Ile Ser Leu Val Thr Lys Asp Pro Pro His Arg Pro His Pro His Glu Leu Val Gly Lys Asp Cys Arg Asp Gly Phe Tyr Glu Ala Glu Leu Cys Pro Asp Arg Cys Ile His Ser Phe Gln Asn Leu Gly Ile Gln Cys Val Lys Lys Arg Asp Leu Glu Gln Ala Ile Ser Gln Arg Ile Gln Thr Asn Asn Asn Pro Phe Gln Val Pro Ile Glu Glu Gln Arg Gly Asp Tyr Asp Leu Asn Ala Val Arg Leu Cys Phe Gln Val Thr Val Arg Asp Pro Ser Gly Arg Pro Leu Arg Leu Pro Pro Val Leu Pro His Pro Ile Phe Asp Asn Arg Ala Pro Asn Thr Ala Glu Leu Lys Ile Cys Arg Val Asn Arg Asn Ser Gly Ser Cys Leu Gly Gly Asp Glu Ile Phe Leu Leu Cys Asp Lys 450 455 Val Gln Lys Glu Asp Ile Glu Val Tyr Phe Thr Gly Pro Gly Trp Glu Ala Arg Gly Ser Phe Ser Gln Ala Asp Val His Arg Gln Val Ala Ile Val Phe Arg Thr Pro Pro Tyr Ala Asp Pro Ser Leu Gln Ala Pro Val Arg Val Ser Met Gln Leu Arg Arg Pro Ser Asp Arg Glu Leu Ser Glu Pro Met Glu Phe Gln Tyr Leu Pro Asp Thr Asp Asp Arg His Arg Ile Glu Glu Lys Arg Lys Arg Thr Tyr Glu Thr Phe Lys Ser Ile Met Lys Lys Ser Pro Phe Ser Gly Pro Thr Asp Pro Arg Pro Pro Pro Arg Arg Ile Ala Val Pro Ser Arg Ser Ser Ala Ser Val Pro Lys Pro Ala Pro 580 585 Gln Pro Tyr Pro Phe Thr Ser, Ser Leu Ser Thr Ile Asn Tyr Asp Glu Phe Pro Thr Met Val Phe Pro Ser Gly Gln Ile Ser Gln Ala Ser Ala Leu Ala Pro Ala Pro Pro Gln Val Leu Pro Gln Ala Pro Ala Pro Ala Pro Ala Pro Ala Met Val Ser Ala Leu Ala Gln Ala Pro Ala Pro Val Pro Val Leu Ala Pro Gly Pro Pro Gln Ala Val Ala Pro Pro Ala Pro Lys Pro Thr Gln Ala Gly Glu Gly Thr Leu Ser Glu Ala Leu Leu Gln Leu Gln Phe Asp Asp Glu Asp Leu Gly Ala Leu Leu Gly Asn Ser Thr Asp Pro Ala Val Phe Thr Asp Leu Ala Ser Val Asp Asn Ser Glu Phe

70		_			71					71					720	
GI	n Gl	n Le	u Lei	ມ As: 72		n Gly	y Ile	e Pro	o Va. 73		a Pr	o Hi	s Th		r Glu	
Pr	o Me	t Le	u Me			r Pro	Glı	ı Ala			r Ar	g Lei	ı Va	73! 1 Th:	r Gly	
			740	)				749	5				75	0		
		75	5				760	)				765	5		Gly	
	77	0				775	5				78	0		r Ile	e Ala	
As:	p Me <sup>.</sup> 5	t As	p Phe	e Sei	r Ala 790		ı Leı	ı Ser	Glr	ı Ile 79		r Sei	:			
	ı		2) IN SEQUE							144:	:					
			) LEN													
		(B)	TYP	E: n	ucle	ic a	cid									
			STR TOP					е								
		(D)	101	OLCG	X: 1	inea	r									
	(	ii)	MOLE	CULE	TYP	E: c	DNA									
	(	ix)	FEAT	URE:												
		(E	A) NAI B) LOO D) OTI	CATI HER	ON: INFO	1:	3378 ION:			110	<b>.</b>					
	•	<b>A1</b> /	SEQUI	CINCE	DES	CRIP.	LTON	: SE	Q ID	NO:	144:					
ATG	GAG	CGG	GCC	GGC	CCC	AGC	TTC	GGG	CAG	CAG	CGA	CAG	CAG	CAG	CAG	48
Met 1	Glu	Arg	Ala	Gly 5	Pro	Ser	Phe	Gly		Gln	Arg	Gln	Gln		Gln	
-				3					10					15		
CCC	CAG	CAG	CAG	AAG	CAG	CAG	CAG	AGG	GAT	CAG	GAC	TCG	GTC	GAA	GCA	96
Pro	Gln	Gln	Gln 20	Lys	Gln	Gln	Gln	Arg 25	Asp	Gln	Asp	Ser	Val 30	Glu	Ala	
TGG	CTG	GAC	GAT	CAC	TGG	GAC	TTT	ACC	TTC	TCA	TAC	TTT.	GTT	AGA	AAA	144
Trp	Leu	Asp	Asp	His	Trp	Asp	Phe	Thr	Phe	Ser	Tyr	Phe	Val	Arg	Lys	
		35					40					45				
GCC	ACC	AGA	GAA	ATG	GTC	AAT	GCA	TGG	TTT	GCT	GAG	AGA	GTT	CAC	ACC	192
Ala	Thr	Arg	Glu	Met	Val	Asn	Ala	Trp	Phe	Ala	Glu	Arg	Val	His	Thr	172
	50					55					60					
ATC	ССТ	GTG	TGC	AAG	GAA	GGT	ATC	AGA	GGC	CAC	ACC	GAA	ጥርጥ	ጥርረ	ጥረጥ	240
Ile	Pro	Val	Cys	Lys	Glu	Gly	Ile	Arg	Gly	His	Thr	Glu	Ser	Cys	Ser	240
65					70					75					80	

TGT CCC TTG CAG CAG AGT CCT CGT GCA GAT AAC AGT GTC CCT GGA ACA

Cys Pro Leu Gln Gln Ser Pro Arg Ala Asp Asn Ser Val Pro Gly Thr

85 90 95 CCA ACC AGG AAA ATC TCT GCC TCT GAA TTT GAC CGG CCT CTT AGA CCC 336 Pro Thr Arg Lys Ile Ser Ala Ser Glu Phe Asp Arg Pro Leu Arg Pro 100 105 ATT GTT GTC AAG GAT TCT GAG GGA ACT GTG AGC TTC CTC TCT GAC TCA 384 Ile Val Val Lys Asp Ser Glu Gly Thr Val Ser Phe Leu Ser Asp Ser 115 120 GAA AAG AAG GAA CAG ATG CCT CTA ACC CCT CCA AGG TTT GAT CAT GAT 432 Glu Lys Lys Glu Gln Met Pro Leu Thr Pro Pro Arg Phe Asp His Asp 130 135 140 GAA GGG GAC CAG TGC TCA AGA CTC TTG GAA TTA GTG AAG GAT ATT TCT 480 Glu Gly Asp Gln Cys Ser Arg Leu Leu Glu Leu Val Lys Asp Ile Ser 145 150 155 160 AGT CAT TTG GAT GTC ACA GCC TTA TGT CAC AAA ATT TTC TTG CAT ATC 528 Ser His Leu Asp Val Thr Ala Leu Cys His Lys Ile Phe Leu His Ile CAT GGA CTG ATA TCT GCT GAC CGC TAT TCC CTG TTC CTT GTC TGT GAA 576 His Gly Leu Ile Ser Ala Asp Arg Tyr Ser Leu Phe Leu Val Cys Glu 180 GAC AGC TCC AAT GAC AAG TTT CTT ATC AGC CGC CTC TTT GAT GTT GCT 624 Asp Ser Ser Asn Asp Lys Phe Leu Ile Ser Arg Leu Phe Asp Val Ala 200 GAA GGT TCA ACA CTG GAA GAA GTT TCA AAT AAC TGT ATC CGC TTA GAA 672 Glu Gly Ser Thr Leu Glu Glu Val Ser Asn Asn Cys Ile Arg Leu Glu 215 TGG AAC AAA GGC ATT GTG GGA CAT GTG GCA GCG CTT GGT GAG CCC TTG Trp Asn Lys Gly Ile Val Gly His Val Ala Ala Leu Gly Glu Pro Leu AAC ATC AAA GAT GCA TAT GAG GAT CCT CGG TTC AAT GCA GAA GTT GAC 768 Asn Ile Lys Asp Ala Tyr Glu Asp Pro Arg Phe Asn Ala Glu Val Asp 245 250 CAA ATT ACA GGC TAC AAG ACA CAA AGC ATT CTT TGT ATG CCA ATT AAG 816 Gln Ile Thr Gly Tyr Lys Thr Gln Ser Ile Leu Cys Met Pro Ile Lys 260 265 AAT CAT AGG GAA GAG GTT GTT GGT GTA GCC CAG GCC ATC AAC AAA 864 Asn His Arg Glu Glu Val Val Gly Val Ala Gln Ala Ile Asn Lys Lys 275 280 285 TCA GGA AAC GGT GGG ACA TTT ACT GAA AAA GAT GAA AAG GAC TTT GCT Ser Gly Asn Gly Gly Thr Phe Thr Glu Lys Asp Glu Lys Asp Phe Ala

290

295

GCT	TAT	TTG	GCA	TTT	TGT	GGT	ATT	GTT	CTT	CAT	ААТ	GCT	CAG	CTC	TAT	960
Ala	Tyr	Leu	Ala	Phe	Cys	Gly	Ile	Val	Leu	His	Asn	Ala	Gln	Leu	Tyr	
305					310					315					320	
GAG	ACT	TCA	CTG	CTG	GAG	AAC	AAG	AGA	AAT	CAG	GTG	CTG	CTT	GAC	CTT	1008
Glu	Thr	Ser	Leu	Leu	Glu	Asn	Lys	Arg	Asn	Gln	Val	Leu	Leu	Asp,	Leu	
				325					330					335		
	AGT															1056
Ala	Ser	Leu		Phe	Glu	Glu	Gln	_	Ser	Leu	Glu	Val		Leu	Lys	
			340					345					350			
222	2002	CCM	CCC	3.00	3 000	3 m/a	mam	mmo	3.000	<b>~~</b>	omo.	~>~		maa		1104
	ATA														•	1104
гЛS	Ile	355	AId	THE	He	me	360	Pne	Met	GIN	vaı		гÀ2	Cys	inr	
		333					300					365				
עוייירע	TTC	בידב	CITC	СУТ	CAA	ርልጥ	шсс.	TYCC	СУТ	цСц	עארעי	тст	ልርጥ	CTC	undah.	1152
_	Phe	_	_		_											1132
110	370		•		O.Lu	375	Cys	DCI	r.cp	DCI	380	DCI	DCI	VUI	1110	
	5.0					5,5					300					
CAC	ATG	GAG	TGT	GAG	GAA	тта	GAA	AAA	TCA	тст	GAT	ACA	TTA	ACA	AGG	1200
_	Met	_		_												
385			-		390			-		395	-				400	
GAA	CAT	GAT	GCA	AAC	AAA	ATC	ААТ	TAC	ATG	TAT	GCT	CAG	TAT	GTC	AAA	1248
Glu	His	Asp	Ala	Asn	Lys	Ile	Asn	Tyr	Met	Tyr	Ala	Gln	Tyr	Val	Lys	
				405					410					415		
AAT	ACT	ATG	GAA	CCA	CTT	AAT	ATC	CCA	GAT	GTC	AGT	AAG	GAT	AAA	AGA	1296
Asn	Thr	Met		Pro	Leu	Asn	Ile	Pro	Asp	Val	Ser	Lys	-	Lys	Arg	
			420					425					430			
		maa						~~-		~			~~~			1244
	CCC															1344
Prie	Pro	435	THE	THE	GIU	ASI	440	GIŸ	ASI	vai	ASI		GIN	суs	116	
		400					440					445				
AGA	AGT	TTTC	Стт	ጥርጥ	מרמ	CCT	מידמ	222	<u>አ</u> አጥ	CCA	ΔΔC	AAG	ייעע	מממ	Cum	1392
	Ser															1372
5	450			0,70		455		_,_		013	460	2,2		_,0	<b>1</b> 44	
ATA	GGG	GTT	TGC	CAA	CTT	GTT	ААТ	AAG	ATG	GAG	GAG	AAT	ACT	GGC	AAG	1440
Ile	Gly	Val	Cys	Gln	Leu	Val	Asn	Lys	Met	Glu	Glu	Asn	Thr	Gly	Lys	
465					470					475					480	
GTT	AAG	CCT	TTC	AAC	CGA	TAA	GAC	GAA	CAG	TTT	CTG	GAA	GCT	TTT	GTC	1488
Val	Lys	Pro	Phe	Asn	Arg	Asn	qzA	Glu <sup>.</sup>	Gln	Phe	Leu	Glu	Ala	Phe	Val	
				485					490					495		
	TTT															1536
Ile	Phe	Cys		Leu	Gly	Ile	Gln		Thr	Gln	Met	Tyr		Ala	Val	
			500					505					510			
030	202	~~~	»			<i>~</i>	3 m~	OFF-C	202	mm~	~~	- CETT	~~~	m	m» m	1504
	AGA															1584
GIU	Arg	via	rie c	wrq	ηλρ	GIII	nec	val	TIIL	reu	GIU	vaı	ьeu	ser	TAL	

515 520 525 CAT GCT TCA GCA GCA GAG GAA GAA ACA AGA GAG CTA CAG TCG TTA GCG 1632 His Ala Ser Ala Ala Glu Glu Glu Thr Arg Glu Leu Gln Ser Leu Ala 535 540 GCT GCT GTG GTG CCA TCT GCC CAG ACC CTT AAA ATT ACT GAC TTT AGC 1680 Ala Ala Val Val Pro Ser Ala Gln Thr Leu Lys Ile Thr Asp Phe Ser 550 555 TTC AGT GAC TTT GAG CTG TCT GAT CTG GAA ACA GCA CTG TGC ACA ATT 1728 Phe Ser Asp Phe Glu Leu Ser Asp Leu Glu Thr Ala Leu Cys Thr Ile 565 570 CGG ATG TTT ACT GAC CTC AAC CTT GTG CAG AAC TTC CAG ATG AAA CAT 1776 Arg Met Phe Thr Asp Leu Asn Leu Val Gln Asn Phe Gln Met Lys His 580 585 GAG GTT CTT TGC AGA TGG ATT TTA AGT GTT AAG AAG AAT TAT CGG AAG Glu Val Leu Cys Arg Trp Ile Leu Ser Val Lys Lys Asn Tyr Arg Lys 600 595 AAT GTT GCC TAT CAT AAT TGG AGA CAT GCC TTT AAT ACA GCT CAG TGC 1872 Asn Val Ala Tyr His Asn Trp Arg His Ala Phe Asn Thr Ala Gln Cys 610 615 ATG TTT GCT GCT CTA AAA GCA GGC AAA ATT CAG AAC AAG CTG ACT GAC 1920 Met Phe Ala Ala Leu Lys Ala Gly Lys Ile Gln Asn Lys Leu Thr Asp CTG GAG ATA CTT GCA TTG CTG ATT GCT GCA CTA AGC CAC GAT TTG GAT 1968 Leu Glu Ile Leu Ala Leu Leu Ile Ala Ala Leu Ser His Asp Leu Asp 645 650 2016 CAC CGT GGT GTG AAT AAC TCT TAC ATA CAG CGA AGT GAA CAT CCA CTT His Arg Gly Val Asn Asn Ser Tyr Ile Gln Arg Ser Glu His Pro Leu 665 GCC CAG CTT TAC TGC CAT TCA ATC ATG GAA CAC CAT CAT TTT GAC CAG 2064 Ala Gln Leu Tyr Cys His Ser Ile Met Glu His His His Phe Asp Gln 680 TGC CTG ATG ATT CTT AAT AGT CCA GGC AAT CAG ATT CTC AGT GGC CTC 2112 Cys Leu Met Ile Leu Asn Ser Pro Gly Asn Gln Ile Leu Ser Gly Leu 690 695 TCC ATT GAA GAA TAT AAG ACC ACG TTG AAA ATA ATC AAG CAA GCT ATT 2160 Ser Ile Glu Glu Tyr Lys Thr Thr Leu Lys Ile Ile Lys Gln Ala Ile 710 715 TTA GCT ACA GAC CTA GCA CTG TAC ATT AAG AGG CGA GGA GAA TTT TTT 2208 Leu Ala Thr Asp Leu Ala Leu Tyr Ile Lys Arg Arg Gly Glu Phe Phe

730

Glu	CTT Leu	Ile	Arg 740	Lys	Asn	Gln	Phe	Asn 745	Leu	Glu	Asp	Pro	His 750	Gln	Lys	2256
	TTG Leu															2304
_	AAA Lys 770				_	_										2352
	TTT Phe															2400
	ACT Thr															2448
_	GTT Val	_	_	_		_										2496
	CAC His															2544
	AGG Arg 850															2592
_	AAT Asn	_	_								Trp					2640
	GAT Asp															2688
	GGG Gly															2736
	AAG Lys															2784
	CTG Leu 930															2832
	CCC Pro			_	_											2880

945					950					955					960	
														GCC Ala 975		2928
														GAC Asp		2976
						Glu					Gly			CTG Leu		3024
Asn		ATC			Lys					Lys				AAC Asn		3072
				Leu					Asn					TAT Tyr		3120
			Lys					Ile					Lys	ATC Ile 1055		3168
		Ile					Val					His		CAG Gln		3216
	Thr					Gly					Pro			CAC His		3264
Leu					Ala					Pro				CGC		3312
				Leu					Ala					CTC		3360
			Leu	TAC Tyr 1125	AAG Lys	TAA										3381

## (2) INFORMATION FOR SEQ ID NO:145:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1126 amino acids

(B) TYPE: amino acid(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:145:

		_	_ •		_	_	_,				_				
1				5					Gln 10		_			15	
Pro	Gln	Gln	Gln 20	Lys	Gln	Gln	Gln	Arg 25	Asp	Gln	Asp	Ser	Val 30	Glu	Ala
Trp	Leu	Asp 35	Asp	His	Trp	Asp	Phe 40	Thr	Phe	Ser	Tyr	Phe 45	Val	Arg	Lys
Ala	Thr 50	Arg	Glu	Met	Val	Asn 55	Ala	Trp	Phe	Ala	Glu 60	Arg	Val	His	Thr
Ile 65	Pro	Val	Cys	Lys	Glu 70	Gly	Ile	Arg	Gly	His 75	Thr	Glu	Ser	Cys	Ser 80
Cys	Pro	Leu	Gln	Gln 85	Ser	Pro	Arg	Ala	Asp 90	Asn	Ser	Val	Pro	Gly 95	Thr
Pro	Thr	Arg	Lys 100	Ile	Ser	Ala	Ser	Glu 105	Phe	Asp	Arg	Pro	Leu 110	Arg	Pro
Ile	Val	Val 115	Lys	Asp	Ser	Glu	Gly 120	Thr	Val	Ser	Phe	Leu 125	Ser	Asp	Ser
Glu	Lys 130	Lys	Glu	Gln	Met	Pro 135	Leu	Thr	Pro	Pro	Arg 140	Phe	Asp	His	Asp
Glu 145	Gly	Asp	Gln	Cys	Ser 150	Arg	Leu	Leu	Glu	Leu 155	Val	Lys	Asp	Ile	Ser 160
Ser	His	Leu	Asp	Val 165	Thr	Ala	Leu	Cys	His 170	Lys	Ile	Phe	Leu	His 175	Ile
His	Gly	Leu	Ile 180	Ser	Ala	Asp	Arg	Tyr 185	Ser	Leu	Phe	Leu	Val 190	Cys	Glu
Asp	Ser	Ser 195	Asn	Asp	Lys	Phe	Leu 200	Ile	Ser	Arg	Leu	Phe 205	Asp	Val	Ala
Glu	Gly 210	Ser	Thr	Leu	Glu	Glu 215	Val	Ser	Asn	Asn	Cys 220	Ile	Arg	Leu	Glu
Trp 225	Asn	Lys	Gly	Ile	Val 230	Gly	His	Val	Ala	Ala 235	Leu	Gly	Glu	Pro	Leu 240
Asn	Ile	Lys	Asp	Ala 245	Tyr	Glu	Asp	Pro	Arg 250	Phe	Asn	Ala	Glu	Val 255	Asp
Gln	Ile	Thr	Gly 260	Tyr	Lys	Thr	Gln	Ser 265	Ile	Leu	Cys	Met	Pro 270	Ile	Lys
Asn	His	Arg 275	Glu	Glu	Val	Val	Gly 280	Val	Ala	Gln	Ala	Ile 285	Asn	Lys	Lys
Ser	Gly 290	Asn	Gly	Gly	Thr	Phe 295	Thr	Glu	Lys	Asp	Glu 300	Lys	Asp	Phe	Ala
Ala 305	Tyr	Leu	Ala	Phe	Cys 310	_	Ile	Val	Leu	His		Ala	Gln	Leu	Tyr 320
	Thr	Ser	Leu	Leu 325			Lys	Arg	Asn 330			Leu	Leu	Asp	Leu
Ala	Ser	Leu	Ile 340		Glu	Glu	Gln	Gln 345		Leu	Glu	Val	11e 350		Lys
Lys	Ile	Ala 355		Thr	Ile	Ile	Ser 360		Met	Gln	Val	Gln 365		Cys	Thr
Ile	Phe 370		Val	Asp	Glu	Asp 375		Ser	Asp	Ser	Phe 380		Ser	Val	Phe

His 385	Met	Glu	Cys	Glu	Glu 390	Leu	Glu	Lys	Ser	Ser 395	Asp	Thr	Leu	Thr	Arg 400
Glu	His	Asp	Ala	Asn 405	Lys	Ile	Asn	Tyr	Met 410	Tyr	Ala	Gln	Tyr	Val 415	Lys
Asn	Thr	Met	Glu 420	Pro	Leu	Asn	Ile	Pro 425	Asp	Val	Ser	Lys	Asp 430	Lys	Arg
Phe	Pro	Trp 435	Thr	Thr	Glu	Asn	Thr 440	Gly	Asn	Val	Asn	Gln 445	Gln	Cys	Ile
Arg	Ser 450	Leu	Leu	Cys	Thr	Pro 455	Ile	Lys	Asn	Gly	Lys 460	Lys	Asn	Lys	Val
Ile 465	Gly	Val	Cys	Gln	Leu 470	Val	Asn	Lys	Met	Glu 475	Glu	Asn	Thr	Gly	Lys 480
Val	Lys	Pro	Phe	Asn 485	Arg	Asn	Asp	Glu	Gln 490	Phe	Leu	Glu	Ala	Phe 495	Val
Ile	Phe	Cys	Gly 500	Leu	Gly	Ile	Gln	Asn 505	Thr	Gln	Met	Tyr	Glu 510	Ala	Val
Glu	Arg	Ala 515	Met	Ala	Lys	Gln	Met 520	Val	Thr	Leu	Glu	Val 525	Leu	Ser	Tyr
His	Ala 530	Ser	Ala	Ala	Glu	Glu 535	Glu	Thr	Arg	Glu	Leu 540	Gln	Ser	Leu	Ala
Ala 545	Ala	Val	Val	Pro	Ser 550	Ala	Gln	Thr	Leu	Lys 555	Ile	Thr	Asp	Phe	Ser 560
Phe	Ser	Asp	Phe	Glu 565	Leu	Ser	Asp	Leu	Glu 570	Thr	Ala	Leu	Суѕ	Thr 575	Ile
Arg	Met	Phe	Thr 580	Asp	Leu	Asn	Leu	Val 585	Gln	Asn	Phe	Gln	Met 590	Lys	His
Glu	Val	Leu 595	Cys	Arg	Trp	Ile	Leu 600	Ser	Val	Lys	Lys	Asn 605	Tyr	Arg	Lys
Asn	Val 610	Ala	Tyr	His	Asn	Trp 615	Arg	His	Ala	Phe	Asn 620	Thr	Ala	Gln	Cys
Met 625	Phe	Ala	Ala	Leu	Lys 630		Gly	Lys	Ile	Gln 635	Asn	Lys	Leu	Thr	Asp 640
Leu	Glu	Ile	Leu	Ala 645	Leu	Leu	Ile	Ala	Ala 650	Leu	Ser	His	Asp	Leu 655	Asp
His	Arg	Gly	Val 660	Asn	Asn	Ser	Tyr	Ile 665	Gln	Arg	Ser	Glu	His 670	Pro	Leu
Ala	Gln	Leu 675	Tyr	Cys	His	Ser	Ile 680	Met	Glu	His	His	His 685	Phe	Asp	Gln
_	690					695		_	Asn		700				
Ser 705	Ile	Glu	Glu	Tyr	Lys 710	Thr	Thr	Leu	Lys	Ile 715	Ile	Lys	Gln	Ala	Ile 720
Leu	Ala	Thr	Asp	Leu 725	Ala	Leu	Tyr	Ile	Lys 730	Arg	Arg	Gly	Glu	Phe 735	Phe
Glu	Leu	Ile	Arg 740	Lys	Asn	Gln	Phe	Asn 745	Leu	Glu	Asp	Pro	His 750	Gln	Lys
Glu	Leu	Phe 755	Leu	Ala	Met	Leu	Met 760	Thr	Ala	Суѕ	Asp	Leu 765	Ser	Ala	Ile
Thr	Lys 770	Pro	Trp	Pro	Ile	Gln 775	Gln	Arg	Ile	Ala	Glu 780	Leu	Val	Ala	Thr
Glu 785	Phe	Phe	Asp	Gln	Gly 790	Asp	Arg	Glu	Arg	Lys 795	Glu	Leu	Asn	Ile	Glu 800
Pro	Thr	Asp	Leu	Met 805	Asn	Arg	Glu	Lys	Lys 810	Asn	Lys	Ile	Pro	Ser 815	Met

Gln Val Gly Phe Ile Asp Ala Ile Cys Leu Gln Leu Tyr Glu Ala Leu 825 Thr His Val Ser Glu Asp Cys Phe Pro Leu Leu Asp Gly Cys Arg Lys 845 840 835 Asn Arg Gln Lys Trp Gln Ala Leu Ala Glu Gln Gln Glu Lys Met Leu 855 Ile Asn Gly Glu Ser Gly Gln Ala Lys Arg Asn Trp Val Pro Arg Ala 865 870 875 Arg Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe 885 890 Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly 900 905 His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly 915 920 925 Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro 940 930 935 Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser 945 950 955 Arg Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met 965 970 Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly 980 985 990 Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val 1000 1005 Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile 1010 1015 1020 Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile 1030 1035 Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg 1045 1050 1055 His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln 1060 1065 1070 Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr 1075 1080 1085 Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp 1090 1095 1100 His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly 105 1110 1115 Met Asp Glu Leu Tyr Lys 1125

#### (2) INFORMATION FOR SEQ ID NO:146:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2760 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: Coding Sequence
  - (B) LOCATION: 1...2757

### (D) OTHER INFORMATION:

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:146:

ATG Met .								48
ACC Thr								96
GAG (								144
TTC Phe								192
TTC Phe 65								240
TTT Phe								288
GAC Asp								336
ACG Thr								384
GGG Gly								432
ATG Met 145	 	 						480
CGC Arg								528
GTA .								576
GAT Asp								624

195	20	00	205
		ys Cys Ser Leu	AAC CCT GAG TGG AAT 672 Asn Pro Glu Trp Asn 220
			AAA GAC AGA AGA CTG 720 Lys Asp Arg Arg Leu 240
			AGG AAT GAC TTC ATG 768 Arg Asn Asp Phe Met 255.
_			AAG GCC AGT GTT GAT 816 Lys Ala Ser Val Asp 270
	Leu Leu Ser G		GAG TAC TTC AAT GTG 864 Glu Tyr Phe Asn Val 285
_		lu Ala Asn Glu	GAA CTG CGG CAG AAA 912 Glu Leu Arg Gln Lys 300
			GTC CCG GAA GAA AAG 960 Val Pro Glu Glu Lys 320
			GGC AAC AGA GAC CGG 1008 Gly Asn Arg Asp Arg 335
			CTG GGG AAA GGC AGC 1056 Leu Gly Lys Gly Ser 350
	Met Leu Ser G		ACA GAT GAG CTC TAT 1104 Thr Asp Glu Leu Tyr 365
		sp Val Val Ile	CAA GAT GAT GAC GTG 1152 Gln Asp Asp Asp Val 380
			CTG CCT GGG AAG CCG 1200 Leu Pro Gly Lys Pro 400
			ACC ATG GAC CGC CTG 1248 Thr Met Asp Arg Leu 415

									GGC Gly							1296
									CAT His							1344
_						Phe			CAG Gln							1392
									CTC Leu							1440
									GAA Glu 490							1488
									GAC Asp							1536
									GTG Val							1584
									CAG Gln							1632
									ATG Met							1680
Pro	Lys	Ser	Met	Ser 565	Lys	Glu	Ala	Val	GCC Ala 570	Ile	Cys	Lys	Gly	Leu 575	Met	1728
Thr	Lys	His	Pro 580	Gly	Lys	Arg	Leu	Gly 585	Cys	Gly	Pro	Glu	Gly 590	Glu		1776
Asp	Ile	Lys 595	Glu	His	Ala	Phe	Phe 600	Arg	Tyr	Ile	Asp	Trp 605	Glu	Lys	Leu	1824
Glu	Arg 610	Lys	Glu	Ile	Gln	Pro 615	Pro	Tyr	Lys	Pro	Lys 620	Ala	Cys	Gly	CGA Arg	1872
															CTA Leu	1920

625	630	635	640
<del>-</del>	n Glu Val Ile Arg	AAT ATT GAC CAA TCA GA Asn Ile Asp Gln Ser Gl 650 65	u Phe
		TTT TTA AAA CCC GAA GT Phe Leu Lys Pro Glu Va 670	
		GTG AGC AAG GGC GAG GA Val Ser Lys Gly Glu G 685	
		GAG CTG GAC GGC GAC G Glu Leu Asp Gly Asp V 700	
		GGC GAG GGC GAT GCC AGGly Glu Gly Asp Ala TO	
	u Lys Phe Ile Cys	ACC ACC GGC AAG CTG C Thr Thr Gly Lys Leu P 730 7	
		ACC TAC GGC GTG CAG T Thr Tyr Gly Val Gln C 750	
		CAC GAC TIC TTC AAG T His Asp Phe Phe Lys S 765	
		ACC ATC TTC TTC AAG G Thr Ile Phe Phe Lys A 780	
		AAG TTC GAG GGC GAC A Lys Phe Glu Gly Asp T 795	
	u Leu Lys Gly Ile	GAC TTC AAG GAG GAC G Asp Phe Lys Glu Asp G 810	
		TAC AAC AGC CAC AAC G Tyr Asn Ser His Asn V 830	
		ATC AAG GTG AAC TTC A Ile Lys Val Asn Phe I 845	

CGC	CAC	AAC	ATC	GAG	GAC	GGC	AGC	GTG	CAG	CTC	GCC	GAC	CAC	TAC	CAG	2592
Arg	His	Asn	Ile	Glu	Asp	Gly	Ser	Val	Gln	Leu	Ala	Asp	His	Tyr	Gln	
	850					855					860					
CAG	AAC	ACC	CCC	ATC	GGC	GAC	GGC	CCC	GTG	CTG	CTG	CCC	GAC	AAC	CAC	2640
Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	Pro	Asp	Asn	His	
865					870					875					880	
TAC	CTG	AGC	ACC	CAG	TCC	GCC	CTG	AGC	AAA	GAC	CCC	AAC	GAG	AAG	CGC	2688
Tyr	Leu	Ser	Thr	Gln	Ser	Ala	Leu	Ser	Lys	Asp	Pro	Asn	Glu	Lys	Arg	
				885					890					895		
GAT	CAC	ATG	GTC	CTG	CTG	GAG	TTC	GTG	ACC	GCC	GCC	GGG	ATC	ACT	CTC	2736
Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	Ala	Ala	Gly	Ile	Thr	Leu	
			900					905					910			
GGC	ATG	GAC	GAG	CTG	TAC	AAG	TAA									2760
Gly	Met	Asp	Glu	Leu	Tyr	Lys										
		915														

#### (2) INFORMATION FOR SEQ ID NO:147:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 919 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:147:

Met Ala Asp Pro Ala Ala Gly Pro Pro Pro Ser Glu Gly Glu Glu Ser 5 10 Thr Val Arg Phe Ala Arg Lys Gly Ala Leu Arg Gln Lys Asn Val His 25 Glu Val Lys Asn His Lys Phe Thr Ala Arg Phe Phe Lys Gln Pro Thr 45 40 Phe Cys Ser His Cys Thr Asp Phe Ile Trp Gly Phe Gly Lys Gln Gly 55 Phe Gln Cys Gln Val Cys Cys Phe Val Val His Lys Arg Cys His Glu 70 Phe Val Thr Phe Ser Cys Pro Gly Ala Asp Lys Gly Pro Ala Ser Asp 85 90 Asp Pro Arg Ser Lys His Lys Phe Lys Ile His Thr Tyr Ser Ser Pro 105 Thr Phe Cys Asp His Cys Gly Ser Leu Leu Tyr Gly Leu Ile His Gln 120 Gly Met Lys Cys Asp Thr Cys Met Met Asn Val His Lys Arg Cys Val 130 135 140 Met Asn Val Pro Ser Leu Cys Gly Thr Asp His Thr Glu Arg Arg Gly 155 150

Arg	Ile	Tyr	Ile	Gln 165	Ala	His	Ile	Asp	Arg 170	Asp	Val	Leu	Ile	Val 175	Leu
Val	Arg	Asp	Ala 180	Lys	Asn	Leu	Val	Pro 185	Met	Asp	Pro	Asn	Gly 190	Leu	Ser
Asp	Pro	Тут 195	Val	Lys	Leu	Lys	Leu 200	Ile	Pro	Asp	Pro	Lys 205	Ser	Glu	Ser
_	210	_		_		215	_	_			220			Trp	
225					230		_			235				Arg	240
				245					250					Phe 255	
_			260		_			265			_		270	Val	_
_		275	_				280					285		Asn	
	290				_	295					300			Gln	_
305		-		_	310			_		315				Glu	320
				325		_		_	330		-		_	335	_
	-		340	_				345				_	350	Gly	
	_	355					360			_		365		Leu	_
	370				_	375					380		_	Asp	
385	_				390	_				395			_	Lys	400
				405				-	410				_	Arg 415	
			420					425					430	His	
		435	_			_	440					445	_	Ala	
	450			-		455					460	_		Ile	
465					470					475				His	480
				485					490					495	Val
		_	500		_	_		505	_	_			510		Ile
		515			_	_	520			_	_	525			Gly
	530					535					540				Glu
Asp 545	Glu	Asp	Glu	Leu	Phe 550	Gln	Ser	Ile	Met	Glu 555	His	Asn	Val	Ala	Тут 560
	_			565	_				570		_	_		Leu 575	
Thr	Lys	His	Pro 580	Gly	Lys	Arg	Leu	Gly 585	Cys	Gly	Pro	Glu	Gly 590	Glu	Arg

Asp	Ile	Lys 595	Glu	His	Ala	Phe	Phe 600	Arg	Tyr	Ile	Asp	Trp 605	Glu	Lys	Leu
Glu	Arg 610	Lys	Glu	Ile	Gln	Pro 615	Pro	Tyr	Lys	Pro	Lys 620	Ala	Cys	Gly	Arg
Asn 625	Ala	Glu	Asn	Phe	Asp 630	Arg	Phe	Phe	Thr	Arg 635	His	Pro	Pro	Val	Leu 640
Thr	Pro	Pro	Asp	Gln 645	Glu	Val	Ile	Arg	Asn 650	Ile	Asp	Gln	Ser	Glu 655	Phe
Glu	Gly	Phe	Ser 660	Phe	Val	Asn	Ser	Glu 665	Phe	Leu	Lys	Pro	Glu 670	Val	Lys
Ser	Ser	Asp 675	Pro	Pro	Val	Ala	Thr 680	Met	Val	Ser	Lys	Gly 685	Glu	Glu	Leu
Phe	Thr 690	Gly	Val	Val	Pro	Ile 695	Leu	Val	Glu	Leu	Asp 700	Gly	Asp	Val	Asn
Gly 705	His	Lys	Phe	Ser	Val 710	Ser	Gly	Glu	Gly	Glu 715	Gly	Asp	Ala	Thr	Tyr 720
Gly	Lys	Leu	Thr	Leu 725	Lys	Phe	Ile	Cys	Thr 730	Thr	Gly	Lys	Leu	Pro 735	Val
Pro	Trp	Pro	Thr 740	Leu	Val	Thr	Thr	Leu 745	Thr	Tyr	Gly	Val	Gln 750	Cys	Phe
Ser	Arg	Tyr 755	Pro	Asp	His	Met	Lys 760	Gln	His	Asp	Phe	Phe 765	Lys	Ser	Ala
Met	Pro 770	Glu	Gly	Tyr	Val	Gln 775	Glu	Arg	Thr	Ile	Phe 780	Phe	Lys	Asp	Asp
Gly 785	Asn	Tyr	Lys	Thr	Arg 790	Ala	Glu	Val	Lys	Phe 795	Glu	Gly	Asp	Thr	Leu 800
Val	Asn	Arg	Ile	Glu 805	Leu	Lys	Gly	Ile	Asp 810	Phe	Lys	Glu	Asp	Gly 815	Asn
Ile	Leu	Gly	His 820	Lys	Leu	Glu	Tyr	Asn 825	Tyr	Asn	Ser	His	Asn 830	Val	Tyr
Ile	Met	Ala 835	Asp	Lys	Gln	Lys	Asn 840	Gly	Ile	Lys	Val	Asn 845	Phe	Lys	Ile
Arg	His 850	Asn	Ile	Glu	Asp	Gly 855	Ser	Val	Gln	Leu	Ala 860	Asp	His	Tyr	Gln
Gln 865	Asn	Thr	Pro	Ile	Gly 870	Asp	Gly	Pro	Val	Leu 875	Leu	Pro	Asp	Asn	His 880
Tyr	Leu	Ser	Thr	Gln 885	Ser	Ala	Leu	Ser	Lys 890	Asp	Pro	Asn	Glu	Lys 895	Arg
Asp	His	Met	Val 900	Leu	Leu	Glu	Phe	Val 905	Thr	Ala	Ala	Gly	Ile 910	Thr	Leu
Gly	Met	Asp		Leu	Tyr	Lys									

## (2) INFORMATION FOR SEQ ID NO:148:

### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3009 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 1...3006(D) OTHER INFORMATION:

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:148:

ATG	GCT	CAG	CAG	ACA	AGC	CCG	GAC	ACT	TTA	ACA	GTA	ССТ	GAA	GTG	GAT	48
Met 1	Ala	Gln	Gln	Thr 5	Ser	Pro	Asp	Thr	Leu 10	Thr	Val	Pro	Glu	Val 15	Asp	
				CCA						_			_			96
ASN	Pro	HIS	cys 20	Pro	ASN	Pro	Trp	25	Asn	GIU	Asp	Leu	30	ьуs	ser	
				CTG Leu												144
Leu	Arg	35	ASII	neu	neu	GIII	40	Giu	гу	Ser	гуs	45	Ala	Arg	БУЗ	
				AAG Lys												192
ser	50	261	PIO	Буз	Deu	55	PIO	Vai	116	Ser	60	Arg	ASII	Ser	PIO	
				AGA												240
65	Leu .	Leu	Arg	Arg	70	Leu	Leu	ser	ser	75	116	PIO	ьуѕ	GIN	80 80	
				GCA Ala												288
Arg	rne	1111	Vai	85	nis	1111	Cys.	riie	90	vai	Asp	ASII	GIY	95	Ser	
				CCC Pro										_		336
Ala	GIY	Arg	100	FIO	ъец	чэр	PIO	105	1111	361	PIO	GLY	110	Gly	Dea	
				AAT Asn												384
110	Deu	115			1	Vul	120	DCI	0211	9	9	125	Der		Dea	
				AGC Ser												432
ıyı	130	Ser	nsp	Der	nsp	135	лэр	Deu	Jei	rio	140	Der	nec	Ser		
				GCC Ala												480
145	Ser	Jer	110	niu	150	പാറ്റ	110	1113	Gry	155	Asp	Dea	110	vai	160	
				GTC Val												528
FIO	THE	and.	3111	165	nea.	ALG	DEL	<u> Leu</u>	170	****	VUI	an g	Aaii	175		
				AAT		_			_							576
ATS	ATG	ьeu	180	Asn	ьeu	GIN	ASP	Arg 185	AIG	PTO	ser	гуз	Arg 190	ser	PIO	

ATG TGC Met Cys									624
TAC CAG Tyr Gln 210	Lys								672
GAC CAG Asp Gln 225									720
TCC AAC Ser Asn							_		768
GAA ATG Glu Met									816
TTC TTA									864
GAA AAG Glu Lys 290	Glu								912
AAA TTG Lys Leu 305						_		_	960
GGA GTT Gly Val									1008
GTG AAC Val Asn						Glu			1056
AAC CGG Asn Arg									1104
TTA TTA Leu Leu 370	Lys								1152
ATG ACT Met Thr 385									1200
ATC CAT									1248

CCT GCT TTG GAG GCT GTG TTT ACA GAT TTG GAG ATT CTT GCA GCA ATT Pro Ala Leu Glu Ala Val Phe Thr Asp Leu Glu Ile Leu Ala Ala Ile TTT GCC AGT GCA ATA CAT GAT GTA GAT CAT CCT GGT GTG TCC AAT CAA Phe Ala Ser Ala Ile His Asp Val Asp His Pro Gly Val Ser Asn Gln TTT CTG ATC AAT ACA AAC TCT GAA CTT GCC TTG ATG TAC AAT GAT TCC Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met Tyr Asn Asp Ser TCA GTC TTA GAG AAC CAT CAT TTG GCT GTG GGC TTT AAA TTG CTT CAG Ser Val Leu Glu Asn His His Leu Ala Val Gly Phe Lys Leu Leu Gln GAA GAA AAC TGT GAC ATT TTC CAG AAT TTG ACC AAA AAA CAA AGA CAA Glu Glu Asn Cys Asp Ile Phe Gln Asn Leu Thr Lys Lys Gln Arg Gln TCT TTA AGG AAA ATG GTC ATT GAC ATC GTA CTT GCA ACA GAT ATG TCA Ser Leu Arg Lys Met Val Ile Asp Ile Val Leu Ala Thr Asp Met Ser AAA CAC ATG AAT CTA CTG GCT GAT TTG AAG ACT ATG GTT GAA ACT AAG Lys His Met Asn Leu Leu Ala Asp Leu Lys Thr Met Val Glu Thr Lys AAA GTG ACA AGC TCT GGA GTT CTT CTT GAT AAT TAT TCC GAT AGG Lys Val Thr Ser Ser Gly Val Leu Leu Leu Asp Asn Tyr Ser Asp Arg ATT CAG GTT CTT CAG AAT ATG GTG CAC TGT GCA GAT CTG AGC AAC CCA Ile Gln Val Leu Gln Asn Met Val His Cys Ala Asp Leu Ser Asn Pro ACA AAG CCT CTC CAG CTG TAC CGC CAG TGG ACG GAC CGG ATA ATG GAG Thr Lys Pro Leu Gln Leu Tyr Arg Gln Trp Thr Asp Arg Ile Met Glu GAG TTC TTC CGC CAA GGA GAC CGA GAG AGG GAA CGT GGC ATG GAG ATA Glu Phe Phe Arg Gln Gly Asp Arg Glu Arg Glu Arg Gly Met Glu Ile AGC CCC ATG TGT GAC AAG CAC AAT GCT TCC GTG GAA AAA TCA CAG GTG Ser Pro Met Cys Asp Lys His Asn Ala Ser Val Glu Lys Ser Gln Val GGC TTC ATA GAC TAT ATT GTT CAT CCC CTC TGG GAG ACA TGG GCA GAC Gly Phe Ile Asp Tyr Ile Val His Pro Leu Trp Glu Thr Trp Ala Asp

	CTC	GTC	CAC	ССТ	GAC	GCC	CAG	GAT	TTA	TTG	GAC	ACT	TTG	GAG	GAC	AAT	1920	
	Leu	Val	His	Pro	Asp	Ala	Gln	Asp	Ile	Leu	Asp	Thr	Leu	Glu	Asp	Asn		
	625					630					635					640		
		GAA													_		1968	
	Arg	Glu	Trp	Tyr		Ser	Thr	TTE	Pro		Ser	Pro	Ser	Pro		Pro		
					645					650					655			
	СУТ	GAC	CCA	GAG	GAG	GGC	CGG	CAG	GGT	CAA	ΔСΤ	GAG	ΑΔΑ	ጉጥ	CAG	ጥጥተ	2016	
		Asp															2010	
				660			5		665					670				
	GAA	CTA	ACT	TTA	GAG	GAA	GAT	GGT	GAG	TCA	GAC	ACG	GAA	AAG	GAC	AGT	2064	
	Glu	Leu	Thr	Leu	Glu	Glu	Asp	Gly	Glu	Ser	Asp	Thr	Glu	Lys	Asp	Ser		
			675					680					685					
																	0110	
		AGT	_														2112	
	GIĀ	Ser 690	GIN	vai	GIU	GIU	ASP 695	Thr	ser	Cys	Ser	700	Ser	rys	тиц	Leu		
		090					093					,00						
	TGT	ACT	CAA	GAC	TCA	GAG	TCT	ACT	GAA	АТТ	ccc	CTT	GAT	GAA	CAG	GTT	2160	
		Thr																
	705					710					715					720		
	GAA	GAG	GAG	GCA	GTA	GGG	GAA	GAA	GAG	GAA	AGC	CAG	CCT	GAA	GCC	TGT	2208	
	Glu	Glu	Glu	Ala		Gly	Glu	Glu	Glu		Ser	Gln	Pro	Glu		Cys		
					725					730					735			
	CITIC	ATA	CAM	Cam	CCT	m~m	CCT	CAC	NCC.	NCC.	CCN	אינוייע	CTC	CAG	TYCC	ACG	2256	
		Ile															2230	
	Val	110	ımp	740	9				745		0_3			750				
	GTA	CCG	CGG	GCC	CGG	GAT	CCA	CCG	GTC	GCC	ACC	ATG	GTG	AGC	AAG	GGC	2304	
	Val	Pro	Arg	Ala	Arg	Asp	Pro	Pro	Val	Ala	Thr	Met	Val	Ser	Lys	Gly		
			755					760					765					
														~~~			5350	
		GAG										_	-			_	2352	
•	GIU	Glu 770	Leu	РПЕ	THE	GIY	775	vaı	PLO	116	Leu	780	GIU	Leu	ASD	GIY		
		770					,,,					, 00						
	GAC	GTA	AAC	GGC	CAC	AAG	TTC	AGC	GTG	TCC	GGC	GAG	GGC	GAG	GGC	GAT	2400	
		Val																
	785					790					795					800		
		ACC															2448	
	Ala	Thr	Tyr	Gly		Leu	Thr	Leu	Lys		Ile	Cys	Thr	Thr		Lys		
					805					810					815			
	Cuv	CCC	CTY	CCC	ייבאנו	CCC	ΔCC	ריזירי -	Cuta	ACC	ACC	ביזים	ACC	ΤΔС	GGC	GTG	2496	
		Pro													_	_		
				820		_ •		🕶	825			==	_	830				
		TGC															2544	
	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	His	Asp	Phe	Phe		

845

AAG TCC GCC ATG CCC GAA GGC TAC GTC CAG GAG CGC ACC ATC TTC TTC 2592 Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe 855 AAG GAC GAC GGC AAC TAC AAG ACC CGC GCC GAG GTG AAG TTC GAG GGC 2640 Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly 870 GAC ACC CTG GTG AAC CGC ATC GAG CTG AAG GGC ATC GAC TTC AAG GAG 2688 Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu 885 GAC GGC AAC ATC CTG GGG CAC AAG CTG GAG TAC AAC TAC AAC AGC CAC 2736 Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His 900 905 AAC GTC TAT ATC ATG GCC GAC AAG CAG AAG AAC GGC ATC AAG GTG AAC 2784 Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn 915 920 TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC GTG CAG CTC GCC GAC 2832 Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp 935 CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC CCC GTG CTG CTC His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro 950 955 GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG AGC AAA GAC CCC AAC 2928 Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn 965 970 GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC GTG ACC GCC GGG 2976 Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly 980 985 990 ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA 3009 Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 995 1000

840

#### (2) INFORMATION FOR SEQ ID NO:149:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1002 amino acids

(B) TYPE: amino acid

835

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein
(v) FRAGMENT TYPE: internal

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:149:

		<b>~1</b>	<b>01</b> -	ml	G	D	3	mh an	T	mh.ss	1.7. T	Dwa	C1	1/07	7
Met 1	Ala	GIn	Gin	1nr 5	ser	Pro	Asp	Thr	Leu 10	THE	Vai	PIO	GIU	15	Asp
Asn	Pro	His	Cys 20	Pro	Asn	Pro	Trp	Leu 25	Asn	Glu	Asp	Leu	Val 30	Lys	Ser
Leu	Arg	Glu 35	Asn	Leu	Leu	Gln	His 40	Glu	Lys	Ser	Lys	Thr 45	Ala	Arg	Lys
Ser	Val 50	Ser	Pro	Lys	Leu	Ser 55	Pro	Val	Ile	Ser	Pro 60	Arg	Asn	Ser	Pro
Arg 65	Leu	Leu	Arg	Arg	Met 70	Leu	Leu	Ser	Ser	Asn 75	Ile	Pro	Lys	Gln	Arg 80
Arg	Phe	Thr	Val	<b>Al</b> a 85	His	Thr	Cys	Phe	Asp 90	Val	Asp	Asn	Gly	Thr 95	Ser
Ala	Gly	Arg	Ser 100	Pro	Leu	Asp	Pro	Met 105	Thr	Ser	Pro	Gly	Ser 110	Gly	Leu
Ile	Leu	Gln 115	Ala	Asn	Phe	Val	His 120	Ser	Gln	Arg	Arg	Glu 125	Ser	Phe	Leu
Tyr	Arg 130	Ser	Asp	Ser	Asp	Tyr 135	Asp	Leu	Ser	Pro	Lys 140	Ser	Met	Ser	Arg
Asn	Ser	Ser	Ile	Ala	Ser	Asp	Ile	His	Gly	Asp	Asp	Leu	Ile	Val	Thr
145					150					155					160
Pro	Phe	Ala	Gln	Val 165	Leu	Ala	Ser	Leu	Arg 170	Thr	Val	Arg	Asn	Asn 175	Phe
Ala	Ala	Leu	Thr 180	Asn	Leu	Gln	Asp	Arg 185	Ala	Pro	Ser	Lys	Arg 190	Ser	Pro
	-	195					200		Ala			205			
-	210					215			Glu		220				
Asp 225	Gln	Leu	Glu	Thr	Leu 230	Gln	Thr	Arg	His	Ser 235	Val	Ser	Glu	Met	Ala 240
	Asn	Lys	Phe	Lys 245		Met	Leu	Asn	Arg 250	Glu	Leu	Thr	His	Leu 255	Ser
Glu	Met	Ser	Arg 260	Ser	Gly	Asn	Gln	Val 265	Ser	Glu	Phe	Ile	Ser 270	Asn	Thr
Phe	Leu	Asp 275	Lys	Gln	His	Glu	Val 280	Glu	Ile	Pro	Ser	Pro 285	Thr	Gln	Lys
Glu	Lys 290		Lys	Lys	Lys	Arg 295		Met	Ser	Gln	Ile 300	Ser	Gly	Val	Lys
Lvs		Met	His	Ser	Ser		Leu	Thr	Asn	Ser		Ile	Pro	Ara	Phe
305					310					315				_	320
	Val	Lys	Thr	Glu 325		Glu	Asp	Val	Leu 330	Ala	Lys	Glu	Leu	Glu 335	Asp
Val	Asn	Lys	Trp 340	Gly	Leu	His	Val	Phe	Arg	Ile	Ala	Glu	Leu 350	Ser	Gly
Asn	Arg	Pro 355	Leu	Thr	Val	Ile	Met 360	His	Thr	Ile	Phe	Gln 365	Glu	Arg	Asp
Leu	Leu 370		Thr	Phe	Lys	Ile 375		Val	Asp	Thr	Leu 380	Ile	Thr	Tyr	Leu
Met 385		Leu	Glu	Asp	His 390		His	Ala	Asp	Val 395	Ala	Tyr	His	Asn	Asn 400
	His	Ala	Ala	Asp 405		Val	Gln	Ser	Thr 410		Val	Leu	Leu	Ser 415	

Pro	Ala	Leu	Glu 420	Ala	Val	Phe	Thr	Asp 425	Leu	Glu	Ile	Leu	Ala 430	Ala	Ile
Phe	Ala	Ser 435	Ala	Ile	His	Asp	Val 440	Asp	His	Pro	Gly	Val 445	Ser	Asn	Gln
Phe	Leu 450	Ile	Asn	Thr	Asn	Ser 455	Glu	Leu	Ala	Leu	Met 460	Tyr	Asn	Asp	Ser
465					470				Val	475		_			480
			-	485					Leu 490		-	-		495	
			500				_	505	Val				510		
-		515					520		Lys			525			
_	530				-	535			Leu		540				
545	<b>01</b>	vui	Deu	02	550	1100	vui		Cys	555	າພຸ	<b>D</b> C <b>u</b>	501		560
	-			565		-			Trp 570		_			575	
			580					585	Arg				590		
		595			-		600		Ser			605			
_	610		_	_		615			Leu	_	620				
625	Val	nis	PIO	ASD	630	GIII	Asp	rre	Leu	635	TILL	Leu	GIU	Asp	640
Arg	Glu	Trp	Tyr	Gln 645	Ser	Thr	Ile	Pro	Gln 650	Ser	Pro	Ser	Pro	Ala 655	Pro
_	-		660		_			665	Gln				670		
		675				_	680		Ser			685			
_	690					695			Cys		700	•			
705			_		710				Glu	715					720
				725					730					735	
vaı	11e	Asp	740	Arg	ser	PIO	Asp	745	Thr	GIY	11e	Leu	750	ser	THE
		755					760		Ala			765			
	770				_	775			Ile		780				
Asp 785	Val	Asn	GIY	His	Lys 790	Phe	Ser	Val	Ser	795	Glu	GIY	GIu	GIA	Asp 800
Ala	Thr	Tyr	Gly	Lys 805	Leu	Thr	Leu	Lys	Phe 810	Ile	Cys	Thr	Thr	Gly 815	Lys
			820					825	Thr				830		
Gln	Cys	Phe 835	Ser	Arg	Tyr	Pro	Asp 840	His	Met	Lys	Gln	His 845	Asp	Phe	Phe

•

rys	850	Ala	met	Pro	GIU	855	ıyr	vai	GIN	GIU	860	THE	TTE	Pne	Pne		
Lys		Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu	Gly		
865					870					875					880		
Asp	Thr	Leu	Val	Asn 885	Arg	Ile	Glu	Leu	Lys 890	Gly	Ile	Asp	Phe	Lys 895	Glu		
Asp	Gly	Asn	Ile 900	Leu	Gly	His	Lys	Leu 905	Glu	Tyr	Asn	Tyr	Asn 910	Ser	His		
Asn	Val	Tyr 915	Ile	Met	Ala	Asp	Lys 920	Gln	Lys	Asn	Gly	Ile 925	Lys	Val	Asn		
Phe	Lys 930	Ile	Arg	His	Asn	Ile 935	Glu	Asp	Gly	Ser	Val 940	Gln	Leu	Ala	Asp		
His		Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	Leu	Leu	Pro		
945					950					955					960		
Asp	Asn	His	Tyr	Leu 965	Ser	Thr	Gln	Ser	Ala 970	Leu	Ser	Lys	Asp	Pro 975	Asn		
Glu	Lys	Arg	Asp 980	His	Met	Val	Leu	Leu 985	Glu	Phe	Val	Thr	Ala 990	Ala	Gly		
Ile	Thr	Leu 995	Gly	Met	Asp		Leu 1000	Tyr	Lys								
		(2	) INI	FORM	ATIOI	v FOI	R SE	Q ID	NO:	150:							
	(:	•					RIST										
							se pa	airs									
					ucle:			_									
					DNES: Y: 1:		ingl r	е									
		(D)	1010		1. 1.	IIICa.	<b>L</b>										
	(:	ii) 1	MOLE	TULE	TYP	E: c	DNA										
	(:	ix)	FEAT	JRE:													
		/ A	ו או	ME/K	rv. (	odi:	ng S	ലെട്	nce							•	
					ON:		_	eque	iice								
					INFO												
	(:	xi)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	150:						
ATG	GAG	GCA	GAG	GGC	AGC	AGC	GCG	CCG	GCC	CGG	GCG	GGC	AGC	GGA	GAG		48
Met	Glu	Ala	Glu	Gly	Ser	Ser	Ala	Pro	Ala	Arg	Ala	Gly	Ser	Gly	Glu		
1				5					10					15			
GGC	AGC	GAC	AGC	GCC	GGC	GGG	GCC	ACG	CTC	AAA	GCC	CCC	AAG	CAT	CTC		96
Gly	Ser	Asp	Ser	Ala	Gly	Gly	Ala	Thr	Leu	Lys	Ala	Pro	Lys	His	Leu		
			20					25					30				
TGG	AGG	CAC	GAG	CAG	CAC	CAC	CAG	TAC	CCG	CTC	CGG	CAG	CCC	CAG	TTC	1	144
Trp	Arg	His	Glu	Gln	His	His	Gln	Tyr	Pro	Leu	Arg		Pro	Gln	Phe		
		35					40					45					
CGC	CTC	CTG	CAT	CCC	CAT	CAC	CAC	CTG	CCC	CCG	CCG	CCG	CCA	CCC	TCG	1	192
Arg	Leu	Leu	His	Pro	His	His	His	Leu	Pro	Pro		Pro	Pro	Pro	Ser		
	50					55					60						

CCC CAG CCC Pro Gln Pro 65						240
CTG CCG CCG Leu Pro Pro						288
TCG AGC GGG Ser Ser Gly			g His Arg		Asp Thr	336
GAG CGC TAC Glu Arg Tyr 115						384
GAG ACC GGC Glu Thr Gly						432
TCC TCG TTC Ser Ser Phe						480
GCG GGA CGG . Ala Gly Arg						528
ATT CTC CAA			r Gln Arg		Phe Leu	576
TAT CGA TCC Tyr Arg Ser .						624
AAC TCC TCC Asn Ser Ser 210	_					672
CCA TTT GCT (Pro Phe Ala (225	Gln Val Leu 230	Ala Ser Le	Arg Thr 235	Val Arg Asn	Asn Phe 240	720
GCT GCA TTA	Thr Asn Leu 245	Gln Asp Arg	g Ala Pro 250	Ser Lys Arg	Ser Pro 255	768
ATG TGC AAC ( Met Cys Asn (	_	_	Ala Thr		Glu Ala	816
TAC CAG AAA ( Tyr Gln Lys I	_					864

GAC CAG CTA GAG ACC CTA CAG ACC AGG CAC TCC GTC AGT GAG ATG GCC Asp Gln Leu Glu Thr Leu Gln Thr Arg His Ser Val Ser Glu Met Ala TCC AAC AAG TTT AAA AGG ATG CTT AAT CGG GAG CTC ACC CAT CTC TCT Ser Asn Lys Phe Lys Arg Met Leu Asn Arg Glu Leu Thr His Leu Ser GAA ATG AGT CGG TCT GGA AAT CAA GTG TCA GAG TTT ATA TCA AAC ACA Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Phe Ile Ser Asn Thr TTC TTA GAT AAG CAA CAT GAA GTG GAA ATT CCT TCT CCA ACT CAG AAG Phe Leu Asp Lys Gln His Glu Val Glu Ile Pro Ser Pro Thr Gln Lys GAA AAG GAG AAA AAG AAA AGA CCA ATG TCT CAG ATC AGT GGA GTC AAG Glu Lys Glu Lys Lys Lys Arg Pro Met Ser Gln Ile Ser Gly Val Lys AAA TTG ATG CAC AGC TCT AGT CTG ACT AAT TCA AGT ATC CCA AGG TTT Lys Leu Met His Ser Ser Ser Leu Thr Asn Ser Ser Ile Pro Arg Phe GGA GTT AAA ACT GAA CAA GAA GAT GTC CTT GCC AAG GAA CTA GAA GAT Gly Val Lys Thr Glu Gln Glu Asp Val Leu Ala Lys Glu Leu Glu Asp GTG AAC AAA TGG GGT CTT CAT GTT TTC AGA ATA GCA GAG TTG TCT GGT Val Asn Lys Trp Gly Leu His Val Phe Arg Ile Ala Glu Leu Ser Gly AAC CGG CCC TTG ACT GTT ATC ATG CAC ACC ATT TTT CAG GAA CGG GAT Asn Arg Pro Leu Thr Val Ile Met His Thr Ile Phe Gln Glu Arg Asp TTA TTA AAA ACA TTT AAA ATT CCA GTA GAT ACT TTA ATT ACA TAT CTT Leu Leu Lys Thr Phe Lys Ile Pro Val Asp Thr Leu Ile Thr Tyr Leu ATG ACT CTC GAA GAC CAT TAC CAT GCT GAT GTG GCC TAT CAC AAC AAT Met Thr Leu Glu Asp His Tyr His Ala Asp Val Ala Tyr His Asn Asn ATC CAT GCT GCA GAT GTT GTC CAG TCT ACT CAT GTG CTA TTA TCT ACA Ile His Ala Ala Asp Val Val Gln Ser Thr His Val Leu Leu Ser Thr CCT GCT TTG GAG GCT GTG TTT ACA GAT TTG GAG ATT CTT GCA GCA ATT Pro Ala Leu Glu Ala Val Phe Thr Asp Leu Glu Ile Leu Ala Ala Ile

		CAT His									1536
 		 AAC Asn				 	 				1584
	_	CAT His	_							_	1632
		ATT Ile 550	_	_				_		_	1680
		GTC Val									1728
 		CTG Leu			_						1776
		GGA Gly									1824
		AAT Asn		_							1872
		CTG Leu 630						_		_	1920
 	 	 GGA Gly				 -	 				1968
		AAG Lys									2016
		ATT Ile								GAC Asp	2064
										AAT Asn	2112
									_	CCT Pro	2160

705	710	715	720
GAT GAC CCA GAG G Asp Asp Pro Glu G 7			
		GAG TCA GAC ACG ( Glu Ser Asp Thr ( 745	
		AGC TGC AGT GAC T Ser Cys Ser Asp S	
		GAA ATT CCC CTT ( Glu Ile Pro Leu A 780	
		GAG GAA AGC CAG ( Glu Glu Ser Gln 1 795	
Val Ile Asp Asp A		ACG ACG GGA ATT 0 Thr Thr Gly Ile 1 810	
		GTC GCC ACC ATG (Val Ala Thr Met V825	
		CCC ATC CTG GTC Pro Ile Leu Val	
		GTG TCC GGC GAG (Val Ser Gly Glu 6	
		AAG TTC ATC TGC . Lys Phe Ile Cys 6 875	
Leu Pro Val Pro 1		GTG ACC ACC CTG . Val Thr Thr Leu 890	
		CAC ATG AAG CAG His Met Lys Gln : 905	
•		GTC CAG GAG CGC . Val Gln Glu Arg	

AAG	GAC	GAC	GGC	AAC	TAC	AAG	ACC	CGC	GCC	GAG	GTG	AAG	TTC	GAG	GGC	2832
Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu	Gly	
	930					935					940					
010		ama	CMC.	220	000	2000	020	omo.		000	3.000	G1.G	mmc	220	CNC	2000
				AAC Asn												2880
945	THE	neu	vai	ASII	950	116	Giu	Deu	гуэ	955	116	ASP	FILE	гу	960	
243					550					,,,,					300	
GAC	GGC	AAC	ATC	CTG	GGG	CAC	AAG	CTG	GAG	TAC	AAC	TAC	AAC	AGC	CAC	2928
Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Tyr	Asn	Ser	His	
				965					970					975		
											~~~					2076
				ATG												2976
ASII	vaı	ıyı	980	Met	Ala	Asp	гуѕ	985	гуs	ASII	GIY.	116	990	Vai	ASII	
			200					,,,,					,,,,			
TTC	AAG	ATC	CGC	CAC	AAC	ATC	GAG	GAC	GGC	AGC	GTG	CAG	CTC	GCC	GAC	3024
Phe	Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Ser	Val	Gln	Leu	Ala	Asp	
		995				:	1000					1005				
																2000
-				AAC Asn												3072
	1010	GIII	GIII	ASII		1015	116	GIY	ASP	-	1020	vaı	Leu	nea	FIO	
	-0-0															
GAC	AAC	CAC	TAC	CTG	AGC	ACC	CAG	TCC	GCC	CTG	AGC	AAA	GAC	CCC	AAC	3120
Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Ala	Leu	Ser	Lys	Asp	Pro	Asn	
1025					1030					1035					1040	
010		000	CAM	CAC	3.000	CITIC	CITIC	cmc	CAC	mmc	CITIC	200		ccc	ccc	3168
				His					•						GGG Glv	3100
Gra	шуэ	9	_	1045	1100	<b>V</b> 01	Deu		1050		•			1055	027	
ATC	ACT	CTC	GGC	ATG	GAC	GAG	CTG	TAC	AAG	TAA			•			3201
Ile	Thr		_	Met	Asp	Glu			Lys							
		:	1060				:	1065								
		12	) TNT	FORM	מדדמ	N FO	R SFY	מד כ	NO ·	151.						

#### (2) INFORMATION FOR SEQ ID NO:151:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1066 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:151:

Met Glu Ala Glu Gly Ser Ser Ala Pro Ala Arg Ala Gly Ser Gly Glu

1 5 5 10 10 15

Gly Ser Asp Ser Ala Gly Gly Ala Thr Leu Lys Ala Pro Lys His Leu
20 25 30

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Trp	Arg	His 35	Glu	Gln	His	His	Gln 40	Tyr	Pro	Leu	Arg	Gln 45	Pro	Gln	Phe
Arg	Leu 50	Leu	His	Pro	His	His 55	His	Leu	Pro	Pro	Pro 60	Pro	Pro	Pro	Ser
Pro 65	Gln	Pro	Gln	Pro	Gln 70	Cys	Pro	Leu	Gln	Pro 75	Pro	Pro	Pro	Pro	Pro 80
Leu	Pro	Pro	Pro	Pro 85	Pro	Pro	Pro	Gly	Ala 90	Ala	Arg	Gly	Arg	Tyr 95	Ala
Ser	Ser	Gly	Ala 100	Thr	Gly	Arg	Val	Arg 105	His	Arg	Gly	Tyr	Ser 110	Asp	Thr
Glu	Arg	Tyr 115	Leu	Tyr	Cys	Arg	Ala 120	Met	Asp	Arg	Thr	Ser 125	Tyr	Ala	Val
Glu	Thr 130	Gly	His	Arg	Pro	Gly 135	Leu	Lys	Lys	Ser	Arg 140	Met	Ser	Trp	Pro
Ser 145	Ser	Phe	Gln	Gly	Leu 150	Arg	Arg	Phe	Asp	Val 155	Asp	Asn	Gly	Thr	Ser 160
Ala	Gly	Arg	Ser	Pro 165	Leu	Asp	Pro	Met	Thr 170	Ser	Pro	Gly	Ser	Gly 175	Leu
Ile	Leu	Gln	Ala 180	Asn	Phe	Val	His	Ser 185	Gln	Arg	Arg	Glu	Ser 190	Phe	Leu
Tyr	Arg	Ser 195	Asp	Ser	Asp	Tyr	Asp 200	Leu	Ser	Pro	Lys	Ser 205	Met	Ser	Arg
Asn	Ser 210	Ser	Ile	Ala	Ser	Asp 215	Ile	His	Gly	qaA	Asp 220	Leu	Ile	Val	Thr
Pro 225	Phe	Ala	Gln	Val	Leu 230	Ala	Ser	Leu	Arg	Thr 235	Val	Arg	Asn	Asn	Phe 240
Ala	Ala	Leu	Thr	Asn 245	Leu	Gln	Asp	Arg	Ala 250	Pro	Ser	Lys	Arg	Ser 255	Pro
Met	Cys	Asn	Gln 260	Pro	Ser	Ile	Asn	Lys 265	Ala	Thr	Ile	Thr	Glu 270	Glu	Ala
Tyr	Gln	Lys 275	Leu	Ala	Ser	Glu	Thr 280	Leu	Glu	Glu	Leu	Asp 285	Trp	Суѕ	Leu
_	Gln 290					295					300				
305	Asn	_			310					315					320
	Met			325					330					335	
	Leu	_	340					345					350	_	
	Lys	355					360					365			
	10 370					375					380				
385	Val	-			390		-			395					400
	Asn	_		405					410					415	
	Arg		420					425					430		
	Leu	435					440					445			
Met	Thr 450	ren	GIU	чар	піЗ	455	пIS	ATG	Asp	vaı	460	īŸĽ	піз	ASII	ASII

The His Ala Ala Asp Val Val Gln Ser Thr His Val Leu Leu Ser Thr 465																
Pro Ala   Leu Glu   Ala   Val   Phe Thr   Asp   Leu Glu   11e   Leu   Ala   Ala   11e   Ala		His	Ala	Ala	Asp		Val	Gln	Ser	Thr		Val	Leu	Leu	Ser	
Phe Ala		Ala	Leu	Glu	Ala		Phe	Thr	Asp	Leu		Ile	Leu	Ala	Ala	
So			_				_		_		_			_		
See   Val   Leu   Glu   Ass   His   His   Leu   Ala   Val   Gly   Phe   Lys   Leu   Leu   Gln   Solo   So	Phe	Ala	Ser		Ile	His	Asp	Val	-	His	Pro	GIĀ	Val		Asn	Gin
Ser	Phe	Leu		Asn	Thr	Asn	Ser		Leu	Ala	Leu	Met	_	Asn	Asp	Ser
Single   S	Ser	Val		Glu	Asn	His	His		Ala	Val	Gly	Phe		Leu	Leu	Gln
545         556         557         558         569         569         565         570         Val Leu Ala Thr Asp Met Ser 575         570         583         583         583         585         590         590         11																
Ser   Leu   Arg   Lys   Met   Val   Ite   Asp   Ite   Val   Leu   Ata   Thr   Asp   Met   Ser   Ser		Glu	Asn	Cys	Asp		Phe	Gln	Asn	Leu		Lys	Lys	Gln	Arg	
Levy   His   Met   Asn   Levy   Levy   Ala   Asp   Levy   Levy   Levy   Sep   Sep		Leu	Arg	Lys	Met		Ile	Asp	Ile	Val		Ala	Thr	Asp	Met	
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S95	цуѕ	птэ	riec		Leu	Deu	AIG	ASP		рур	1111	mec	Vai		1111	цуs
	Lys	Val		Ser	Ser	Gly	Val		Leu	Leu	Asp	Asn	_	Ser	Asp	Arg
Thr         Lys         Pro         Leu         Gln         Lys         Arg         Gln         Thr         Asp         Arg         Lys         Ho         G35         Lys         Ho         G1u         Arg         Glu         Arg         Glu         Arg         Glu         Arg         Glu         Arg         Gly         Me         Cys         Asp         Lys         His         Asn         Ala         Ser         Val         Glu         Lys         Ser         G10         Cys         Asp         Asp         Lys         His         Asp         Ala         Ser         Val         Glu         Lys         Ser         G10         Cys         Asp	Ile		Val	Leu	Gln	Asn		Val	His	Cys	Ala	-	Leu	Ser	Asn	Pro
Glu Phe Phe Arg Gln Gly Asp Arg Glu Arg Glu Arg Gly Met Glu Ile 645	Thr		Pro	Leu	Gln	Leu		Arg	Gln	Trp	Thr		Arg	Ile	Met	Glu
Ser         Pro         Met         Cys         Asp         Lys         His         Asn         Ala         Ser         Val         Glu         Lys         Gln         Val         665							_					_				
Carre   Carr	Glu	Phe	Phe	Arg		Gly	Asp	Arg	Glu	_	Glu	Arg	Gly	Met		Ile
Fig.	Ser	Pro	Met	_	Asp	Lys	His	Asn		Ser	Val	Glu	Lys		Gln	Val
Leu   Val   His   Pro   Asp   Ala   Gln   Asp   Tle   Leu   Asp   Thr   Leu   Glu   Asp   Asp   Asp   Glo   G95   For   Tor   Tor	Gly	Phe		Asp	Tyr	Ile	Val		Pro	Leu	Trp	Glu		Trp	Ala	Asp
Arg Glu Trp Tyr Gln Ser Thr Ile Pro Gln Ser Pro Ser Pro Ala Pro 705	Leu			Pro	Asp	Ala			Ile	Leu	Asp			Glu	Asp	Asn
Asp Asp Pro Glu Glu Gly Arg Gln Gly Gln Thr Glu Lys Phe Gln Phe 725	Arg		Trp	Tyr	Gln	Ser		Ile	Pro	Gln	Ser		Ser	Pro	Ala	Pro
Glu Leu Thr         Leu Glu Glu Glu Asp Gly Glu Ser Asp Thr Glu Lys Asp Ser 740         735         736         735         736         735         736 <td></td> <td><b>3</b></td> <td>Desa</td> <td><b>61</b></td> <td>C1</td> <td></td> <td>7</td> <td>C1 =</td> <td>C1</td> <td>C1-</td> <td></td> <td>C1.,</td> <td>T 1 10</td> <td>Dho</td> <td>Cln</td> <td></td>		<b>3</b>	Desa	<b>61</b>	C1		7	C1 =	C1	C1-		C1.,	T 1 10	Dho	Cln	
Gly Ser Gln Val       Glu Glu Glu Asp       Thr Ser Cys       Ser Asp       Ser Lys       Thr Leu 755         Cys       Thr Gln Asp       Ser Glu Ser Glu Ser Thr Glu Ile Pro Leu Asp Glu Gln Val 770       Tr 775       Tr 775       Tr 780       <	ASP	ASP	PIO	GIU		GIY	Arg	GIII	GIY		1111	Giu	БУБ	FILE		FILE
Cys       Thr Gln Asp Ser Glu Ser Thr Glu Ile Pro Leu Asp Glu Gln Val 770       Thr Gln Asp Ser Glu Ser Thr Glu Ile Pro Leu Asp Glu Gln Val 770       Thr Glu Glu Glu Ala Val Gly Glu Glu Glu Glu Glu Glu Glu Ser Gln Pro Glu Ala Cys 785       Thr Glu Glu Glu Ala Val Gly Glu Glu Glu Glu Glu Ser Gln Pro Glu Ala Cys 800         Val Ile Asp Asp Asp Arg Ser Pro Ro	Glu	Leu	Thr		Glu	Glu	Asp	Gly		Ser	Asp	Thr	Glu	-	Asp	Ser
Cys Thr Gln Asp Ser Glu Ser Thr Glu Ile Pro Leu Asp Glu Gln Val 770	Gly	Ser		Val	Glu	Glu	Asp		Ser	Суѕ	Ser	Asp		Lys	Thr	Leu
Glu Glu Glu Ala Val Gly Glu	Cys			Asp	Ser	Glu			Glu	Ile	Pro			Glu	Gln	Val
785       790       795       800         Val Ile Asp Asp Asp Asp Ser Pro Asp Thr Bit Gly Ile Leu Gln Ser Thr Roll Ile Pro Asp Asp Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly 820       815         Val Glu Leu Phe Thr Gly Val Val Val Pro Bit Leu Val Glu Leu Asp Gly 835       840         Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp 850       855         Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys 865       870         Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val	Glu		Glu	Ala	Val	Glv		Glu	Glu	Glu	Ser		Pro	Glu	Ala	Cvs
Nai																
Glu Glu Leu Phe Thr Gly Val Glu Val Glu Leu Rap Gly 835       825       836         Asp Val Asn Gly His Lys Phe 855       840       845       845         Asp Val Asn Gly His Lys Phe 855       855       860       860         Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe 11e Cys Thr Thr Gly Lys 865       870       870       875       875       880         Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val       880       880	Val	Ile	Asp	Asp	_	Ser	Pro	Asp	Thr		Gly	Ile	Leu	Gln		Thr
Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly 835	Val	Pro	Arg		Arg	Asp	Pro	Pro		Ala	Thr	Met	Val		Lys	Gly
Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp 850 855 865  Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys 865 870 870 875 885  Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val	Glu	Glu			Thr	Gly	Val			Ile	Leu	Val			Asp	Gly
Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys 865 870 875 880 Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val	Asp			Gly	His	Lys			Val	Ser	Gly			Glu	Gly	Asp
865 870 875 880 Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val	Δl=		ጥኒም	ദിഗ	TAVE	Leu		Len	Live	Phe	Tle		Thr	ጥኮተ	Glv	Ive
	865		_	_		870			_		875					880
	Leu	Pro	Val	Pro		Pro	Thr	Leu	Val		Thr	Leu	Thr	Tyr		Val

	Cys	Phe		Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	His		Phe	Phe	
T	<b></b>		900	<b>D</b>	<b>~</b> 1	<b>61</b>	<b></b>		<b>~</b> 1	<b>~1</b>		<b></b>	910	-1	_,	
Lys S	ser	915	Met	Pro	GIU	GIĀ	920	vaı	GIN	GIU	Arg	925	116	Phe	Pne	
Lys A	Asp 930	Asp	Gly	Asn	Tyr	Lys 935	Thr	Arg	Ala	Glu	Val 940	Lys	Phe	Glu	Gly	
Asp ?	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	Lys	Glu	
945					950					955					960	
Asp (	Gly	Asn	Ile	Leu 965	Gly	His	Lys	Leu	Glu 970	Tyr	Asn	Tyr	Asn	Ser 975	His	
Asn V	Val	Tyr	Ile 980	Met	Ala	Asp	Lys	Gln 985	Lys	Asn	Gly	Ile	Lys 990	Val	Asn	
Phe I	Lys	Ile 995	Arg	His	Asn		Glu L000	Asp	Gly	Ser		Gln 1005	Leu	Ala	Asp	
His ?	Tyr		Gln	Asn	Thr			Gly	Asp	Gly			Leu	Leu	Pro	
	010					1015		-	-		1020					
Asp A	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Ala	Leu	Ser	Lys	Asp	Pro	Asn	
025				1	L030					1035					L040	
Glu I	Lys	Arg	_		Met	Val	Leu			Phe	Val	Thr			Gly	
~1 - 6		<b>-</b>		1045		G1	<b>.</b>		1050					1055		
Ile	ınr		GIŻ	met	Asp	GIU		1yr 1065	Lys							
		-	.000				•	1003			-					
		(2)	IN	ORM	OITA	v FOI	R SEG	O ID	NO:	152:					•	
	( i	i) SI	QUE	VCE (	CHARA	ACTE	RIST:	ics:								
		(A)	LENC	TH:	3024	1 bas	se pa	airs								
							-									
		(B)	TYPE		ıcle:		_									
		(C)	STRA	E: nu ANDEI	ones	ic ad	cid ingle									
		(C)	STRA	E: nu ANDEI	ıcle	ic ad	cid ingle									
	/ -	(C) (D)	STRA TOPO	E: nu ANDEI OLOGY	oness 7: 1:	ic ad S: s: inean	eid ingle									
		(C) (D)	STRA TOPO	E: nu ANDEI OLOGY CULE	ones	ic ad S: s: inean	eid ingle									
		(C) (D)	STRA TOPO	E: NU ANDEI OLOGY CULE	oness 7: 1:	ic ad S: s: inean	eid ingle									
		(C) (D) Li) N	STRA TOPO OLEO FEATU	E: nu ANDEI DLOGY TULE JRE:	oness 7: 1:	ic ad S: s: inean	eid ingle	e	nce							
		(C) (D) (ii) N (x) H	STRA TOPO MOLEC FEATU NAM	E: nu ANDEI DLOGY CULE JRE:	nclei ONESS Y: 1: TYPI	ic ad S: s: inean E: cI	eid ingle ONA	e	nce							
		(C) (D) (ii) M (x) H (A) (B)	STRATOPO MOLECTEATUM NAM	E: NO ANDEI DLOGY CULE JRE: ME/KI	TYPE	ic ad S: s: inean E: cI Codin	eid ingle	e	nce							
	(i)	(C) (D) (ii) N (x) F (A) (B) (D)	STRATOPO MOLECTEATUM NAM LOC	E: nu ANDEI DLOGY TULE JRE: ME/KI CATIO HER I	TYPE  EY: CON: 1	ic ac S: s: inear E: cI Codir	eid ingle ONA ng Se 3021	e eque								
	(i)	(C) (D) (ii) N (x) F (A) (B) (D)	STRATOPO MOLECTEATUM NAM LOC	E: nu ANDEI DLOGY TULE JRE: ME/KI CATIO HER I	TYPE  EY: CON: 1	ic ac S: s: inear E: cI Codir	eid ingle ONA ng Se 3021	e eque	nce Q ID	NO::	152:					
ATG i	i) (c)	(C) (D) (ii) N (x) (A) (B) (D)	STRATOPO	E: MANDEI DLOGY CULE JRE: ME/KI CATIC HER I	TYPH EY: CON: 1 INFOR	ic ac S: s: inear E: cI Codir I3	eid ingle CONA ng Se 3021 CON:	eque:	Q ID			GGG	GCC	TGG	GAA	48
ATG A	(i	(C) (D) (ii) N (x) F (A) (B) (D) (ci) S	TCA	E: NO ANDER DLOGY TULE JRE: JE/KI CATIC HER I	TYPE  EY: CON: 1  INFOR	ic ac S: s: inear E: cI Codir IS RMAT: CRIP:	eid ingle  ONA  ng Se  3021 ION: FION:	eque: : SEX	) ID CAG	ACA	TGT					48
	(i	(C) (D) (ii) N (x) F (A) (B) (D) (ci) S	TCA	E: NO ANDER DLOGY TULE JRE: JE/KI CATIC HER I	TYPE  EY: CON: 1  INFOR	ic ac S: s: inear E: cI Codir IS RMAT: CRIP:	eid ingle  ONA  ng Se  3021 ION: FION:	eque: : SEX	) ID CAG	ACA	TGT					.48
Met s	(i	(C) (D) (ii) N (x) F (A) (B) (D) (ci) S	TCA	E: NO ANDER DLOGS CULE JRE: JE/KH CATIC HER I ENCE CCT Pro	TYPE  EY: CON: 1  INFOR	ic ac S: s: inear E: cI Codir IS RMAT: CRIP:	eid ingle  ONA  ng Se  3021 ION: FION:	eque: : SEX	Q ID CAG Gln	ACA	TGT			Trp		48
Met s	() AGC Ser	(C) (D) (ii) N (A) (A) (B) (D) (C) TGG	STRATION TOPO	E: nu ANDEI CULE IRE: CATIC CATIC CCT Pro 5	TYPE TYPE TYPE TYPE TYPE TYPE TYPE TYPE	ic ac ic ac inean E: cI Codin I CTG Leu	cid ingle	equer : SEX ACG Thr	CAG Gln 10	ACA Thr	TGT Cys	Gly	Ala	Trp 15	Glu	48
Met S	(; AGC Ser	(C) (D) (A) (A) (A) (B) (D) TGG TTP	STRATOPO MOLECTORY NAM LOX OTH TCA Ser CGC	E: nu ANDEI CULE IRE: E/KH CATIC CCT Pro 5	TYPE  TYPE	ic ac ic ac inean E: cI Codin 13 CTG Leu	cid ingle continued and see co	equer : SEX ACG Thr	CAG Gln 10	ACA Thr	TGT Cys	Gly GTC	Ala ATC	Trp 15 CGA	Glu	
Met s	(; AGC Ser	(C) (D) (A) (A) (A) (B) (D) TGG TTP	STRATOPO MOLECTORY NAM LOX OTH TCA Ser CGC	E: nu ANDEI CULE IRE: E/KH CATIC CCT Pro 5	TYPE  TYPE	ic ac ic ac inean E: cI Codin 13 CTG Leu	cid ingle continued and see co	equer : SEX ACG Thr	CAG Gln 10	ACA Thr	TGT Cys	Gly GTC	Ala ATC	Trp 15 CGA	Glu	
Met 1 ATG A	() () AAGC Ser AAAA Llys	(C) (D) (A) (A) (A) (A) (B) (D) (C) (C) TCG GAG GAG GAG	TOPO  MOLECTOR  NAM  LOC  OTH  TCA  Ser  CGC  Arg  20	E: nt ANDEI CULE IRE: IE/KI IE	TYPE TYPE TYPE TYPE TYPE TYPE TYPE TYPE	ic ac S: s: inear E: cI Codir 13 CRIP: CTG Leu ACA	cid ingle in	equel : SE ACG Thr GGA G1y 25	CAG Gln 10 TTT Phe	ACA Thr GGA Gly	TGT Cys AAT Asn	Gly GTC Val	ATC Ile 30	Trp 15 CGA Arg	Glu TGG Trp	96
Met :  ATG :  Met :  CAC :	(; () AAGC Ser AAA Llys	(C) (D) (A) (A) (A) (A) (A) (A) (A) (A) (A) (A	STRATOPO MOLECTORY NAM LOCATORY TCA Ser  CGC Arg 20 GAA	E: nt ANDEI CULE RE: RE/KI CATIC CATIC CATIC CATIC CATIC CATIC CATIC CATIC ACA	TYPE TYPE TYPE TYPE TYPE TYPE TYPE TYPE	ic ac S: s: inear E: cI Codir I3 CTG Leu ACA Thr	cid ingle in	equel SEC ACG Thr GGA Gly 25	CAG Gln 10 TTT Phe	ACA Thr GGA Gly	TGT Cys AAT Asn	Gly GTC Val	Ala ATC Ile 30 TGC	Trp 15 CGA Arg	Glu TGG Trp CAG	
Met 1 ATG A	(; () AAGC Ser AAA Llys	(C) (D) (A) (A) (A) (A) (A) (A) (A) (A) (A) (A	STRATOPO MOLECTORY NAM LOCATORY TCA Ser  CGC Arg 20 GAA	E: nt ANDEI CULE RE: RE/KI CATIC CATIC CATIC CATIC CATIC CATIC CATIC CATIC ACA	TYPE TYPE TYPE TYPE TYPE TYPE TYPE TYPE	ic ac S: s: inear E: cI Codir I3 CTG Leu ACA Thr	cid ingle in	equel SEC ACG Thr GGA Gly 25	CAG Gln 10 TTT Phe	ACA Thr GGA Gly	TGT Cys AAT Asn	Gly GTC Val CAG Gln	Ala ATC Ile 30 TGC	Trp 15 CGA Arg	Glu TGG Trp CAG	96
Met 1 ATG 2 Met 1	(; () AAGC Ser AAA Llys	(C) (D) (A) (A) (A) (A) (A) (A) (A) (A) (A) (A	STRATOPO MOLECTORY NAM LOCATORY TCA Ser  CGC Arg 20 GAA	E: nt ANDEI CULE RE: RE/KI CATIC CATIC CATIC CATIC CATIC CATIC CATIC ACA	TYPE TYPE TYPE TYPE TYPE TYPE TYPE TYPE	ic ac S: s: inear E: cI Codir I3 CTG Leu ACA Thr	cid ingle in	equel SEC ACG Thr GGA Gly 25	CAG Gln 10 TTT Phe	ACA Thr GGA Gly	TGT Cys AAT Asn	Gly GTC Val	Ala ATC Ile 30 TGC	Trp 15 CGA Arg	Glu TGG Trp CAG	96
Met 1 ATG 2 Met 1	() AGC Ser AAA Lys AAT ASn	(C) (D) (A) (A) (A) (A) (A) (A) (A) (A) (A) (A	STRATUPO	E: nu ANDEI CULE URE: ME/KI CATIC HER I COT Pro 5 CTT Leu ACA Thr	TYPE  TYPE	ic ac S: s: inear E: cI CCodir 11 CTG Leu ACA Thr	cid ingle in	eequents: SEC ACG Thr GGA Gly 25 ATT Ile	CAG Gln 10 TTT Phe GCC Ala	ACA Thr GGA Gly ATC Ile	TGT Cys AAT Asn AAG Lys	Gly GTC Val CAG Gln 45	Ala ATC Ile 30 TGC Cys	Trp 15 CGA Arg CGG Arg	Glu TGG Trp CAG Gln	96
Met S  ATG A  Met I  CAC A  His A	(; AGC Ser AAA Lys AAT ASn	(C) (D) (A) (A) (A) (A) (A) (A) (A) (A) (A) (A	STRATOPO MOLECTEATURE NAM LOX OTH TCA Ser CGC Arg 20 GAA Glu CCCC	E: nu ANDEI CULE URE: E/KI E/KI EATIC CATI Pro 5 CTT Leu ACA Thr	TYPE  TYPE	ic ac S: s: inear E: cI Codin 1 CTG Leu ACA Thr	cid ingle in	equents: SEC ACG Thr GGA Gly 25 ATT Ile	CAG Gln 10 TTT Phe GCC Ala	ACA Thr GGA Gly ATC Ile	TGT Cys AAT Asn AAG Lys	Gly GTC Val CAG Gln 45	Ala ATC Ile 30 TGC Cys	Trp 15 CGA Arg CGG Arg	Glu TGG Trp CAG Gln ATC	96 144

55 ATG AGA AGG CTG ACC CAC CCC AAT GTG GTG GCT GCC CGA GAT GTC CCT 240 Met Arg Arg Leu Thr His Pro Asn Val Val Ala Ala Arg Asp Val Pro 70 75 GAG GGG ATG CAG AAC TTG GCG CCC AAT GAC CTG CCC CTG CTG GCC ATG 288 Glu Gly Met Gln Asn Leu Ala Pro Asn Asp Leu Pro Leu Leu Ala Met 85 90 GAG TAC TGC CAA GGA GGA GAT CTC CGG AAG TAC CTG AAC CAG TTT GAG 336 Glu Tyr Cys Gln Gly Gly Asp Leu Arg Lys Tyr Leu Asn Gln Phe Glu 100 105 AAC TGC TGT GGT CTG CGG GAA GGT GCC ATC CTC ACC TTG CTG AGT GAC 384 Asn Cys Cys Gly Leu Arg Glu Gly Ala Ile Leu Thr Leu Leu Ser Asp 115 120 ATT GCC TCT GCG CTT AGA TAC CTT CAT GAA AAC AGA ATC ATC CAT CGG 432 Ile Ala Ser Ala Leu Arg Tyr Leu His Glu Asn Arg Ile Ile His Arg 135 GAT CTA AAG CCA GAA AAC ATC GTC CTG CAG CAA GGA GAA CAG AGG TTA 480 Asp Leu Lys Pro Glu Asn Ile Val Leu Gln Gln Gly Glu Gln Arg Leu 150 155 ATA CAC AAA ATT ATT GAC CTA GGA TAT GCC AAG GAG CTG GAT CAG GGC 528 Ile His Lys Ile Ile Asp Leu Gly Tyr Ala Lys Glu Leu Asp Gln Gly 165 170 AGT CTT TGC ACA TCA TTC GTG GGG ACC CTG CAG TAC CTG GCC CCA GAG 576 Ser Leu Cys Thr Ser Phe Val Gly Thr Leu Gln Tyr Leu Ala Pro Glu 180 185 CTA CTG GAG CAG CAG AAG TAC ACA GTG ACC GTC GAC TAC TGG AGC TTC Leu Leu Glu Gln Gln Lys Tyr Thr Val Thr Val Asp Tyr Trp Ser Phe 195 200 205 GGC ACC CTG GCC TTT GAG TGC ATC ACG GGC TTC CGG CCC TTC CTC CCC 672 Gly Thr Leu Ala Phe Glu Cys Ile Thr Gly Phe Arg Pro Phe Leu Pro 215 220 210 AAC TGG CAG CCC GTG CAG TGG CAT TCA AAA GTG CGG CAG AAG AGT GAG 720 Asn Trp Gln Pro Val Gln Trp His Ser Lys Val Arg Gln Lys Ser Glu 225 230 GTG GAC ATT GTT GTT AGC GAA GAC TTG AAT GGA ACG GTG AAG TTT TCA 768 Val Asp Ile Val Val Ser Glu Asp Leu Asn Gly Thr Val Lys Phe Ser 245 AGC TCT TTA CCC TAC CCC AAT AAT CTT AAC AGT GTC CTG GCT GAG CGA Ser Ser Leu Pro Tyr Pro Asn Asn Leu Asn Ser Val Leu Ala Glu Arg

265

CTG GAG Leu Glu										864
GGC ACG Gly Thr 290										912
GAC ATC Asp Ile 305										960
ACC ATC Thr Ile										1008
AAG GCC Lys Ala	Arg							_	_	1056
CTG CTG Leu Leu								_		1104
CAG TGT Gln Cys 370		 			 					1152
GAT CTT Asp Leu 385							_		_	1200
ATC TCC Ile Ser						_			_	1248
CCC AAG Pro Lys	Arg					_		_	_	1296
GTC TGG Val Trp										1344
CAG GGA Gln Gly 450										1392
CTC TCC Leu Ser 465										1440
GCC AAG Ala Lys										1488

		485			490				495		
			GAG Glu								1536
			GAG Glu								1584
			GTA Val								1632
			AGC Ser 550				_	_	_		1680
			CAA Gln								1728
			CGA Arg								1776
			ATT Ile								1824
			AAA Lys								1872
 			GAA Glu 630								1920
			CTG Leu								1968
			TGT Cys								2016
			GCC Ala							ATG Met	2064
			GCC Ala							AAG Lys	2112

			CTC TGC ACC CTG Leu Cys Thr Leu 715	_
AAT GCC ATA CAG Asn Ala Ile Gln			GAC CAG AGT TTC Asp Gln Ser Phe	
			GAA GAG CAC AGC Glu Glu His Ser 750	Cys Leu
			GAT CCA CCG GTC Asp Pro Pro Val 765	
			GGG GTG GTG CCC Gly Val Val Pro 780	_
			AAG TTC AGC GTG Lys Phe Ser Val 795	
			CTG ACC CTG AAG Leu Thr Leu Lys	
	Lys Leu Pro		CCC ACC CTC GTG Pro Thr Leu Val 830	Thr Thr
			TAC CCC GAC CAC Tyr Pro Asp His 845	
Gln His Asp Phe 850	Phe Lys Ser 855	Ala Met Pro	GAA GGC TAC GTC Glu Gly Tyr Val 860	. Gln Glu
			TAC AAG ACC CGC Tyr Lys Thr Arc 875	
			CGC ATC GAG CTC Arg Ile Glu Leu	
		_	GGG CAC AAG CTC Gly His Lys Let 910	Glu Tyr
			GCC GAC AAG CAC Ala Asp Lys Glr	

915 920 GGC ATC AAG GTG AAC TTC AAG ATC CGC CAC AAC ATC GAG GAC GGC AGC 2832 Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser 935 940 GTG CAG CTC GCC GAC CAC TAC CAG CAG AAC ACC CCC ATC GGC GAC GGC 2880 Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly 950 CCC GTG CTG CCC GAC AAC CAC TAC CTG AGC ACC CAG TCC GCC CTG 2928 Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu 965 970 AGC AAA GAC CCC AAC GAG AAG CGC GAT CAC ATG GTC CTG CTG GAG TTC 2976 Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe 980 985 GTG ACC GCC GCG ATC ACT CTC GGC ATG GAC GAG CTG TAC AAG TAA 3024 Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys 995 1000 (2) INFORMATION FOR SEQ ID NO:153: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1007 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:153:

(v) FRAGMENT TYPE: internal

Met Ser Trp Ser Pro Ser Leu Thr Thr Gln Thr Cys Gly Ala Trp Glu 10 Met Lys Glu Arg Leu Gly Thr Gly Gly Phe Gly Asn Val Ile Arg Trp 25 His Asn Gln Glu Thr Gly Glu Gln Ile Ala Ile Lys Gln Cys Arg Gln 40 Glu Leu Ser Pro Arg Asn Arg Glu Arg Trp Cys Leu Glu Ile Gln Ile 55 Met Arg Arg Leu Thr His Pro Asn Val Val Ala Ala Arg Asp Val Pro 70 75 Glu Gly Met Gln Asn Leu Ala Pro Asn Asp Leu Pro Leu Leu Ala Met 90 Glu Tyr Cys Gln Gly Gly Asp Leu Arg Lys Tyr Leu Asn Gln Phe Glu 105 Asn Cys Cys Gly Leu Arg Glu Gly Ala Ile Leu Thr Leu Leu Ser Asp 120 125 Ile Ala Ser Ala Leu Arg Tyr Leu His Glu Asn Arg Ile Ile His Arg 135 130

Asp 145	Leu	Lys	Pro	Glu	Asn 150	Ile	Val	Leu	Gln	Gln 155	Gly	Glu	Gln	Arg	Leu 160
Ile	His	Lys	Ile	Ile 165	Asp	Leu	Gly	Tyr	Ala 170	Lys	Glu	Leu	Asp	Gln 175	Gly
Ser	Leu	Суѕ	Thr 180	Ser	Phe	Val	Gly	Thr 185	Leu	Gln	Tyr	Leu	Ala 190	Pro	Glu
Leu	Leu	Glu 195	Gln	Gln	Lys	Tyr	Thr 200	Val	Thr	Val	Asp	Tyr 205	Trp	Ser	Phe
Gly	Thr 210	Leu	Ala	Phe	Glu	Cys 215	Ile	Thr	Gly	Phe	Arg 220	Pro	Phe	Leu	Pro
Asn 225	Trp	Gln	Pro	Val	Gln 230	Trp	His	Ser	Lys	Val 235	Arg	Gln	Lys	Ser	Glu 240
Val	Asp	Ile	Val	Val 245	Ser	Glu	Asp	Leu	Asn 250	Gly	Thr	Val	Lys	Phe 255	Ser
Ser	Ser	Leu	Pro 260	Tyr	Pro	Asn	Asn	Leu 265	Asn	Ser	Val	Leu	Ala 270	Glu	Arg
		275					280		Met			285	_		_
	290					295			Gly		300				
Asp 305	Ile	Leu	Asn	Leu	Lys 310	Leu	Val	His	Ile	Leu 315	Asn	Met	Val	Thr	Gly 320
Thr	Ile	His	Thr	Tyr 325	Pro	Val	Thr	Glu	Asp 330	Glu	Ser	Leu	Gln	Ser 335	Leu
_			340					345	Ile				350		
Leu	Leu	Gln 355	Glu	Ala	Gly	Leu	Ala 360	Leu	Ile	Pro	Asp	Lys 365	Pro	Ala	Thr
	370					375			Glu	_	380			_	
Asp 385	Leu	Val	Phe	Leu	Phe 390	Asp	Asn	Ser	Lys	Ile 395	Thr	Tyr	Glu	Thr	Gln 400
Ile	Ser	Pro	Arg	Pro 405	Gln	Pro	Glu	Ser	Val 410	Ser	Cys	Ile	Leu	Gln 415	Glu
Pro	Lys	Arg	Asn 420	Leu	Ala	Phe	Phe	Gln 425	Leu	Arg	Lys	Val	Trp 430	Gly	Gln
	_	435					440	_	Glu	_	_	445			
	450		_			455			Leu		460				_
	Ser	Lys	Met	Lys		Ser	Met	Ala	Ser		Ser	Gln	Gln	Leu	_
465	Tuc	LON	7 cm	Dhe	470 Pho	Tare	ጥኮሎ	Sor	Ile	475	Tle	λεν	T 011	Clu	480
	=			485					490					495	_
_			500					505	Thr		_		510		
	-	515					520		Glu		-	525			
	530					535			Met		540			_	
	Asp	Leu	Gln	Arg		Pro	Met	Gly	Arg		Gln	Gly	Gly	Thr	
545	7	T 6	C1	C1	550		7~~	C1	T.CV	555	7~~	7~~	Tou	7	560
ASD	wsp	neu	GIU	565	GIII	via	wrg	GIU	Leu 570	īÀĽ	ALG	wrg	Den	575	GIU

Lys	Pro	Arg	Asp 580	Gln	Arg	Thr	Glu	Gly 585	Asp	Ser	Gln	Glu	Met 590	Val	Arg
Leu	Leu	Leu 595	Gln	Ala	Ile	Gln	Ser 600	Phe	Glu	Lys	Lys	Val 605	Arg	Val	Ile
Tyr	Thr 610	Gln	Leu	Ser	Lys	Thr 615	Val	Val	Cys	Lys	Gln 620	Lys	Ala	Leu	Glu
Leu 625	Leu	Pro	Lys	Val	Glu 630	Glu	Val	Val	Ser	Leu 635	Met	Asn	Glu	Asp	Glu 640
Lys	Thr	Val	Val	Arg 645	Leu	Gln	Glu	Lys	Arg 650	Gln	Lys	Glu	Leu	Trp 655	Asn
Leu	Leu	Lys	Ile 660	Ala	Суѕ	Ser	Lys	Val 665	Arg	Gly	Pro	Val	Ser 670	Gly	Ser
		675					680	Leu				685			
	690					695		Ser			700				
705					710			His		715					720
				725				Glu	730					735	
	_		740					Glu 745					750		
		755					760	Ala				765			
	770					775					780		•		Leu
Val 785	Glu	Leu	Asp	GIĀ	790	Val	Asn	GIĀ	HIS	ьуs 795	Pne	ser	vaı	Sei	Gly 800
				805					810					815	
			820					825					830		Thr
		835					840					845			Lys
	850	_				855					860				Glu
Arg 865		Ile	Phe	Phe	Lys 870	Asp	Asp	GIY	Asn	17yr 875		Thr	Arg	ALA	61u 880
Val	Lys	Phe	Glu	Gly 885		Thr	Leu	Val	Asn 890		Ile	Glu	Leu	Lys 895	Gly
			900					905					910	)	Tyr
	_	915					920					925	,		Asn
_	930					935					940				Ser
Val 945		Leu	Ala	Asp	His 950		Gln	Gln	Asn	Thr 955		Ile	Gly	Asr	960
		Leu	Leu	Pro 965	Asp		His	Tyr	Leu 970	Ser		Glr	Ser	Ala 975	Leu
Ser	Lys	Asp	Pro 980	Asn		Lys	Arg	Asp 985	His		Val	Leu	1 Let 990		ı Phe
Val	Thr	Ala 995		Gly	lle	Thr	Leu 1000	Gly	Met	. Asp	Glu	Let 1005		Lys	5

## (2) INFORMATION FOR SEQ ID NO:154:

130

(2) 1111	0.441111011 1011 01	Q 15 NO.151.	
(A) LENG (B) TYPE (C) STRAI (D) TOPO	CE CHARACTERIST TH: 2793 base point of the control	pairs	
(A) NAM (B) LOC (D) OTH	E/KEY: Coding S ATION: 12790 ER INFORMATION	)	
		C TTT AGA AGG CAT ( D Phe Arg Arg His :	
		r GCG GGA CGG AGT ( r Ala Gly Arg Ser : 25	
		A ATT CTC CAA GCA A	
		G TAT CGA TCC GAC A L Tyr Arg Ser Asp 60	
		G AAC TCC TCC ATT of Asn Ser Ser Ile . 75	
His Gly Asp Asp		r CCA TTT GCT CAG r Pro Phe Ala Gln ' 90	
		r GCT GCA TTA ACT A e Ala Ala Leu Thr A 105	
		C ATG TGC AAC CAA C D Met Cys Asn Gln	

AAA GCC ACC ATA ACA GAG GAG GCC TAC CAG AAA CTG GCC AGC GAG ACC

Lys Ala Thr Ile Thr Glu Glu Ala Tyr Gln Lys Leu Ala Ser Glu Thr

		CAG CTA GAG Gln Leu Glu 155		
 		AAC AAG TTT Asn Lys Phe 170		
		ATG AGT CGG Met Ser Arg		
		TTA GAT AAG Leu Asp Lys		
		AAG GAG AAA Lys Glu Lys 220		
	_	TTG ATG CAC Leu Met His 235		
		GTT AAA ACT Val Lys Thr 250	_	
		AAC AAA TGG Asn Lys Trp		
		CGG CCC TTG Arg Pro Leu		
		TTA AAA ACA Leu Lys Thr 300		
		ACT CTC GAA Thr Leu Glu 315		
		CAT GCT GCA His Ala Ala 330		
		GCT TTG GAG Ala Leu Glu		_
		GCC AGT GCA Ala Ser Ala		

355		360	365	
			ATC AAT ACA AA Ile Asn Thr Ass 380	
		_	TTA GAG AAC CA Leu Glu Asn Hi 395	_
			AAC TGT GAC AT Asn Cys Asp Il	· -
	s Lys Gln Arg	_	AGG AAA ATG GT Arg Lys Met Va 43	l Ile Asp
			ATG AAT CTA CT Met Asn Leu Le 445	
		_	ACA AGC TCT GG Thr Ser Ser Gl 460	
			GTT CTT CAG AA Val Leu Gln As 475	
			CCT CTC CAG CT Pro Leu Gln Le	
	p Arg Ile Met		TTC CGC CAA GG Phe Arg Gln Gl 51	y Asp Arg
			ATG TGT GAC AA Met Cys Asp Ly 525	
			ATA GAC TAT AT Ile Asp Tyr Il 540	
		•	CAC CCT GAC GC His Pro Asp Al 555	
			TGG TAC CAG AG Trp Tyr Gln Se	

.

											·		
				_	CCT Pro								1776
					TTT Phe 600								1824
		_	_		AGT Ser		_		_	_			1872
					CTT Leu								1920
					GTT Val				_				1968
					TGT Cys								2016
					ACG Thr 680			_					2064
					GGC Gly			_	_	_	_	_	2112
					GGC Gly				_				2160
					GAT Asp								2208
					AAG Lys								2256
					GTG Val 760								2304
					TTC Phe					_			2352
					TTC Phe			_				_	2400

785	790	795	800
		CTG GTG AAC CGC A	
Leu Lys Gly I		AAC ATC CTG GGG 0 Asn Ile Leu Gly 1 830	
		TAT ATC ATG GCC O Tyr Ile Met Ala 2 845	
		ATC CGC CAC AAC ABC ABC ABC ABC ABC ABC ABC AB	
		CAG CAG AAC ACC Gln Gln Asn Thr 875	_
		CAC TAC CTG AGC . His Tyr Leu Ser	
Ser Ala Leu S		CGC GAT CAC ATG Arg Asp His Met 910	
		CTC GGC ATG GAC Leu Gly Met Asp 925	
TAC AAG TAA Tyr Lys 930			2793

- (2) INFORMATION FOR SEQ ID NO:155:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 930 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
  (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:155:

Met Met His Val Asn Asn Phe Pro Phe Arg Arg His Ser Trp Ile Cys

1 5 10 15

Phe	Asp	Val	Asp 20	Asn	Gly	Thr	Ser	Ala 25	Gly	Arg	Ser	Pro	Leu 30	Asp	Pro
Met	Thr	Ser 35	Pro	Gly	Ser	Gly	Leu 40	Ile	Leu	Gln	Ala	Asn 45	Phe	Val	His
Ser	Gln 50	Arg	Arg	Glu	Ser	Phe 55	Leu	Tyr	Arg	Ser	Asp 60	Ser	Asp	Tyr	Asp
Leu 65	Ser	Pro	Lys	Ser	Met 70	Ser	Arg	Asn	Ser	Ser 75	Ile	Ala	Ser	Asp	Ile 80
His	Gly	Asp	Asp	Leu 85	Ile	Val	Thr	Pro	Phe 90	Ala	Gln	Val	Leu	Ala 95	Ser
	_		100	_				105					110	Gln	_
_		115		_			120		_			125		Ile	
-	130					135		_		_	140			Glu	
145					150	_				155				Gln	160
_				165					170	-				Met 175	
	_		180					185					190	Asn	
		195					200			_	-	205		Glu	
	210					215	_		_		220		_	Arg	
Met 225	Ser	Gln	Ile	Ser	Gly 230	Val	Lys	Lys	Leu	Met 235	His	Ser	Ser	Ser	Leu 240
Thr	Asn	Ser	Ser	Ile 245	Pro	Arg	Phe	Gly	Val 250	Lys	Thr	Glu	Gln	Glu 255	Asp
			260				_	265		_			270	His	
		275					280					285		Ile	
	290					295					300			Ile	
Val 305	Asp	Thr	Leu	Ile	Thr 310	Tyr	Leu	Met	Thr	Leu 315	Glu	Asp	His	Tyr	His 320
Ala	Asp	Val	Ala	Tyr 325	His	Asn	Asn	Ile	His 330	Ala	Ala	Asp	Val	Val 335	Gln
Ser	Thr	His	Val 340	Leu	Leu	Ser	Thr	Pro 345	Ala	Leu	Glu	Ala	Val 350	Phe	Thr
Asp	Leu	Glu 355	Ile	Leu	Ala	Ala	Ile 360	Phe	Ala	Ser	Ala	Ile 365	His	Asp	Val
Asp	His 370	Pro	Gly	Val	Ser	Asn 375	Gln	Phe	Leu	Ile	Asn 380	Thr	Asn	Ser	Glu
Leu 385	Ala	Leu	Met	Tyr	Asn 390	Asp	Ser	Ser	Val	Leu 395	Glu	Asn	His	His	Leu 400
Ala	Val	Gly	Phe	Lys 405	Leu	Leu	Gln	Glu	Glu 410	Asn	Cys	Asp	Ile	Phe 415	Gln
Asn	Leu	Thr	Lys 420	Lys	Gln	Arg	Gln	Ser 425	Leu	Arg	Lys	Met	Val 430	Ile	Asp
Ile	Val	Leu 435	Ala	Thr	Asp	Met	Ser 440	Lys	His	Met	Asn	Leu 445	Leu	Ala	Asp

Leu	Lys 450	Thr	Met	Val	Glu	Thr 455	Lys	Lys	Val	Thr	Ser 460	Ser	Gly	Val	Leu
Leu 465	Leu	Asp	Asn	Tyr	Ser 470	Asp	Arg	Ile	Gln	Val 475	Leu	Gln	Asn	Met	Val 480
His	Cys	Ala	Asp	Leu 485	Ser	Asn	Pro	Thr	Lys 490	Pro	Leu	Gln	Leu	Tyr 495	Arg
Gln	Trp	Thr	Asp 500	Arg	Ile	Met	Glu	Glu 505	Phe	Phe	Arg	Gln	Gly 510	Asp	Arg
Glu	Arg	Glu 515	Arg	Gly	Met	Glu	Ile 520	Ser	Pro	Met	Cys	Asp 525	Lys	His	Asn
Ala	Ser 530	Val	Glu	Lys	Ser	Gln 535	Val	Gly	Phe	Ile	Asp 540	Tyr	Ile	Val	His
Pro 545	Leu	Trp	Glu	Thr	Trp 550	Ala	Asp	Leu	Val	His 555	Pro	Asp	Ala	Gln	Asp 560
Ile	Leu	Asp	Thr	Leu 565	Glu	Asp	Asn	Arg	Glu 570	Trp	Tyr	Gln	Ser	Thr 575	Ile
Pro	Gln	Ser	Pro 580	Ser	Pro	Ala	Pro	Asp 585	Asp	Pro	Glu	Glu	Gly 590	Arg	Gln
Gly	Gln	Thr 595	Glu	Lys	Phe	Gln	Phe 600	Glu	Leu	Thr	Leu	Glu 605	Glu	Asp	Gly
	610					615			Ser		620				
625					630				Thr	635					640
				645					Glu 650					655	
Glu	Glu	Ser	Gln 660	Pro	Glu	Ala	Cys	Val 665	Ile	Asp	Asp	Arg	Ser 670	Pro	Asp
Thr	Thr	Gly 675	Ile	Leu	Gln	Ser	Thr 680	Val	Pro	Arg	Ala	Arg 685	Asp	Pro	Pro
	690					695	_		Glu		700				
705					710				Val	715					720
				725					Thr 730				•	735	
_			740					745	Pro				750		
		755					760					765			Asp
	770					775			Ser		780				
785					790				Asp	795					800
				805					810					815	
			820					825	Gly				830		
		835					840		Val			845			
	850					855					860				Glu
Asp 865	Gly	Ser	Val	Gln	Leu 870	Ala	Asp	His	Tyr	Gln 875	Gln	Asn	Thr	Pro	Ile 880

Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln	
.885 890 895	
Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu	
900 905 910	
Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu	
915 920 925	
Tyr Lys	
930	
(2) INFORMATION FOR SEQ ID NO:156:	
(2) INFORMATION FOR SEQ ID NO.130.	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 37 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(2)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:156:	
GTAAGCTTCG AACATGATGC ACGTGAATAA TTTTCCC	37
(2) INFORMATION FOR SEQ ID NO:157:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 34 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:157:	
(XI) SEQUENCE SESSECULIZATION. SEQ ES NO. 15.	
GTAAGCTTCG AACATGGAGG CAGAGGGCAG CAGC	34
(2) INFORMATION FOR SEQ ID NO:158:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 34 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:158:	
CONTROL PROPERCY ACCIONAL COCC	34
GTAAGCTTCG AACATGGCTC AGCAGACAAG CCCG	34
(2) INFORMATION FOR SEQ ID NO:159:	
(D) THE OTHER POST OF THE PROPERTY.	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 37 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	

(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:159:	
GTGAATTCCC GTCGTGTCAG GAGAAGCATC ATCTATG	37
(2) INFORMATION FOR SEQ ID NO:160:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 24 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:160:	
GTGAATTCAA CCATGGAGCG GGCC	24
(2) INFORMATION FOR SEQ ID NO:161:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 23 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:161:	
GTGGTACCCA GTTCCGCTTG GCC	23
(2) INFORMATION FOR SEQ ID NO:162:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 24 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:162:	
GTCTCGAGGC AAGATGGCTG ACCC	24
(2) INFORMATION FOR SEQ ID NO:163:	
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 25 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:163:	·	
GTGGATCCGA GCTCTTGACT TCGGG	25	
(2) INFORMATION FOR SEQ ID NO:164:		
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 31 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li></ul>		
(D) TOPOLOGY: linear	•	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:164:	31	
"		
(2) INFORMATION FOR SEQ ID NO:165:		
<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 25 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>		
(ii) MOLECULE TYPE: cDNA		
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:165:		
GTGGTACCCA TGAGGCCTGC TCCAG	25	

Figure 1

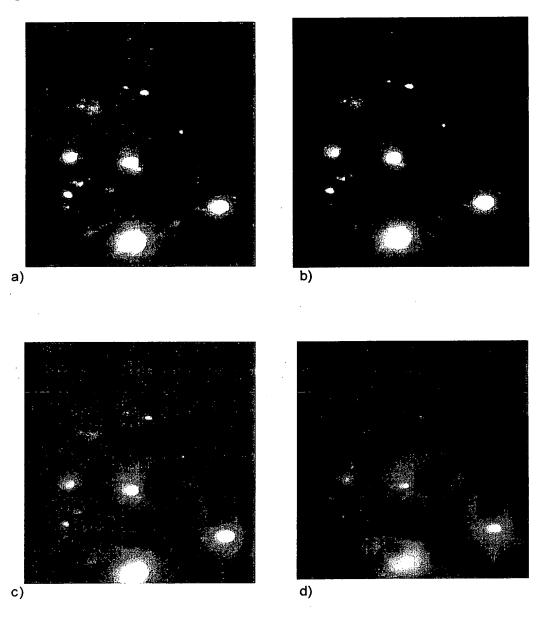


Figure 2

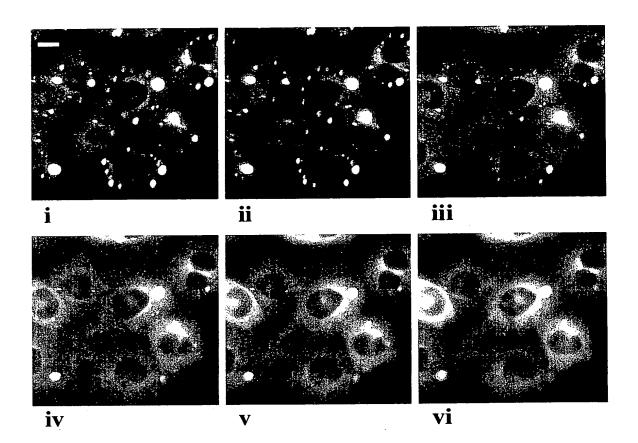
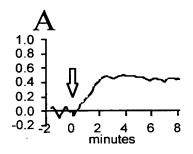
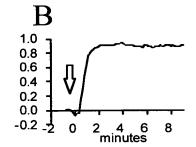
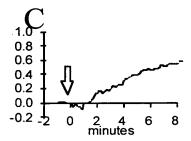
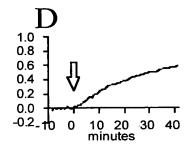


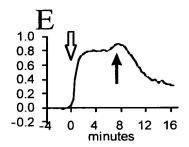
Figure 3

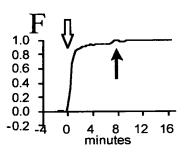


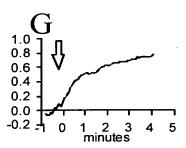


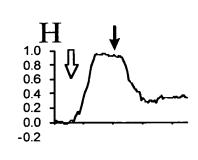












Modtaget PD 15 OKT. 1998

Figure 4

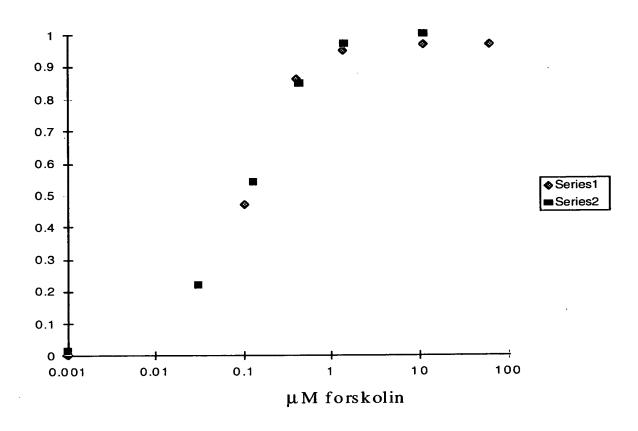
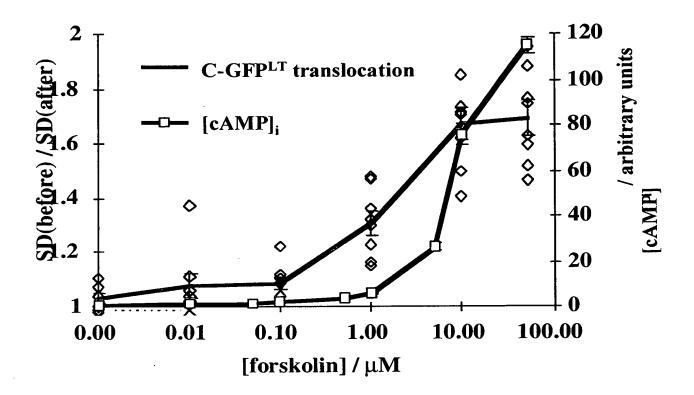


Figure 5

[forskolin]µM	t <sub>1/2max</sub> / s	t <sub>max</sub> /s
1	115±21	310±31
10	69±14	224±47
50	47±10	125±28

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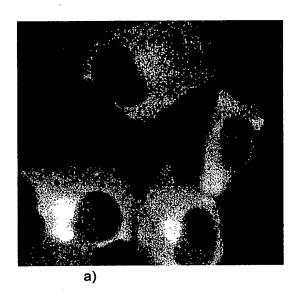
Figure 6

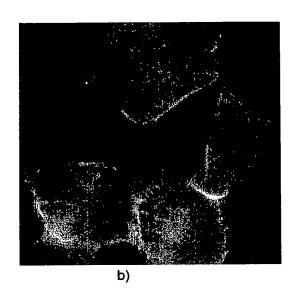


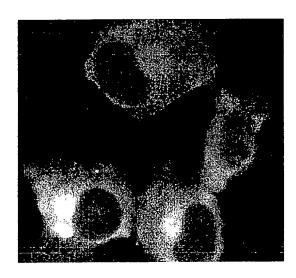
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Figure 7

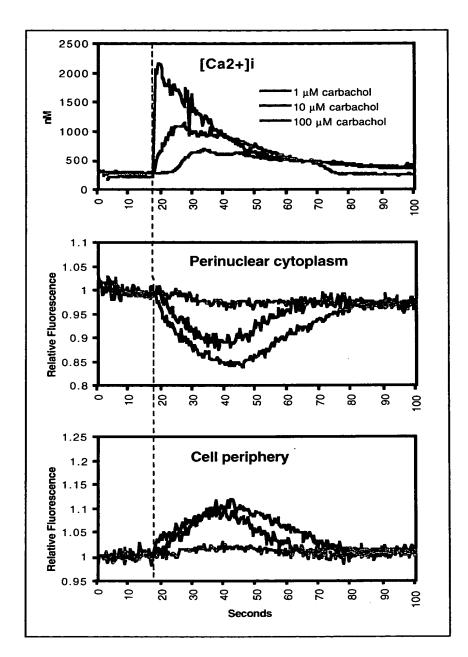






c)

Figure 8



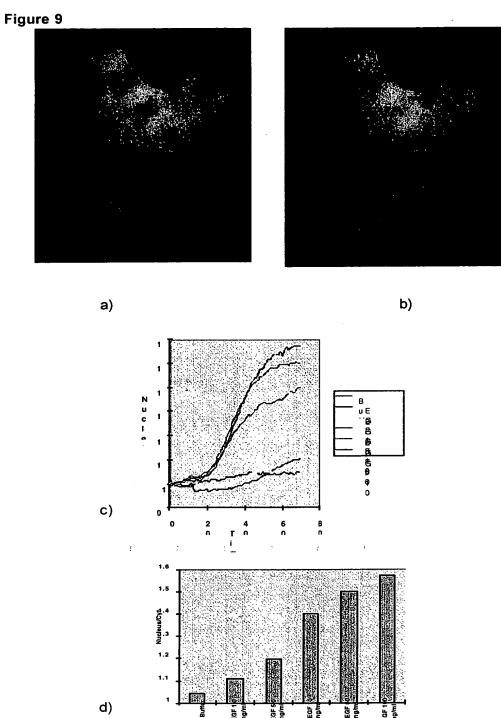
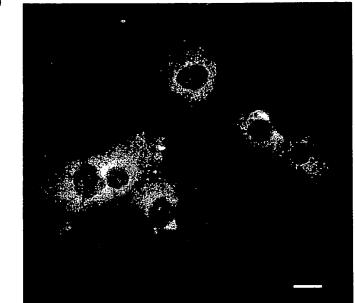
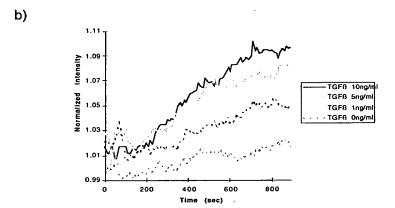


Figure 10

a)





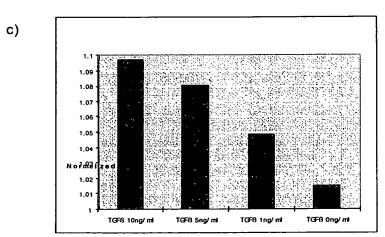


Figure 11

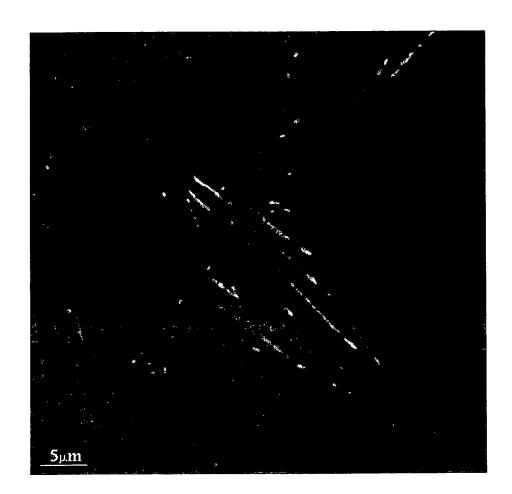
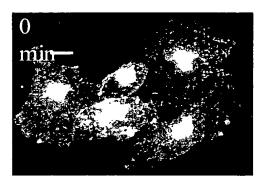
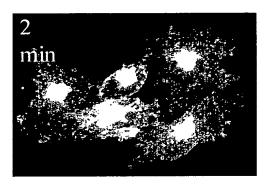
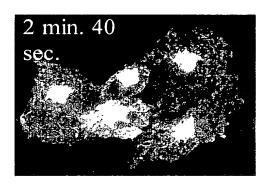
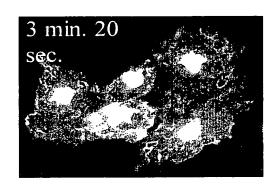


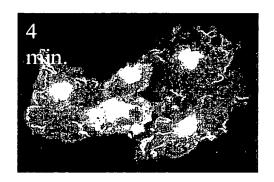
Fig. 12











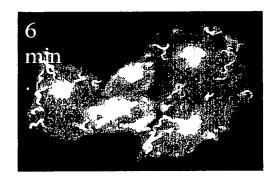


Figure 13

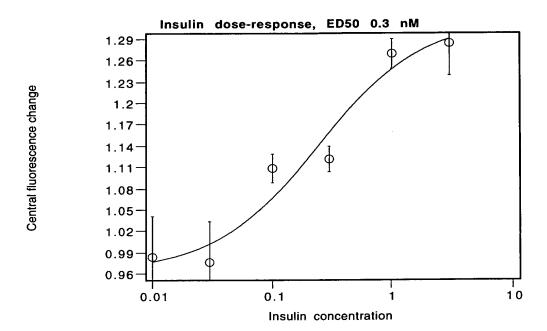


Figure 14

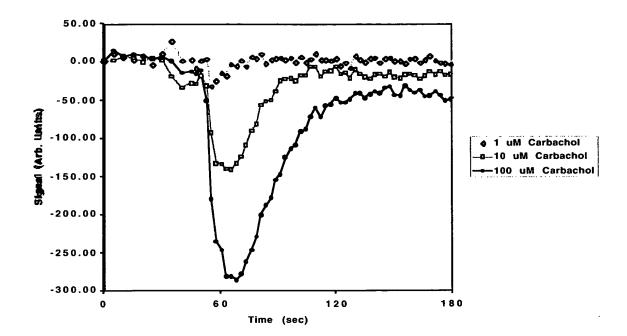
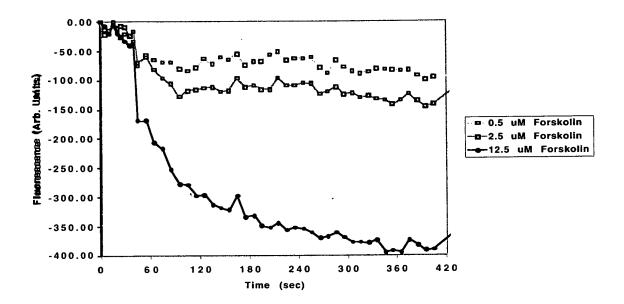
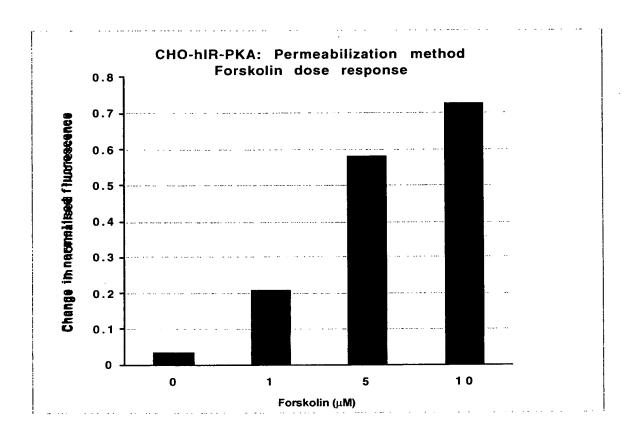


Figure 15

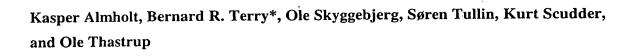


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Figure 16



# Changes in intracellular cAMP visualised using a cAMP-dependent protein kinase-green fluorescent protein hybrid.



BioImage, Novo Nordisk A/S, 28 Mørkhøj Bygade, DK-2860 Søborg.

Keywords: PKA, cAK, C-subunit, cAMP, measurement, live-cell, GFP, redistribution

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#### **ABSTRACT**

A novel method to monitor changes in intracellular cAMP concentration ([cAMP],) within intact living cells has been developed based on a fusion of the catalytic subunit of cAMP-dependent protein kinase to green fluorescent protein (GFP). In stably transfected unstimulated fibroblasts, fusion protein fluorescence was highly concentrated in aggregates throughout the cytoplasm and absent in the nucleus. Stimulation with the adenylate cyclase activator forskolin caused the release of tagged catalytic subunits from the cytoplasmic aggregates within minutes, resulting in an increasingly homogeneous distribution of GFP fluorescence throughout the cytoplasm. The observed redistribution was completely reversible: removal of forskolin led to the return of fluorescence to the cytoplasmic aggregates. Spot-photobleach measurements showed that the rate of exchange of GFP-labelled catalytic subunits at these aggregates increased in proportion to [cAMP]. The localisation of the fusion protein was also sensitive to receptor stimulation. In fibroblasts stably expressing the G-protein coupled glucagon receptor, generation of an increased [cAMP], through glucagon stimulation resulted in a redistribution of tagged catalytic subunit similar to that observed after forskolin addition. Conversely, in fibroblasts overexpressing the G<sub>i</sub>-protein coupled α2a adrenoreceptor, addition of norepinephrine after forskolin stimulation led to a reversal of the fusion protein redistribution.

#### INTRODUCTION

The cAMP-dependent protein kinase  $(cAK)^1$  is a ubiquitous serine/threonine protein kinase. cAK is recognised as the only mediator of intracellular cAMP signals in eukaryotes<sup>2</sup>, with the exception of certain ion channels<sup>3</sup>. The cAK holoenzyme is an  $R_2C_2$  tetramer consisting of a regulatory (R) dimer and two catalytic (C) subunits<sup>2</sup>. Presently, four isoforms of the regulatory subunit (RI $\alpha$ , RI $\beta$ , RII $\alpha$  and RII $\beta$ ) and three isoforms of the catalytic subunit (C $\alpha$ , C $\beta$  and C $\gamma$ ) have been described<sup>2</sup>. Splice variants of C $\alpha$  and C $\beta$ <sup>4</sup> and possible R heterodimers, as reported for RI $\alpha$  and RI $\beta$ <sup>5</sup>, add to the complexity of the cAK holoenzyme. Although the C $\gamma$  isoform is unique with respect to substrate specificity, inhibition and tissue distribution<sup>6</sup>, few reports suggest different roles for C $\alpha$  and C $\beta$  isoforms of the catalytic subunit<sup>7</sup>. In contrast, the RI and RII subunits are reported to be distinct. The cAKI (RI<sub>2</sub>C<sub>2</sub>) holoenzyme is thought to be mainly soluble and cytoplasmic<sup>2</sup> although RI is reported to be associated with

sarcoplasmic membranes<sup>8</sup> and also with a detergent-resistant structure in mammalian sperm<sup>9</sup>. cAKII (RII<sub>2</sub>C<sub>2</sub>) on the other hand is thought to be particulate and RII has been reported to bind to a number of intracellular components, most notably Golgi membranes<sup>10,11</sup> and centrosomes<sup>10,11</sup> but also mitochondria<sup>12</sup>, nuclei<sup>13,14</sup> and cytoskeletal components<sup>11,12</sup>. RII subunits interact with a family of proteins called A-kinase anchoring proteins (AKAP)<sup>15</sup> and this may also be true of RI subunits<sup>16</sup>. The AKAP-RII subunit interaction is presumed to be responsible for localising the cAKII tetramer at these intracellular sites. The NH<sub>2</sub>-terminus of the C subunit is myristoylated<sup>17</sup>, a post-translational modification usually associated with membrane insertion. However, the C subunit does not appear to be membrane attached and while myristoylation may increase the thermostability of the protein, the possible role of myristoylation in its targeting or substrate specificity is still not clear<sup>18</sup>.

The C subunit in the assembled tetramer is believed, although not unanimously<sup>19</sup>, to be catalytically inactive. Activation of cAK is physiologically mediated through G<sub>s</sub>-protein coupled plasma membrane receptors. G<sub>s</sub>-protein activation leads to activation of adenylate cyclases, which generate cAMP. Binding of two molecules of cAMP to each R subunit causes the release and activation of the C subunits. Dissociated C subunits phosphorylate cytoplasmic substrates<sup>20,21</sup> and have been shown to relocalise to the nucleus<sup>22</sup>. The nuclear redistribution mechanism of C subunits may be by simple diffusion through nuclear pores<sup>21</sup>. To date a large number of cytoplasmic and a few nuclear cAK substrates have been reported. An incomplete list of 25 *in vitro* substrates<sup>23</sup> includes several enzymes involved in basic metabolism such as phosphorylase kinase, glycogen synthase and fructose bisphosphatase. Nuclear C subunit regulates transcription of genes under control of the cAMP response element (CRE) by phosphorylating the continuously bound CRE binding protein, (CREB)<sup>24,25</sup>.

Several factors decrease the level of cAK activity. Stimulation of plasma membrane bound G<sub>i</sub>-protein coupled receptors inhibits adenylate cyclases and cAMP is continuously being broken down by a variety of phosphodiesterases. Despite the importance of the cAMP/cAK signalling pathway, there is no easy method to monitor intracellular cAMP concentrations ([cAMP]<sub>i</sub>) in intact living cells. The current method of choice involves fluorescence resonance energy transfer (FRET) between microinjected fluorescently labelled R and C subunits<sup>26</sup>. In the work described herein, the Cα subunit was tagged with a highly fluorescent variant of green fluorescent protein (GFP) containing F64L and S65T amino acid substitutions (GFP<sup>LT</sup>) (International

Publication No. WO97/11094). This approach provides a transfectable probe for monitoring the intracellular trafficking of C subunits in response to changes in [cAMP], and represents the first easy method to evaluate changes in [cAMP], in intact living cells in response to extracellular signals.

## Results

## GFP<sup>LT</sup> tagged C had the expected molecular weight.

Lysates of glucagon receptor-transfected baby hamster kidney cells (BHK/GR) stably expressing the C-GFP<sup>LT</sup> fusion protein were characterised by Western blot analysis using polyclonal antibodies directed against the NH<sub>2</sub>-terminus of  $C\alpha$  (Fig. 1). In a separate experiment, lysates of BHK cells, transiently expressing either of the two fusion proteins, were characterised by Western blot analysis using polyclonal antibodies that recognise GFP (data not shown). Taken together, these experiments show that C-GFP<sup>LT</sup> fusion protein is recognised as a unique protein of the expected size by the anti- $C\alpha$  antibody in stably transfected cells and that both fusion proteins have the same molecular weight.

# The fusion protein localised to cytoplasmic aggregates.

The localisation of the two fusion proteins, when transiently expressed in Chinese hamster ovary (CHO) cells, was very different. While GFP<sup>LT</sup>-C was evenly distributed throughout the cytoplasm (Fig. 2A), C-GFP<sup>LT</sup> was found in highly fluorescent aggregates in the cytoplasm (Fig. 2B). These distinct patterns for the two fusions was also seen in transiently transfected human embryonic kidney (HEK293) and BHK/GR cells (data not shown). For unknown reasons it was not possible to make stable transfectants expressing the GFP<sup>LT</sup>-C fusion, whereas this procedure was straightforward with the C-GFP<sup>LT</sup> fusion. The distribution of GFP<sup>LT</sup>-C in transiently transfected CHO cells did not change when [cAMP]<sub>i</sub> was raised by the addition of 50 μM forskolin (n=6, data not shown). The following results are therefore based only on work with the C-GFP<sup>LT</sup> fusion.

# Increased [cAMP]<sub>i</sub> caused the release of fusion protein from cytoplasmic aggregates.

Within 2-3 minutes of treatment of CHO/C-GFPLT cells with forskolin, C-GFPLT fluorescence dispersed from the bright aggregates and filled the cytoplasm (Fig. 3A, 1 µM forskolin), remaining in this distribution for as long as forskolin was present (cells were followed up to two hours). The probe did not enter the nuclear compartment to any clearly observable extent. Higher doses of forskolin increased the rate and extent of probe redistribution. The responses depicted in Figure 3B-G have all been quantified from image data, as described in the experimental protocol. Table 1 gives a comparison of the average temporal profiles of fusion protein redistribution in response to the three forskolin concentrations shown in Figure 3B. Addition of 1 mM dibutyryl cAMP (dbcAMP) (n=6), a membrane permeable cAMP analogue, which is not degraded by phosphodiesterases, caused a similar but slower response (Fig. 3C). Addition of 100 μM 3-isobutyl-1-methylxanthine (IBMX) (n=4), a cell permeable phosphodiesterase inhibitor, caused a similar, slow response (Fig. 3D), even in the absence of adenylate cyclase stimulation. Addition of buffer (n=2) had no effect (data not shown). As a control for the behaviour of the fusion protein, GFPLT alone was expressed in CHO cells and these also given 50  $\mu M$  forskolin (n=5); the uniform diffuse distribution characteristic of GFP in these cells was unaffected by such treatment (data not shown).

To test the reversibility of the fusion protein redistribution, CHO/C-GFP<sup>LT</sup> cells were treated with 10  $\mu$ M forskolin (n=2) and washed repeatedly (5-8 times) with 37°C buffer. Although the plant terpenoid forskolin is lipophilic, it is possible to remove its effect by washing with aqueous buffer<sup>22</sup>. In these experiments, fusion protein began to return to its prestimulatory localisation within 2-3 min (Fig. 3E). In fact the fusion protein returned to a pattern of fluorescent cytoplasmic aggregates virtually indistinguishable from that observed before forskolin stimulation. To test whether the return of fusion protein to the cytoplasmic aggregates reflected a decreased [cAMP], cells were treated with a combination of 10  $\mu$ M forskolin and 100  $\mu$ M IBMX (n=2); when washed repeatedly (5-8 times) with 37°C buffer containing 100  $\mu$ M IBMX the fusion protein did not return to its prestimulatory localisation after removal of forskolin (Fig. 3E).

To test the probe's response to receptor activation of adenylate cyclase, stably transfected BHK/GR,C-GFP<sup>LT</sup> cells were exposed to glucagon stimulation. In these cells, addition of 100 nM glucagon (n=2) caused the release of C-GFP<sup>LT</sup> from the cytoplasmic aggregates and a resulting permanent redistribution of the fusion protein to

a more even cytoplasmic distribution within 2-3 min (Fig. 3F). Similar but less pronounced effects were seen at lower glucagon concentrations (n=2, data not shown). Addition of buffer (n=2) had no effect over time (data not shown). CHO/C-GFP<sup>LT</sup> cells, transiently transfected with the α2a adrenoreceptor (ARα2a), were treated with 10 μM forskolin then, in the continued presence of forskolin, exposed to 10 μM norepinephrine to stimulate the exogenous adrenoreceptors. This treatment led to reaggregation of C-GFP<sup>LT</sup> within the fluorescent structures, consistent with a receptor-induced decrease in [cAMP]<sub>i</sub> (Fig. 3G).

# Rate of recovery from photobleach of C-GFP<sup>LT</sup> aggregates is dependent on forskolin concentration.

Photobleach measurements were made to confirm that changes seen in the distribution of C-GFP<sup>LT</sup> fluorescence were a result of changes in the rate of turnover of C-GFP<sup>LT</sup> upon the aggregates. The fluorescence of an entire C-GFP<sup>LT</sup> aggregate within a cell could be effectively bleached within 2 to 5 seconds by a stationary laser beam at full intensity. After bleaching, aggregates recovered their fluorescence, indicating a dynamic exchange of C-GFP<sup>LT</sup> at these loci (Fig. 4A). The rate of recovery from spot photobleach was highly reproducible at each particular concentration of forskolin even in different cells (Fig. 4B). Both the extent and rate of recovery increased with the forskolin treatment given. Most recovery curves required at least two exponentials to fit them adequately. Given the limits of the experimental procedure, the curves are used here only to estimate half-times of recovery. To an approximation, half times for recovery can be estimated directly from the slope of reciprocal plots of the fluorescence displacement for the first few time points<sup>27</sup>. Values for half times estimated within the first 3.0 seconds of recovery (Fig. 4C) are plotted as a dose response curve in Figure 5, giving an estimated ½-maximal concentration for forskolin of about 3 μM

## Fusion protein redistribution correlated with [cAMP]<sub>i</sub>

As described above, the time it took for a response to come to completion was inversely related to the forskolin dose (Table 1). In addition the extent of a response was also dose dependent. In an automated imaging system we stimulated CHO/C-GFP<sup>LT</sup> cells with 5 increasing doses of forskolin (n=8). Images were analysed with the same algorithm used

to construct Figure 3B-G. From the results shown in Figure 5, a half maximal stimulation was observed at 1.7 μM forskolin by this method. In parallel, CHO/C-GFP<sup>LT</sup> cells were stimulated with 8 increasing concentrations of forskolin (n=N) and the relative amount of cAMP produced was measured in a scintillation proximity assay (SPA). The ½-maximal concentration for forskolin in the SPA assay was determined to be 9.3 μM (Fig. 5).

## Co-localisation of C-GFP<sup>LT</sup> with labelled ceramide distributions

Figure 6A is an overlay of green and red fluorescence emissions from CHO/C-GFP<sup>LT</sup> cells stained with BODIPY<sup>®</sup> FL C<sub>5</sub>-ceramide (ceramide-FL). The green channel contains the ceramide-FL and GFP<sup>LT</sup> fluorescence; the red channel shows only the ceramide-FL excimer emission. The ceramide-FL probe preferentially accumulates in Golgi membranes<sup>28</sup>. This is most obvious in images formed from the red excimer emissions of the FL-ceramide. The GFP<sup>LT</sup>-bright structures do not stain with the ceramide probe indicating that they are clearly distinct from Golgi membranes.

## Structure of the $GFP^{LT}$ -bright aggregates

Figure 6B shows an iso-surface rendering of 25 deconvolved and reconstructed through-focus wide-field images of a single large C-GFP<sup>LT</sup> aggregate. Each aggregate appears to have the structure of a convoluted tubule or glomerulus, and this is more obvious in the stereo pair (Fig. 6C) derived from the same data set from which the iso-surface rendering was made. It is not completely clear whether each structure is formed from a single fully connected tubule or a small number of discrete tubules in close apposition. The structure is however clearly compact and more complex and structured than a simple amorphous aggregation of C-GFP<sup>LT</sup> molecules. Figure 6B-C is typical of the larger aggregates which are of the order of 2 to 4 μm across. The more numerous smaller aggregates (less than 1 μm across) appear to share the same underlying structural component(s) as their larger counterparts.

#### Discussion

The aim of the present study was to develop a transfectable probe for monitoring changes in [cAMP]. Since cAK is by far the major intracellular effector for cAMP<sup>2</sup>, a measure of its activation should closely reflect physiologically relevant changes in [cAMP].

NH<sub>2</sub>- and COOH-terminal fusions of C subunit were made to a highly fluorescent variant of GFP. Only the C-GFP<sup>LT</sup> fusion responded to changes in [cAMP]<sub>i</sub>. The three-dimensional structure of the C subunit<sup>29,30</sup> reveals that both the NH<sub>2</sub>- and COOH-termini, while far apart, are both located opposite the catalytic cleft and close to the surface of the protein. Comparison with the closely related cGMP-dependent protein kinase, whose R and C subdomains are contained within the same polypeptide chain in R-C order<sup>31</sup>, suggests that the R subunit of cAK may be expected to interact with the NH<sub>2</sub>-terminal region of the C subunit. Furthermore, the surface of the C subunit in the NH<sub>2</sub>-terminal region is hydrophobic<sup>29</sup>, supportive of a protein-protein interaction in this area. An NH<sub>2</sub>-terminal GFP<sup>LT</sup> tag would also prevent post-translational myristoylation (of the NH<sub>2</sub>-terminus) of the C subunit as reported specifically for mouse Cα<sup>18</sup>, while the C-GFP<sup>LT</sup> fusion may well be myristoylated. These factors may explain the very different behaviours of the NH<sub>2</sub>- and COOH-terminal fusions of C subunit to GFP<sup>LT</sup>.

There are reasons to believe, that the C-GFP<sup>LT</sup> fusion protein behaves like the endogenous kinase both with regard to localisation and activation kinetics. Li *et al.* (1996)<sup>11</sup> have, for instance, reported that RII subunits occur as "intensely fluorescent spots" within perinuclear cytoplasm. Skålhegg *et al.* (1997)<sup>32</sup> also reported a granular distribution of RII in both human B and T lymphocytes. Also, the time frame of fusion protein redistribution in response to forskolin addition reported here, corresponds well to the observation of dissociation of microinjected RI $\alpha_2$ C $\alpha_2$  holoenzyme in response to forskolin within 1-2 minutes<sup>26</sup> and the dissociation of endogenous RII<sub>2</sub>C<sub>2</sub> in response to forskolin observed by immunofluorescence after less than 5 min<sup>22</sup>.

In contrast with previous work with microinjected RIIα<sub>2</sub>Cα<sub>2</sub> holoenzyme and Cα subunit<sup>21</sup>, we did not observe any translocation of C-GFP<sup>LT</sup> to the nucleus. A possible explanation could be the increased size of the fusion protein relative to endogenous C subunit. Nuclear pores are thought to allow passage by diffusion of globular proteins of less than 45-60 kDa<sup>33</sup>. The putative size limit of 45-60 kDa may adequately explain the exclusion of the fusion protein (68 kDa), yet passage of endogenous C subunit (41 kDa).

Consistent with this, a microinjected 65 kDa fusion protein of glutathione S-transferase and mouse Ca subunit (GST-C) was excluded from the nucleus<sup>21</sup>.

That the C-GFP<sup>LT</sup> fusion can be released by dbcAMP or treatments which increase [cAMP]<sub>i</sub> suggests that it must recognise and attach to endogenous R subunits (or some subset of the same) and therefore that these R subunits are naturally collected at or on the structures seen in Figures 3A and 6. Reversal of elevated [cAMP]<sub>i</sub>, e.g. by removal of forskolin or stimulation of G<sub>i</sub>-coupled receptors, results in rapid return of fluorescence to the original prestimulatory locations within cytoplasm. These anchoring structures therefore appear to be persistent features within the cytoplasm of CHO/C-GFP<sup>LT</sup> cells. Similar structures and C-GFP<sup>LT</sup> behaviour were also found in transfected BHK cells.

The distribution of fluorescence between aggregates and cytoplasm should reflect the position of a dynamic equilibrium within each cell, determined principally by [cAMP]. This is confirmed by results from spot-photobleach measurements. The rate of fluorescence recovery of aggregates following photobleach measures the net rate of turnover of C subunits at these sites. The rate of recovery is the sum of on and off rates for the association of catalytic with regulatory subunits at these loci, both of which will be governed principally by the concentration of cAMP within the cell (the off rate being governed directly by [cAMP]; the on rate being dependent on the concentration of free C-GFP<sup>LT</sup> in the cytoplasm). Most aggregates completely disappear after full stimulation with forskolin. However, often one aggregate remains, and this is always the biggest and brightest from the unstimulated cell. Nevertheless, as photobleaching can demonstrate, there is active turnover of C-GFP<sup>LT</sup> even at these large fluorescent aggregates which remain in fully stimulated cells. As a further observation, there appears to be considerable mobility of catalytic subunits within the structure of an aggregate, since a stationary laser beam (approx. 0.5-1.0 µm diameter) is able to bleach fluorescence from an entire aggregate of 2-3 µm diameter in 2 to 5 seconds.

The lack of colocalisation of C-GFP<sup>LT</sup> and ceramide fluorescence, the position of aggregates within the cell and their unusual form, suggest that these structures are definitely not associated with Golgi, but may well be constructed of membrane tubules with C-GFP<sup>LT</sup> on the outer surface. Although we have been unable as yet to ascertain the identity of these structures, we have ruled out Golgi membranes. They may however be membranous since fusion protein is apparently freely mobile on them, possible tubular judging by the 3-D recontructed image, and clearly the catalytic subunits are able to

bind to and release from R subunits with ease, suggesting that the latter are anchored to the surface of these structures. They are also persistent within the cytoplasm, and found in all cells transfected thus far with the C-GFP<sup>LT</sup> construct (CHO, HEK293 and BHK).

Figure 5 gives a comparison of an SPA assay conducted in parallel with two different forskolin dose response experiments using the cAK fusion protein. These experiments showed a direct correlation of three parameters: level of [cAMP], turnover rate of C-GFP<sup>LT</sup> at cytoplasmic aggregates, and overall degree of fusion protein redistribution. Data from these three greatly varying methods agree on an ½-maximal concentration for forskolin of between 1.7 to 9.3 μM in this system. As these results show, the cAK fusion protein represents a novel and reliable probe by which dynamic changes in [cAMP], can be measured in intact living cells as they respond to extracellular signals.

## **Experimental protocol**

## Hybrid cDNA construction

Hybrid cDNAs encoding  $\mathrm{NH_{2}}\text{-}$  and COOH-terminal fusions of murine  $\mathrm{C}\alpha$  subunit<sup>34</sup> to GFP<sup>LT</sup> were inserted into the multiple cloning site of the pZeoSV (Invitrogen Corp., San Diego, CA, USA) mammalian expression vector, generating the fusion constructs C-GFPLT and GFPLT-C. Briefly, cDNAs encoding C and GFPLT were amplified by PCR 5'-C, following primers: the using TTGGACACAGCTTTGGACACCCTCAGGATATGGGCAACGCCGCCGCCCCCC 3'-C, AAG; GTCATCTTCTCGAGTCTTTCAGGCGCCCCAAACTCAGTAAACTCCTTGCCA 5'-GFPLT, CAC TTGGACACAGCTTTGGACACGGCGCGCCATGAGTAAAGGAGAAGAACTTT 3'-GFP<sup>LT</sup>, TC and GTCATCTTCTCGAGTCTTACTCCTGAGGTTTGTATAGTTCATCCATGCCATGT . HindIII/AscI restriction endonuclease digested C subunit PCR amplification product and AscI/XhoI digested GFPLT PCR product were ligated with the HindIII/XhoI digested vector for the generation of the C-GFP<sup>LT</sup> fusion construct. Correspondingly the GFP<sup>LT</sup>-C construct was generated by ligating HindIII/Bsu36I digested GFPLT PCR product and Bsu36I/XhoI digested C subunit PCR product with the HindIII/XhoI digested vector. To

generate a similar construct which allowed the expression of GFP<sup>LT</sup> alone, the GFP<sup>LT</sup> PCR product was digested with HindIII/XhoI and ligated with the HindIII/XhoI digested vector.

#### Cell cultures

CHO cells were transfected with the vectors containing hybrid cDNA for the C-GFP<sup>LT</sup> or the GFP<sup>LT</sup>-C fusion proteins using the calcium phosphate precipitate method in HEPES-buffered saline<sup>35</sup>. Stable transfectants were selected using 1000 μg Zeocin/ml (Invitrogen) in the growth medium (DMEM with 1000 mg glucose/l, 10 % foetal bovine serum (FBS), 100 μg penicillin-streptomycin mixture ml<sup>-1</sup>, 2 mM L-glutamine purchased from Life Technologies Inc., Gaithersburg, MD, USA). Untransfected CHO cells were used as the control. To assess the effect of glucagon on fusion protein redistribution, the constructs were stably expressed in BHK/GR cells (Novo Nordisk, Bagsværd, Denmark) overexpressing the human GR. Untransfected BHK/GR cells were used as the control. Expression of GR was maintained with 500 μg G418/ml (*Neo* marker) and C-GFP<sup>LT</sup> was maintained with 500 μg Zeocin/ml (*Sh ble* marker). CHO cells were also simultaneously co-transfected with vectors containing cDNAs for C-GFP<sup>LT</sup> and the human ARα2a (ATCC). Transfected cells are referred to as *e.g.* CHO/C-GFP<sup>LT</sup> cells in the text.

For fluorescence microscopy, cells were allowed to adhere to Lab-Tek chambered coverglasses (Nalge Nunc Int., Naperville, IL, USA) for at least 24 hours and cultured to about 80% confluence. Prior to experiments, the cells were cultured over night without selection pressure in HAM's F12 medium with glutamax (Life Technologies), 100 µg penicillin-streptomycin mixture ml¹ and 0.3 % FBS. This medium has low autofluorescence enabling fluorescence microscopy of cells straight from the incubator.

### **Immunoblotting**

Samples containing 10 µg of protein, determined according to the method of Bradford<sup>36</sup> using the Bio-Rad Protein Assay (Bio-Rad Laboratories, Hercules, CA, USA), were added to SDS sample buffer<sup>35</sup> and run on precast 7.5 % SDS-PAGE gels with a 4 % stacking gel (Bio-Rad). The proteins were transferred to PH79 nitrocellulose membranes (Scleicher & Schuell GmbH., Dassel, Germany) for an hour at 4°C using a Bio-Rad Transfer Blot apparatus (80 V). Non-specific adhesion was blocked by

incubating the membranes over night in 3 % bovine serum albumin Fraction V (Sigma Chemical Company, St. Louis, MO, USA) in Tris-buffered saline (TBS) containing 50 mM Tris pH 7.5 and 0.15 M NaCl and for an hour in 3 % skim milk powder (Difco Laboratories, Detroit, MI, USA) in TBS with 0.1 % Tween20 (TBST). The membranes were incubated for an hour in TBST with 3 % skim milk powder and the primary polyclonal rabbit anti-Cα antibody (Upstate Biotechnology Inc., Lake Placid, NY, USA), which was raised against a peptide corresponding to a 16 amino acid N-terminal stretch of human Ca, diluted 1:1000. After 4 washes of 5 min each with TBST, (horse radish peroxidase-conjugated donkey secondary antibody immunoglobulin from Amersham International plc, Buckinghamshire, UK) diluted 1:5000 in TBS with 3 % skim milk powder was added and incubated for an hour. After 4 washes in TBST and one in TBS, immunoreactivity was detected by enhanced chemiluminescence (ECL) as described by the manufacturer (Amersham) and exposed on Biomax® MR film (Eastman Kodak Company, Rochester, NY, USA). All the steps were performed at room temperature unless otherwise stated.

## Time-lapse recording of fusion protein movement.

Cells were cultured in HAM's F12 medium as described above. The chambers were placed on a temperature regulated microscope stage and kept at 37°C. Fluorescence images were captured using an Axiovert 135 inverted light microscope (Carl Zeiss, Oberkochen, Germany) equipped with a Fluar x40, NA 1.3 oil immersion objective (Zeiss) and a cooled (-40°C) CH1 charged coupled device (CCD) camera (Photometrics Ltd., Tucson, AZ, USA). The microscope was equipped with a 470±20 nm excitation filter, a 505 nm dichroic mirror and a 515±15 nm emission filter (Delta Lys & Optik, Lyngby, Denmark). The excitation light source was a 100W HBO arc lamp.

Redistribution of the C-GFP<sup>LT</sup> fusion protein was quantified using an image analysis program custom written in LabVIEW (National Instruments, Austin, TX, USA). Fluorescent aggregates are segmented from each image using an automatically found threshold based on maximisation of the information measure between the object and the background. The *a priori* entropy of the image histogram is used as the information measure<sup>37</sup>. The area occupied by aggregates in each image is calculated by counting pixels in the segmented areas. The value thus obtained for each image in a series, or treatment pair, is normalised to the value found for the first (unstimulated) image

collected. A value of zero (0) indicates no redistribution of fluorescence from the starting condition. A value of one (1) by this method equals full redistribution.

### Spot photobleaching

A Zeiss LSM 410 with x40 Fluar (as above) was used in spot scan mode at 488 nm to bleach individual fluorescent C-GFPLT aggregates within CHO cells variously treated with forskolin. Fluorescence recovery at the locus of each aggregate was monitored immediately after bleach with successive small-area raster scans just large enough to include most of the cell in which the aggregate lay. Nominal output of the laser at 488 nm, before launch into the microscope, was 10 mW. Subsequent raster scans were also run with the laser at full intensity and without a confocal aperture to allow the first to be made within 0.2 seconds of bleach, and for each scan to be completed within 0.3 seconds (100 x 100 pixels per scan). The recovery of fluorescence for the majority of bleach experiments was measured over a period of 215 seconds, recorded in three consecutive blocks of 10 scans having successive intervals between frames of 0.5, 1 and 5 seconds, and a final set of 15 scans each 10 seconds apart. A single scan collected prior to each bleach exposure served both to establish depth of bleach and to estimate maximum recoverable fluorescence in each experiment. Bleach recovery scans (8-bit images) were analysed using IPlab Spectrum software (Signal Analytics Corp., Vienna, VI, USA). A small region of interest (ROI) of between 6x6 to 10x10 pixels was used to define the area for which fluorescence recovery would be monitored in each experiment, and the average fluorescence within that ROI was measured for successive frames in each time series. The measurement ROIs were slightly larger than the bleached C-GFPLT aggregates to allow for cytoplasmic movements during the measurement period. The total average fluorescence within each frame was also measured to allow fluorescence recovery within C-GFPLT aggregates to be corrected for the minor effects of photobleaching caused by the series of measurement scans.

Results of the spot-bleach experiments are presented as normalised values of displacement from photobleach,  $\Delta F(t)$ , versus time t:

$$\Delta F(t) = [F(\infty) - F(t)]/[F(\infty) - F(0)]$$

where  $F(\infty) = F_1 \cdot R / R_1$ 

 $F(\infty)$  being the maximum recoverable fluorescence within a measurement ROI calculated from the pre-bleach intensity of the target aggregate,  $F_i$ , corrected for total loss of fluorophores within the cell,  $R_i/R_p$ , during the bleach exposure and recovery periods.

#### SPA

CHO/C-GFP<sup>LT</sup> cells were cultured in HAM's F12 medium as described above, but in 96-well plates. The medium was exchanged with Ca<sup>2+</sup>-HEPES buffer containing 100 μM IBMX. The cells were stimulated with different concentrations of forskolin for 10 min. Reactions were stopped with addition of NaOH to 0.14 M and the amount of cAMP produced was measured with the cAMP-SPA kit, RPA538 (Amersham) as described by the manufacturer.

### Automated imaging

A Diaphot300 microscope (Nikon Corp., Tokyo, Japan) coupled to a camera based on the SITe back illuminated 512 x 512 CCD camera (Princeton Instruments Inc., Trenton, NJ, USA) and integrated with a digital data acquisition system using LabVIEW software was configured to allow automated focusing and image-based analyses in 96-well plates. CHO/C-GFP<sup>LT</sup> cells were cultured as described above but in 96-well plates and kept at 37°C throughout the experiments. A fluorescence micrograph of the same field of cells, initially chosen at random, was acquired before and 30 min after forskolin stimulation and analysed as described above.

## Endomembrane labelling with fluorescently tagged ceramides

Golgi membranes in CHO/C-GFP<sup>LT</sup> cells were labelled with ceramide-FL (Molecular Probes Inc., Eugene, OR, USA) at  $0.5~\mu M$  for 20 minutes before washing. Ceramide-FL excited at 480 nm normally emits in the green at about 510 nm, but when concentrated (as in Golgi membranes) the fluorophore forms excimers, resulting in a shift in the emission maximum to greater than  $600~nm^{38}$ . Images were collected at both  $520~\pm~10~nm$  and beyond 570 nm, allowing good separation of GFP<sup>LT</sup> and ceramide-FL signals.

## Structure of the GFP<sup>LT</sup>-bright aggregates

Through-focus images of individual C-GFP<sup>LT</sup> aggregates were collected from chilled cells with a x63 NA 1.4 oil-immersion objective. The built-in focus motor of the Zeiss LSM 410 was used to advance the objective 0.2 µm between images, 25 images per data set. Effective pixel size in the images was 65.6 nm. Data sets were corrected for bleaching and fluctuations in illumination intensity. Out-of-focus information in the images was removed using iterative, constrained, three-dimensional deconvolution (DeltaVision from Applied Precision Inc., Seattle, WA, USA) based on a theoretically calculated point-spread function. The deconvolved images were then reconstructed into a 3-D rotational projection of 40 images (9 degrees between images) using the method of maximum intensity ray-tracing (DeltaVision, Applied Precision, Inc., Seattle, USA). Two adjacent images in this set, re-sized and pixel-smoothed, were used to create the stereo pair shown in Figure 6C. An iso-surface rendering of the 3-D reconstruction was created using Milan software (BitPlane AG, Zurich, Switzerland) (Fig. 6B).

## **Acknowledgments**

We acknowledge Dr. S. P. Bjørn (Novo Nordisk) for developing a mutagenised cDNA clone for the highly fluorescent derivative of green fluorescent protein used in this study, Dr. G. S. McKnight, Howard Hughes Medical Research Institute, Seattle, USA for providing us with his cDNA clone of the  $C\alpha$  subunit of murine cAK and G. Hagel for expert technical assistance.

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## Figure legends

Table 1. Time from initiation of a response to half maximal  $(t_{1/2 max})$  and maximal  $(t_{max})$  C-GFP<sup>LT</sup> redistribution. The data was extracted from curves such as shown in Figure 3B. All  $t_{1/2 max}$  and  $t_{max}$  values are given as mean±SD and are based on a total of 26-30 cells from 2-3 independent experiments for each forskolin concentration. Since the observed redistribution is sustained over time, the  $t_{max}$  values were taken as the earliest time point at which complete redistribution is reached. Note that the values do not relate to the degree of redistribution.

Figure 1. Western blot analysis of lysates containing C-GFP<sup>LT</sup> fusion proteins. Total lysates of BHK/GR,C-GFP<sup>LT</sup> (A) and control BHK/GR (B) cells were probed with an anti-Cα antibody. 500 ng of purified bovine C subunit (C) was included as a positive control and to identify the endogenous C subunit. Although the antibody clearly reacts unspecifically with several proteins in the total lysates, the fusion protein (f) is recognised as a specific band, migrating with an apparent size of 60 kDa, in the transfected cells (A). The endogenous C subunit (e) migrated as predicted by its molecular weight of 41 kDa. It is possible to compare the expression levels of endogenous hamster C subunit and overexpressed mouse fusion proteins in these blots since the immunogenic peptide is conserved between these two species.

Figure 2. Fluorescence micrographs of CHO cells expressing C subunit fusion proteins. The two fusion proteins of the C subunit of cAK show distinct localisation patterns. A. The NH<sub>2</sub>-terminal GFP<sup>LT</sup>-C fusion protein is localised almost evenly throughout the cytoplasm. B. The COOH-terminal C-GFP<sup>LT</sup> fusion protein is highly concentrated in cytoplasmic aggregates, often in one large and several minor structures per cell. Scale bar 10 μm.

Figure 3. Time-lapse analyses of fluorescence redistribution in CHO/C-GFP<sup>LT</sup> cells treated with various agonists. The raw data of each experiment consisted of 60 fluorescence micrographs acquired at regular intervals including several images acquired before the addition of agonist. Six of these images are shown (A) for the typical response to 1 μM forskolin, taken at the time points indicated. The time point t=0 corresponds to the image acquired immediately before the cells were challenged with agonist. Scale bar

10 μm. The charts (B-G) each show a quantification of the responses in each time series. The total area of the highly fluorescent aggregates (see Experimental Protocol) is plotted versus time for each experiment. (B) Redistribution time profiles of the C-GFP<sup>LT</sup> fusion following treatment of cells with various concentrations of forskolin. (C) Response following addition of 1 mM dbcAMP. (D) The effect of 100 μM IBMX on the fusion protein distribution. (E) Demonstrates the reversibility of the forskolin-induced redistribution of C-GFP<sup>LT</sup>, where 10 μM forskolin (open arrow) is followed shortly by repeated washings with buffer (dark arrow). In a parallel experiment, treatment with 10 μM forskolin plus 100 μM IBMX is followed by repeated washing with buffer containing 100 μM IBMX. (F) BHK/GR,C-GFP<sup>LT</sup> cells treated with 100 nM glucagon. (G) CHO/C-GFP<sup>LT</sup> cells transiently transfected with the ARα2a were pretreated with 10 μM forskolin (open arrow) to increase [cAMP], then given 10 μM norepinephrine in the continued presence of forskolin.

Figure 4. (A) Four frames from the recovery sequence following spot photobleach of a large aggregate (arrow) in a CHO/C-GFP<sup>LT</sup> cell exposed to 25 μM forskolin. Times are seconds after bleach. (B) Normalised displacement curves of the fluorescence recovery process in cells exposed to various levels of forskolin. Measurement points are averages±sem (n=4). (C) Linear fits to the first five points of the normalised recovery curves shown in (B). The slope of each line is used as an estimate of the half-time of recovery from bleach at each forskolin concentration.

Figure 5. Parallel dose response analyses of forskolin effects in CHO/C-GFP<sup>LT</sup> cells on: [cAMP], elevation (□), the rate of recovery from spot photobleach (Δ) and induced change in C-GFP<sup>LT</sup> redistribution (•). [cAMP], was measured by SPA assay, analysing the effects of buffer or 8 increasing concentrations of forskolin in these cells. The graph shows a trace of the mean±sem expressed in arbitrary units (n=4 for each data point). Half times for recovery from spot photobleach were estimated from the first 5 time points of the mean value (n=4) curves in Figure 4B. Changes induced in C-GFP<sup>LT</sup> distribution were quantified as described (Experimental Protocol) using fluorescence micrographs taken of the same field of cells prior to and 30 min after the addition of forskolin. The graph shows a trace of the mean±sem at each forskolin concentration (n=8 for each data point). The fitted curves indicate ½-maximal concentration values for

forskolin as: 1.7  $\mu$ M, image-based assay ( $\square$ ); 3.0  $\mu$ M, spot photobleach assay ( $\Delta$ ); 9.3  $\mu$ M, SPA ( $\bullet$ ).

Figure 6. (A) Two images of CHO/C-GFP<sup>LT</sup> cells stained with ceramide-FL, in emission ranges of  $520 \pm 10$  nm and >570 nm, have been superimposed to demonstrate the distinct separateness of Golgi membranes (orange) and C-GFP<sup>LT</sup> fluorescence (green). Scale bar is  $10 \mu m$ . (B) An iso-surface rendering of a single large C-GFP<sup>LT</sup> aggregate (similar to that arrowed in 6A). The image is a reconstruction from 25 through-focus images deconvolved and processed as described (Experimental Protocol). Scale bar  $1 \mu m$ . (C) Stereo pair of the reconstructed images used to generate the iso-surface seen in (B). Each image is smoothed for presentation, the structure originally being 35 pixels high by 27 wide in this orientation. Scale bar  $1 \mu m$ .

## Figure 1

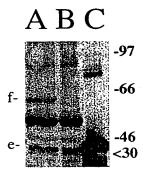
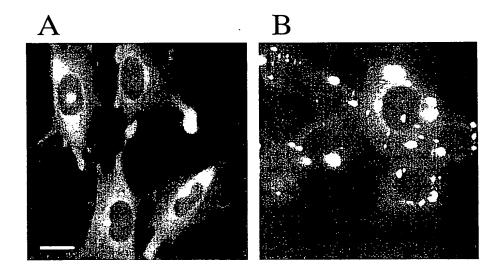


Figure 2



## Figure 3

## A

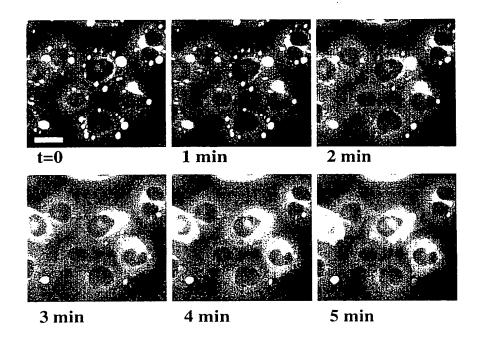


Figure 3

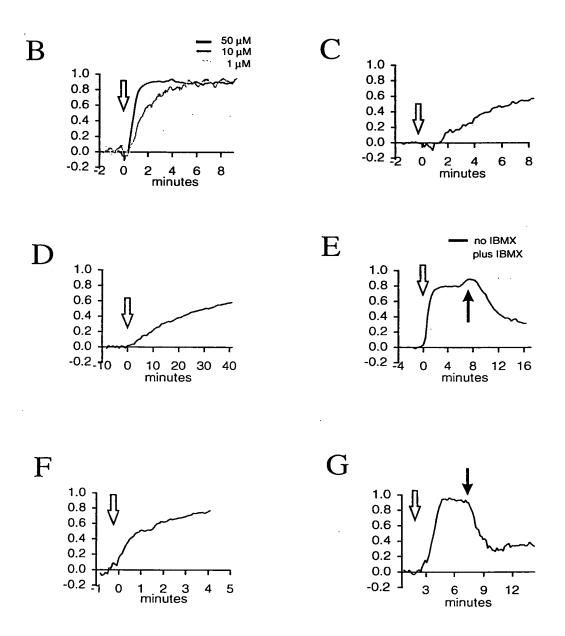
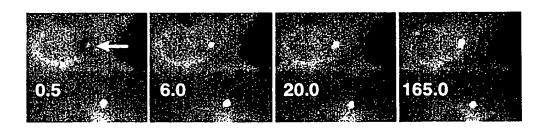
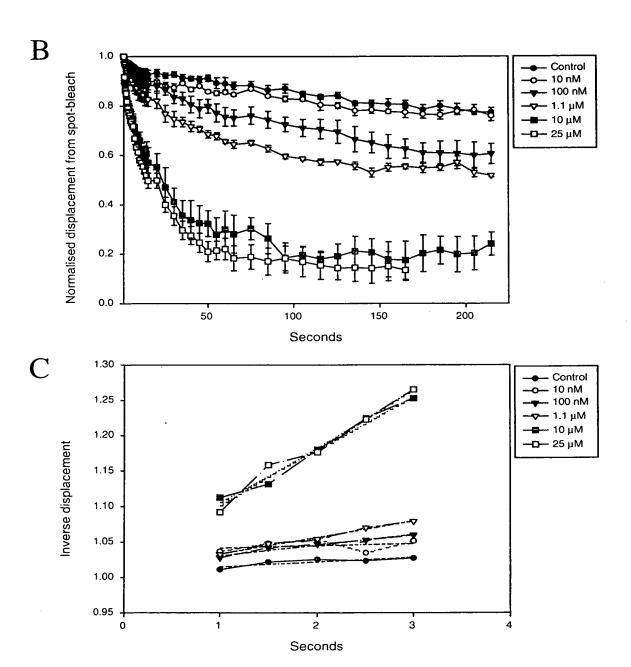


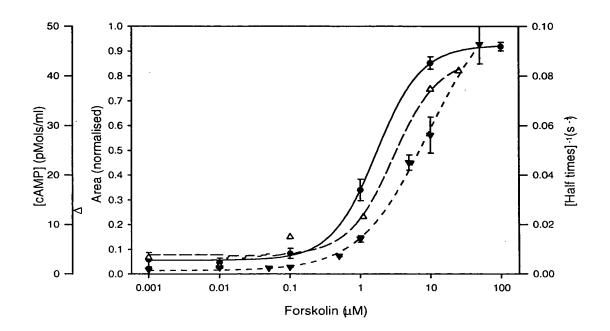
Figure 4

A





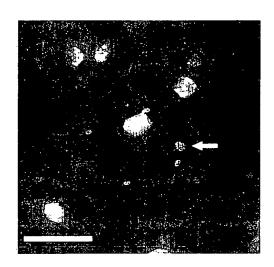
22129DK1 Appendix I

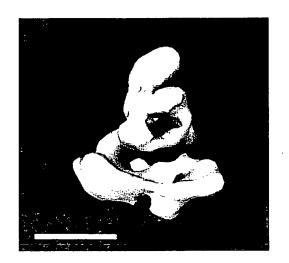


## Figure 6

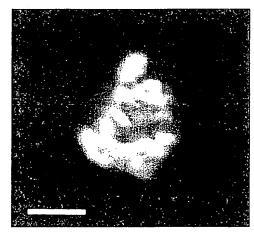
A

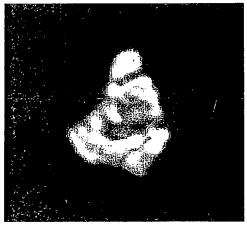
B





C





Left

Right

## Table 1

[forskolin]/µM	t <sub>1/2max</sub> / s	t <sub>max</sub> / s
1	115±21	310±31
10	69±14	224±47
50	47±10	125±28

## METHOD AND APPARATUS FOR HIGH DENSITY FORMAT SCREENING FOR BIOACTIVE MOLECULES

#### FIELD OF THE INVENTION

The invention relates to a method and apparatus for screening large numbers of molecules for biological activities.

#### **BACKGROUND OF THE INVENTION**

Current technology is able to generate large numbers of molecules which may possess potential therapeutic value. Compounds having potentially interesting biological activity may be products of combinatorial or traditional chemistry, a natural product, proteins isolated by one- or two-dimensional gel electrophoresis, or compounds secreted from or expressed by natural or genetically modified animal, plant, microbial or fungal cells (or parts thereof), or displayed by natural or genetically modified viral or phage particles.

Screening methods have been developed which achieve very high throughputs of test compounds. Such methods are termed Ultra High Throughput Screening (UHTS). The present generation of UHTS machines rely upon essentially serial additions of test compounds, usually one test compound per discrete test well. Test well array densities range from between 96 to 3456 wells per plate. Such UHTS machines require sophisticated technologies to dispense microvolumes of many different fluids to selected locations, and also require that the detecting surface for each test molecule generally be separated from other detecting surfaces within the array.

There is a need to develop a method which allows simultaneous screening of large numbers of test compounds for biological activity and potential therapeutic use while avoiding the complications associated with dispensing multiple fluid microvolumes.

#### BRIEF SUMMARY OF THE INVENTION

The invention is directed to a screening method which eliminates the need for delivering microfluid volumes and allows simultaneous parallel screening of large numbers of test compounds. Accordingly, the invention is drawn to a method for screening test

compounds for bioactivity, by contacting an array of test compounds with a detector layer capable of detecting bioactivity, wherein a cell response is indicative of bioactivity.

The method of the invention is a high throughput system for parallel screening of a large number of test compounds. In one embodiment of the method of the invention, 96 to 10,000 test compounds are simultaneously screened for bioactivity in an assay; in a more specific embodiment, 96 to 3456 test compounds are simultaneously screened for bioactivity.

In a more specific embodiment, invention is drawn to a method for screening test compounds for bioactivity, comprising:

- (a) contacting a solid support comprising an array of test compounds with a liquid layer, wherein the liquid layer is in immediate contact with a detector layer and wherein each test compound comes into contact with a localized portion of the liquid layer; and
- (b) registering a response of the detector layer to the test compound, wherein a bioactive test compound is identified.

By "high throughput screening" is meant a method able to screen large number of test compounds for biological activity within a given machine time (i.e. at a rate anywhere from 100 to 100,000 compounds per hour per machine).

The term "parallel screening" refers to a method by which very many compounds are applied simultaneously to the detector layer, and similarly, signals from that detector layer are collected contemporaneously rather than sequentially.

By "array" is meant a regular two-dimensional arrangement of test compounds by which compounds are disposed at the nodes of a rectilinear grid pattern whereby a compound position can be identified by a simple 2-dimensional coordinate.

A "detector layer" means any two-dimensional system which can be used to report biologically relevant information. In one specific embodiment of the method of the invention the detector layer is a monolayer of living cells loaded with a fluorescent reporter dye such as Fluo-3.

By "bioactive" or "bioactivity" is meant an action or influence of a test compound upon the detector layer which results in a response from the detector layer that has direct biological significance or can be interpreted as being a biologically relevant response.

Bioactive agents have the ability to effect physiological parameters of living cells and tissues. Bioactivity includes inducing or suppressing the expression of a protein, activating or inhibiting transcription of a gene, and/or effecting cellular function(s) such as, for example,

intracellular movement and storage of calcium ions, and membrane transportation.

The capacity of a test compound to affect a detector layer, i.e. bioactivity, may be determined in a number of ways known to the art. In specific embodiments of the method of the invention, bioactivity is determined by changes or movements of fluorescent probes present in the detector layer which indicate changes in ionic content, cell metabolism, growth or viability. In a preferred method of the invention, living cells form the detector layer and have specific protein components tagged with a fluorescent agent, such as green fluorescent protein (GFP); changes in GFP fluorescence or distribution within cells indicate a particular cellular response which may be selected for identification of bioactivity.

The phrase "a change in fluorescence" means any change in absorption properties, such as wavelength and intensity, or any change in spectral properties of the emitted light, such as a change of wavelength, fluorescence lifetime, intensity or polarization.

A "solid support comprising an array of multiple test compounds" or similar terms, mean a fixed matrix to which test compounds have been fixed. As an example, the solid support of the invention includes a membrane or other surface comprising an array of printed test compounds. In one specific embodiment of the invention, the test compounds are deposited as discrete spots on a porous track-etched polycarbonate membrane 10 to 20 microns thickness, the spots being between 10 microns to 2 mm diameter. The quantity of compound contained in each discrete spot will depend on the concentration of the stock solution from which it was derived, and the volume of that stock solution applied to the support. In another specific embodiment of the invention, compounds are printed onto a non-porous solid support which is optically clear.

By "test compounds" is meant a fixed array of compounds to be screened for ability to effect physiological parameters of a cell or tissue. In one embodiment, the test compounds are proteins or peptides generated by combinatorial protein chemical methods known to the art. In another embodiment, the test compounds are chemical compounds generated by combinatorial chemistry methods known in the art. In another embodiments, the test compounds are chemical compounds which are naturally occurring compounds more or less purified from their native state, are the products of genetically engineered cells, or are viral or bacteriophage particles engineered to display compounds upon their surfaces (phage display).

In one embodiment, the detector layer is an undemarcated area of living cells growing on a flat culture surface. The cells on this surface may or may not be grown to confluence,

may be transformed and/or engineered cells, or directly derived from animal tissues and grown as primary cell culture.

In one embodiment, a test compound reaches the detector layer by diffusion through a porous membrane to a liquid layer immediately overlaying the detector layer. A variety of commercially available porous membranes are useful in the method of the invention. A preferred porous membrane is a track-etched polyester or polycarbonate support in which parallel channels of identical size are formed by a selective etching process following exposure of the membrane to a source of high energy ions. The method of the invention allows each test compound affixed to a solid support to come into contact with a limited fluid volume, which fluid volume is in immediate contact with the detector layer. In one embodiment, each test compound contacts the detector layer by diffusion through a liquid-containing channel directly adjacent to the detector layer.

One advantage of the method of the invention is that it allows massive parallel screening of a large array of test compounds for biological activity. When living cells are the detector layer of the invention, they are maintained under physiologically viable conditions. Provision of these conditions requires the use of solutions able to supply essential nutrients and buffer pH changes normal to the continued growth of living cells. Such solutions may be complete cell culture media (i.e. any of those commercially available, for instance from Life Technologies Ltd.), optionally supplemented with antibiotics and serum preparations for optimal cell growth conditions. Buffer solutions may also be of the type known as "chemically defined". Cells will also require controlled temperature conditions, in the range 20° to 37°C, and the provision of gases essential to continued cell growth and maintenance of buffer capacity (O<sub>2</sub>, and optionally 5% CO<sub>2</sub>, depending on the type of buffer being used).

These and other objectives, advantages, and features of the invention will become apparent to those persons skilled in the art upon reading the details of the method as more fully described below.

#### BRIEF DESCRIPTION OF THE DRAWINGS

The foregoing features of the present invention may be fully understood from the following detailed disclosure of a specific preferred embodiment in conjunction with the accompanying drawings in which:

Fig. 1 is a schematic representation of the apparatus useful in one specific embodiment of the invention: Light from a high energy light source 1 is collected and collimated by unit 2, directed through a shutter assembly 3 and passes through a excitation filter-changer 4. A light guide 5 directs excitation light into the lensing and epi-illumination optics housed in unit 7. Excitation light emerging from 7 illuminates the horizontal detector layer located in the multi-component assembly having two solid layers 10 and 11 fixed relative to a supporting stage unit 8. Layer 11 is moved vertically downward on guide pins (17 Fig. 2b) controlled by arm 12 driven by unit 13. Four sprung contacts 14 attached to 12 press upon the frame of layer 11 to drive it downwards as arm 12 descends. Specified devices (3, 4, 9, 13, 15, 16) are controlled by central processing unit 6 which issues commands and collects data and status information from the devices attached to it. Unit 6 includes a central processing unit, RAM, multi-channel serial input/output cards with onboard A/D and D/A converters, one of which cards controls the camera 16 and captures images from it.

Figs. 2a-c: Figs. 2a and 2b are side view of the test stage (not to scale); Fig. 2c is a top view of the test stage. A supporting stage 8 has a rectangular central aperture the shape and size of which is the same as the area 19 of Fig. 2c. The position of stage 8 is adjusted in the horizontal and vertical axes by the 3-axis positioner 9. Components of the test stage shown include, solution layer 18, (not shown: detector layer 20 and array of test compounds 21 in Figs 3 and 4). The array 21 is held away from the liquid layer by pins 17 which pass through holes (24 in Fig. 5) in the corners of the frame 11. Arm 12 is moved down by the drive unit 13, and the four sprung contacts 14 it bears exert pressure on the frame 11 moving it down the guide pins 17 and into close proximity to the upper surface of 10, from which it is separated by a thin liquid layer 18.

Fig. 3 is a schematic showing the relative positions of the different layers in the testarray detector layers used in one specific embodiment. The layers are depicted in apposition, as they would appear after arm 12 has pushed component 11 down the support pins 17. An array of discrete spots of test compounds 21 on a porous membrane 19 is in contact with a liquid layer 18 overlaying the detector layer 20 which is supported by an optically transparent solid substrate 10. The compounds fill the parallel capillary spaces in the track-etched membrane 22.

Fig. 4 is a schematic drawing of a second embodiment of the screening method of the invention. The layers are depicted in apposition, as they would appear after arm 12 has pushed component 11 down the support pins 17. A detector layer 20 supported on an optically clear porous membrane 19, and overlayed by a liquid layer 23, is placed onto an optically clear solid substrate 10 bearing an array of test compounds 21. The thin space 18 between components 19 and 10 is filled with solution from 23 which has passed through the porous membrane 19. Bioactivity is detected by measuring changes in fluoresence of the detector layer resulting from responses to the diffusion of test compounds through the porous membrane to the detector layer.

Figs. 5a-c are schematics illustrating transfer printing of an array of compounds onto a surface of a track-etched membrane. Compounds are stored in 16 separate 96-well microtitre plates and defined amounts are transferred simultaneously by a 96-pin printing head to the surface 19 (Fig. 5a). The contents of each successive 96-well plate are printed at a slightly offset position, generating an array after 4 such printing operations (Fig. 5b), and a full array of 1536 compounds after 16 printing operations (Fig. 5c).

### **DETAILED DESCRIPTION**

Before the present method and solutions used in the method are described, it is to be understood that this invention is not limited to particular methods, components, or solutions described, as such methods, components, and solutions may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and

materials are now described. All publications mentioned herein are incorporated herein by reference to disclose and describe the methods and/or materials in connection with which the publications are cited.

Generally, the invention is drawn to a method for high throughput screening of test compounds, by contacting a solid support comprising an array of multiple test compounds with a detector layer, wherein each test compound comes into contact with a localized liquid which is in contact with a detector layer, and detecting a response of the detector layer to the test compound, wherein a bioactive test compound is identified.

The high density format screening system (HDFS) of the invention, rests in part on the realization that the delivery of test compounds to detector surfaces can be greatly simplified by doing away with the need for complicated microfluidics. Test compounds are applied to the detector surface in a massively parallel manner, and the method is applicable to a large range of different types of test compounds.

Central to the specific embodiments of the method and apparatus of the invention, described below, is the use of living cells as detectors, their responses being signalled via changes in the fluorescent or luminescent properties of various specific probes located within. However many different types of detector systems could be used in place of cells in such a system, for example, appropriate variants of Scintillation Proximity Assay (SPA) systems (Amersham Pharmacia Biotech) and enzyme-linked immuno-sorbent assay (ELISA) systems (Amersham).

#### Test Compound Arrays

The array of test compounds is formatted to have the same dimensions as the detector surface. In one specific embodiment of the invention, array and detector layers have a width of 8 cm and length of 12.5 cm, so as to fit within the format of conventional 96-well or 384-well microtiter plates. Preparation of the test arrays will depend on their origin.

Test compounds held in formatted arrays. Current methods for the production of single compounds by combinatorial methods are under development which involve miniaturization and patterned arrays of tethered solid-phase substrates. Thus, test compounds generated by combinatorial methods can be used to synthesize an array directly or indirectly on a carrier sheet. In one embodiment, vapor phase solubilization is used to produce a test compound array on the synthetic substrate, followed by a printing process of the test

compound array on to an absorbent membrane. In this embodiment, the test array is the printed membrane. An attractive feature of this method is that multiple copies of the same test array can be produced at one time to be screened against multiple cell systems for specific activities which minimizes stock handling from library archives.

Currently most compounds to be screened come in 96-well format. However, the 96-well format can be altered by repeated off-set printings, to any chosen density of format that the transfer substrate and assay can support. The optimum density of compounds in the test array will depend very much on the fraction of compounds in an array which generate bioactive responses in the detector layer ("hit rate"). The hit rate will depend on how well the compound library being tested matches the targets in the assay. If the hit rate is low, e.g., 1:20,000 - 100,000 compounds tested, a test array with center to center spacing of 200 µm (giving 240,000 separate compounds in a 12 cm x 8 cm area) may be preferable, providing 2 to 10 hits per plate. At a spacing of 1 mm, 9,600 test compounds may be screened simultaneously.

The density of the format may be adjusted as required without requiring any changes in the hardware used to perform the re-formatting; rather, adjustment may be made in the degree of off-set and the number of print operations used per array.

#### Detection

Fluorescent imaging provides a way to monitor physiological responses of living cells in a non-invasive manner. Ion- and voltage-sensitive probes, as well as the new generation of recombinant fluorescent probes, for instance, hybrid proteins comprising fusions of green fluorescent protein variants (GFPs) to cellular proteins involved in intracellular signaling, can be used singly or in combination to report on many aspects of cellular microphysiology. Due to the strong fluorescence of GFP, the luminescence of cells expressing the probes may easily be detected and analyzed by employing a combination of fluorescence microscopy and image analysis. Furthermore, these probes described are easily introduced into cells, as they can be expressed in the cells of interest after transfection with a suitable expression vector.

Recombinant probes for second messengers and enzyme activity, such as kinase activity, are not only useful in basic research but also in screening programs aiming at identifying novel biologically active substances. As an example, any currently used screening program designed to find compounds that affect cAMP concentration and protein kinase

activity are based on receptor binding and/or immuno detection and/or reporter gene expression. The recombinant probes described herein, on the other hand, make it possible to develop an entirely new types of screening assays able to monitor immediate and transient changes of cAMP concentration and protein kinase activity in intact living cells.

The HDFS method of the invention monitors the response of cell populations to test compounds. Lens systems are currently available which can simultaneously epi-illuminate and image the fluorescence from areas in excess of 8.5 x 13 cm, the size of a standard 96-well plate. The detection method used herein collects a variety of fluorescent signals from all cells in a field, with responses from discrete areas of the field being apparent in the real image of the fluorescence from that field as formed on the surface of the photosensitive detector (imaging camera).

#### Delivery of Test Compounds to Detector Cells

In a first embodiment of the method of the invention, delivery of large arrays of test compounds to cells is achieved with test compounds which are present on or transferred to a porous carrier sheet. In specific embodiments, test compounds are printed on the carrier sheet, and the sheet is applied (overlayed) to a field of cells of the same area. The test compounds reach the detector cells by diffusion through a localized buffer layer immediately in contact with an area of the detector cell layer. This embodiment is shown in the schematic of Figs. 2 & 3.

Porous carrier sheet for delivery of test compounds: Test compound arrays are fixed onto the porous carrier sheet by a variety of methods known to the art. For example, an array of test compounds may be transferred and fixed to the carrier sheet by the method of contact printing, whereby an array of inert flat-ended pins (e.g. made of stainless steel) is used to transfer defined volumes of individual test compounds (in the range 50 nl to 2  $\mu$ l) in solution form to discrete points on a dry carrier sheet.

A porous membrane useful in the delivery of test compounds is a membrane constructed of a non-absorbent material with pores of regular and defined diameter which traverse the membrane directly from the upper to the lower side. The property of orthogonal capillarity is useful in these membranes to limit lateral spread of test compounds applied to the membranes as discrete spots of liquid, since it is important that the compounds remain as discrete spots upon the membrane. A variety of membranes of different thicknesses,

materials, and pore densities are commercially available from a number of manufacturers. For example, porous membranes useful in the method of the invention include a track-etched polycarbonate or polyester membrane (Corning Costar or Whatman/Polyfiltronics). These are available in thicknesses from 6 to 23 microns, with pores of 14 to 0.015 microns, at 100,000 to 1,000,000,000 pores/cm<sup>2</sup>. For delivery of test compounds with maximum ease of handling and loading of test compounds, polycarbonate membranes are preferred, particularly of a thickness of greater than 10 microns, with pores between 1 and 10 microns diameter at densities of between 20,000,000 to 100,000 pores/cm<sup>2</sup>, respectively. One preferred membrane is Nucleopore® from Corning Costar.

Alternative membranes useful for the delivery of compounds include cast cellulose acetate (Membra-fil®), PTFE membranes (e.g. Filinert™), and glass fiber filters, all available from Corning Costar. These thicker membranes encourage lateral spread of liquid samples applied to their surfaces, but are thicker and could thus be used to deliver larger amounts of compounds.

Track-etched and cast cellulosic membranes may also be given hydrophilic or hydrophobic surface treatments. It is useful to have membranes whose surfaces have defined wettability properties.

When the test compound is soluble, the compound will dissolve into the buffer upon contact with the buffer medium, and directly contact the detector layer immediately underlying the buffer layer. In this embodiment, the test compounds dissolve upon contact with the buffer medium, and fall vertically onto the detector layer as a result of having a higher density than the surrounding liquor. It is generally preferred that the thin buffer layer between the test compound membrane and detector layer not be stirred significantly by convection. At the detector layer, the vertical fall of a solution of test compound is expected to spread radially by displacement and diffusion. The radial extent of a measured response may thus be use as an indicator of the bio-potency of the compounds involved.

Test compounds of limited solubility, such as those expressed on the surface of a carrier system, for instance, a cell membrane, viral or phage particle, must be brought into very close proximity, including direct contact, with the detector layers.

<u>Buffer and Detector layer</u>. The detector layer may be a continuous or non-continuous layer of living cells. In a specific embodiment, the detector layer is a continuous cell monolayer corresponding in size to the test compound array. In more specific embodiments,

thin glass substrate, suitably tissue culture treated is preferred for fluorescent probes requiring excitation wavelengths below 400 nm.

Living cells are maintained under physiologically viable conditions, as defined by such parameters as oxygen consumption, membrane potential, mitochondrial potential and cytoplasmic ion balance. Provision of these conditions requires the use of solutions able to supply essential nutrients and buffer pH changes normal to the continued growth of living cells. Such solutions may be complete cell culture media (i.e. any of those commercially available, for instance from Life Technologies Ltd.) optionally supplemented with antibiotics and serum preparations for optimal cell growth conditions. Buffer solutions may also be of the type known as "chemically defined" (e.g. phosphate buffered saline solutions). Cells will also require controlled temperature conditions, in the range 20° to 37°C, and the provision of gases essential to continued cell growth and maintenance of buffer capacity (O<sub>2</sub>, and optionally 5% CO<sub>2</sub>, depending on the type of buffer being used).

Detection of bioactivity. Detection of bioactivity may be determined by a number of methods known in the art. In a preferred embodiment, detection of bioactivity is determined by cellular imaging of fluorescence. For example, imaging may be conducted of a cell layer on a clear glass substrate. A glass substrate having a surface pitted with a regular array of very shallow (approx  $20~\mu m$ ) depressions may be used for this purpose (Corning). This glass substrate is useful because it ensures a regular and defined spacing between the overlying test array and the cells beneath.

In one embodiment, the detector layer is an undemarcated area of living cells growing on a flat culture surface. The cells on this surface may or may not be grown to confluence, may be transformed and/or engineered cells, or directly derived from animal tissues and grown as primary cell culture. In a second embodiment of the method of the invention, the array of test compounds is laid out onto a non-porous substrate (such as thin coverglass sheet) which is transparent or optically clear. Imaging will be through this surface, and through the cell support membrane lying above. The substrate (Fig. 4, 10) should be inert and solvent tolerant. For example, borosilicate glass sheets of about 200 microns thickness, which may be further surface-treated to give either hydrophobic or hydrophilic properties as desired. This embodiment is shown in the schematic of Fig. 4.

Detector layer: In one embodiment of the invention, the detector layer is a layer of

living cells cultured on a thin porous membrane. A porous membrane useful in the culture and transfer of cells is a transparent non-absorbent membrane with pores of regular and defined diameter which traverse the membrane directly from the upper to the lower side. A porous sheet suitable for cell growth is a track-etched polyester membrane about 10 microns thick with pores between 0.015 and 5 microns diameter at densities of between 600,000,000 to 400,000 pores/cm<sup>2</sup> repectively (Nucleopore® from Corning Costar).

Delivery of test compounds to detector layer. The porous membrane which supports the detector layer, complete with the buffer medium which overlays it, is applied onto the (dry) test array. Buffer medium wets the lower surface of the porous membrane (Fig. 4, 19) and forms a continuous thin film 23 between the array of test compounds 21 and the porous membrane 19. Test compounds diffuse up through the pores to the detector layer above. In one embodiment of the invention the detector layer is a monolayer of living cells overlayed with physiological buffer solution. The invention includes the possibility that under some conditions it is desirable to have cells grow processes through the membrane to make direct contact with substances on the test array below, with the use of a membrane having an appropriate pore diameter.

Further embodiments and general considerations. Where a test array is generated as a complex mixture of components, such as from the "teabag" method of combinatorial synthesis, or from cDNA library expression systems, a separation step may first necessary. Separation of test components may be conducted in any number of ways known to the art. In one embodiment, components may be separated by the use of one- or two-dimensional separation techniques in non-denaturing gels. The resulting gels may be used directly as test arrays.

Specific separation methods will be tailored to the components involved. Any bioactive compounds from such an array would be identified from identical copies of the original test gel.

### Detection of Bioactivity.

Lens and illumination system. Specialized light sources and optics are needed to illuminate and image the fluorescence coming from an area the size of a microtiter plate (96-well plate). Such a system is available from: Imaging Research Inc., St Catherines, Ontario, Canada, and consists of a high-power light source directed through a specialized lens which

acts both as a wide-field epi-illuminator and imaging device.

An illumination system useful in the HDFS device is able to deliver excitation light over an area of at least 8.5 by 13 cm at an intensity sufficient to excite measurable fluorescence from that test field (which in most cases will be living cells loaded with fluorescent reporters). The illumination may come from a scanned beam, or be wide-field for simultaneous illumination of the entire area. The imaging system will collect fluorescent light from the entire test area and bring it to focus onto a sensitive imaging photodetector, such as a cooled CCD camera chip.

Screening. The practice of screening large libraries of samples of unknown composition for the few which may contain a compound of specific biological activity is one of the more common methods of new drug discovery. The samples of unknown composition are in most cases biological material, such as plant extracts or microbial fermentation broths. Screening these for biological activity is normally accomplished by performing binding assays or, more recently, functional assays. A binding assay is an attempt to find compounds of interest by identifying those which adhere with some desired affinity to cells or cell products. This can be done using fluorescent, luminescent, or radioactive detection methods. These assays are based not on a biological response, but passive processes of adherence and displacement. They cannot be construed as functional assays or as real-time assays. Another way to determine biological activity is to measure up-regulation or down-regulation of expression of a known gene. This is done by inserting DNA which codes for something which can be readily measured into a cell's genome such that the expression of interest is coupled to expression of the inserted DNA. While this is a true functional assay, it also is not a real time assay. In addition, it is only capable of finding compounds which affect gene expression. In many cases this is not the response of interest.

The CytoSensor described in U.S. Patent No. 4,915,812 and U.S. Patent No. 5,395,503 is a commercial instrument which has been billed as a screening instrument. It is based on the detection of increased cellular proton flux by means of a semiconducting electrode. The instrument is applicable to high through-put screening, but can only detect cellular events that result in changes in extracellular pH. Again, many responses of interest are not associated with changes in extracellular pH.

The growth over the last few decades in the knowledge of cellular signaling has presented extremely rich opportunities for new ways of screening for biologically active

compounds. Armed with knowledge of the biological process which one wants to affect with a new product, it is possible to monitor the actual process as a way of looking for compounds which affect it. The development of fluorescent probe molecules which upon interaction with intracellular signaling molecules (e.g. ions, enzymes, cyclic nucleotides) change their spectral properties has enabled the real-time monitoring of dynamic biological responses within living cells. Most of these probes can be introduced non-invasively into cells and will, depending on the detection system, allow characterization of cellular events in high temporal resolution (microseconds to seconds) and high spatial resolution (nanometers to micrometers). This probe technology, in combination with the technology of cellular imaging which is described below, has had a major impact on cell biology in that it has enabled monitoring of complex, cross-reacting intracellular events that could not be unravelled by conventional invasive biochemical techniques.

Imaging of cellular functions using luminescent probes. Visualization of intracellular function using luminescent (fluorescent or bioluminescent) probes has become one of the mainstay techniques in modern cell biology. Using traditional optical microscopes with quantitative detectors in place of the human eye, both the concentration and distribution in the cell of a variety of intracellular molecules of interest can be measured. While luminescent probes can be measured in large populations of cells using other techniques, imaging is the only way to learn what is going on in single cells or small populations of cells. The imaging capabilities of the HDFS apparatus will be limited to rather low spatial resolution - fluorescent changes will be imaged from the entire field of detector layer up to 8cm by 12.5 cm. When the detector layer comprises living cells, individual cells need not be resolved in the image, only the fluorescent signals from regions in which cells are present.

The imaging times will vary depending on the responses and parameters being monitored. Signaling responses, for instance changes in the level of free calcium in cellular cytoplasm, may first be seen within seconds or minutes following delivery of test compounds to the detector layer. Such changes can be monitored by changes in the fluorescent properties of specific chemical probes, for instance Fluo-3 or Fura 2 may be used to report on cytoplasmic calcium. The way in which these changes develop within cells (time-response profile) is an important diagnostic feature of the signaling processes giving rise to them. Rapid responses are therefore recorded by sequences of images, where the time between images in a sequence is between 0.1 and 30 seconds (depending on the response being

screened for). Transcription mediated events may require minutes to hours to develop.

Monitoring may be continuos or intermittent. For slow responses, two images can be sufficient to gauge the level of response, the first taken before application of test compounds, the second after a period during which the response is estimated to have reached its maximum extent.

Controls relevant to the parameters being measured can be incorporated into the test arrays, both as a check for cell responsiveness and as co-ordinate markers within the arrays. The detector layer is continuous and undemarcated, but because of the close apposition of the test array to the detector layer, the center point of a response in the detector layer corresponds to a conjugate coordinate in the test array. It is helpful to have compounds in the test array which will generate known responses at known coordinates in the detector layer. Responses at the conjugate coordinates in the detector layer act as controls for the system's response, against which responses of the detector layer to unknown compounds may be compared; the points of response to control substances also act as reference points in the detector layer from which the coordinates of other responses can be mapped. For example, when bioactivity is determined as the ability to alter the level of free calcium in cellular cytoplasm, common calcium-mobilizing agonists such as carbamylcholine or adenosine trisphosphate are included in the test array at known coordinates.

As another example, when a change in the cellular ratio of inherently fluorescent NAD(P)H/FAD is the biological parameter being assayed, metabolic inhibitors such as KCN or rotenone may be used as a control and marker compounds.

In many instances, diffusion within a thin fluid layer will be involved in many applications of the screening method of the invention, and a concentration gradient will be established from each test point. Those few compounds in a test array which have bioactivity should be detected as spreading rings of response from the focus point of diffusion, within a field of the detector showing no response. The extent of the response areas (measured over time), compared with those from control substances, will provide an indication of potency and solubility of the compound responsible, and also obviate the need to make serial dilutions of test compounds. Toxic or inhibitory substances may also be determined by causing blank sectors in response rings from known agonists. Inhibitory compounds may be determined by their actions on a (pre-)stimulated detector field. Detection of bioactive compounds may incorporate simple image processing to determine the focus, extent and potency/efficacy from

the areas of activity measured in a detector field.

### **Apparatus**

In specific embodiments, the apparatus and method of the invention are as shown in Figs. 1-4. Fig. 1 shows a high energy light source 1, either a mercury or xenon arc lamp, light from which is collected and collimated by unit 2, directed through a shutter assembly 3 and passes through a excitation filter-changer 4. A high-quality light guide 5, either of fused quartz or a UV-compatible liquid light guide, directs excitation light into the lensing and epi-illumination optics housed in unit 7. Excitation light emerging from 7 evenly illuminates the horizontal detector layer located in the multi-component assembly labeled 10 and 11.

Further details of this assembly are shown in Figs. 2a-c, 3, and 4. The assembly comprises two solid layers of which 10 is fixed relative to the stage unit 8 which supports it, while layer 11 is moved vertically downward on guide pins (17 in Figs. 2a,b,c) to bring test compounds into contact with the detector layer. Vertical movement of 11 is controlled by arm 12 driven by unit 13. Four sprung contacts 14 attached to 12 press upon the frame of layer 11 to drive it downwards as arm 12 descends. A separate drive unit 9 controls position of the stage 8 in the horizontal plane, and also is used to adjust focus by movement along the vertical axis.

Fluorescent light emitted by the detector layer is collected by lensing unit 7, passes through an emission filter-changer 15 and is brought to focus on the photosensitive surface of an imaging detector housed in unit 16.

Specified devices (3, 4, 9, 13, 15, 16) are controlled by a central processing unit 6 which issues commands to, and collects data and status information from the devices attached to it. Collected data (images) can also be analyzed by unit 6, or passed to a subsidiary analysis station (not shown). Unit 6 comprises: central processing unit (Intel Pentium chip, or better), RAM, multi-channel serial input/output cards with onboard A/D and D/A converters, one of which cards controls the camera 16 and captures images from it, also a video controller card, VDU, and hard disk memory units.

Figs. 2a,b,c are schematic diagrams of the test stage, which includes a supporting stage 8 with large rectangular central aperture, the shape and size of which is the same as the area labeled 19. The position of stage 8 is adjusted in the horizontal and vertical axes by the

3-axis positioner 9. These diagrams are drawn for the specific embodiment in which the detector layer is a layer of living cells growing on the upper surface of the solid transparent component 10, which also serves to contain the liquid layer 18 which overlays the cells in the detector layer and provides them with necessary nutrients and conditions to keep them alive. The printed array of test compounds 21 is borne on a sheet of track-etched membrane 19 held by a rectangular rigid frame 11. At the beginning of the screening assay, the array 21 is not in contact with the fluid layer 18. The array 21 is held away from the liquid layer by pins 17 which pass through holes 24 in the corners of the frame 11 and which, by friction or "click-stops", prevent it from falling (Fig. 2a). At the appropriate moment, arm 12 is moved down by the drive unit 13 and the four sprung contacts it bears 14 exert pressure on the frame 11 moving it down the guide pins 14 and into the liquid 18 below to a position where it is in very close proximity to the underlying layer of detector cells 20 grown on top of the solid substrate 10 (Fig. 2b). Throughout this procedure, the entire area of the detector layer corresponding to the size and shape of area 19 is illuminated and imaged from below by the additional apparatus shown in Fig. 1.

The apparatus can also be used in a second embodiment of the screening method of the invention, where the test array is laid out on the upper surface of component 10, and components 11 and 19 are a frame and thin transparent track-etched membrane, respectively. In this specific embodiment, the frame 11 is sufficiently deep to contain culture liquid as required to sustain the detector layer of living cells growing on the upper surface of the membrane 19.

Figs. 3 and 4 are schematics to show the relative positions of the different layers in the test-array/detector layers used in the specific embodiments of the invention. Fig. 3 shows the arrangement in which an array of discrete spots of test compounds 21 on a porous membrane 19 is in contact with a liquid layer 18 overlaying the detector layer 20 which is supported by an optically transparent solid substrate 10. The compounds fill the parallel capillary spaces 22 in the track-etched membrane 19. Bioactivity is detected by measuring changes in fluorescence in the detector layer 20 resulting from responses to the diffusion of test compounds through the porous membrane to the detector layer.

Fig. 4 is a schematic drawing of a second embodiment of the screening method in which a detector layer 20 supported on an optically clear porous membrane 19, and overlayed

by a liquid layer 23, is placed onto an optically clear solid substrate 10 bearing an array of test compounds 21. The thin space 18 between components 19 and 10 is filled with solution from 23 which has passed through the porous membrane 19. Bioactivity is again detected by measuring changes in fluorescence of the detector layer resulting from responses to the diffusion of test compounds through the porous membrane to the detector layer.

Fig. 5 is a schematic illustrating the way in which an array of 1536 compounds can be created on a membrane surface, such as would be useful in the first embodiment described above, by simple transfer printing. Compounds are stored in 16 separate 96-well microtiter plates and defined amounts are transferred simultaneously by a 96-pin printing head to the surface 19. The contents of each successive 96-well plate are printed at a slightly offset position, generating an array as shown in Fig. 5b after 4 such printing operations, and a full array of 1536 compounds (Fig. 5c) after 16 printing operations. The holes 24 in frame 11 are used to position and guide the completed array on the pins 17 indicated in Figs. 2b and 2c. The process illustrated in Fig. 5 can also be used to transfer an array of test compounds to a solid surface such as would be useful for component 10 in the second embodiment of the method described above.

### **EXAMPLE**

### Example 1. Screening of 1536 Test Compounds for Bioactivity.

The following description of the use of one embodiment of the apparatus of the invention in the screening method disclosed. An array of test compounds are supplied in 96-well microtiter plates, as is common practice for compounds produced by methods commonly known as combinatorial chemistry, or for compounds extracted from natural sources. In this example, the compounds are provided in soluble form, and the concentrations and solvents used have previously been tested for compatibility with the apparatus. In this example, 1536 compounds are tested simultaneously against a known cellular target, specifically a G-protein coupled receptor (GPCR) of the Gq type expressed in a transformed cell line. Gq GPCRs give clearly identifiable changes in intracellular calcium when activated.

First, physiologically viable living cells are cultured to a near confluent monolayer in a transparent culture dish (10, Fig. 2a-c) in appropriate culture medium and conditions.

Immediately prior to being used in the experiment, the cells are loaded with the fluorescent

indicator of free cytoplasmic calcium concentration, Fluo-3 (from Molecular Probes, Oregon). This is accomplished by incubating the cells with a 2 to 5  $\mu$ M solution of Fluo-3 acetoxymethyl ester (AM) for a period of 10 to 15 minutes, followed by a series of solution exchanges to wash away excess Fluo-3 AM.

The method of transfer of compounds to the track-etched membrane Fig. 2a-c 19 is illustrated in Fig. 5. In this example, 1536 compounds are printed as an array 21 on a single track-etched membrane 19, from sixteen individual 96-well microtiter plates in the following manner: A 96-pin printing head is used to transfer defined volumes of compounds (in the range 0.05 to 0.5 µl of each compound), one compound per pin, from each 96-well plate in turn (with wash steps between source plates to avoid cross-contamination). Each 96-point print to the membrane occurs in an offset grid, such that 16 print operations are made sequentially on the same membrane and the printed spots of compounds remain discrete and separated from each other (three of these spots are indicated in Fig. 5a, 21). Fig. 5a shows the result of a single 96-point print operation, Fig. 5b after four such operations, and Fig. 5c the finished array after 16 print operations. In this way, just sixteen print operations (and sixteen intermediate wash steps for a single print head) are sufficient to transfer 1536 compounds to a single test array. The procedure can be readily automated, and multiple copies of each printed sheet made for multiple tests.

Completed arrays are fixed to the pins 17 (Figs. 2b-c) projecting from the culture dish 10 such that they are supported some small distance above the thin fluid layer 18 covering the living cells which form the detector layer. Once the test array is fixed in place over the Fluo-3-loaded cells, the entire assembly is placed onto the test stage as shown in Fig. 2a.

The following events are synchronized by sequential instructions from the computer processing unit 6. First, the test stage is centered over the lensing unit 7 (Fig. 1) and the detector layer it supports is brought into focus by the motor unit 9. Fluo-3 is excited by light of 490 nm, and its fluorescent emissions are collected in the range 505-540 nm. The intensity of emission is increased when the dye binds free calcium. Thus the computer brings a 490 nm band-pass excitation filter into line of the light path coming from units 1 and 2 using the filter changer unit 4. At the same time, a band-pass emission filter for the range 505-540 nm is positioned in the imaging path by unit 15. The shutter 3 is opened for a pre-determined exposure period (typically 50 to 500 milliseconds), and during this time the whole area of the

detector layer is illuminated with 490 nm light. Fluorescent emission from the Fluo-3 in the cells is collected by the lens 7 and focused into the camera. The camera captures the image and sends it to the processing unit 6 where it is stored and displayed. At regular intervals thereafter, images are captured in sequence by repeatedly opening the shutter 3. Intervals between successive images are typically in the range 0.5 to 30 seconds, depending on the speed of the response expected. Intervals of 0.5 to 2 seconds are usual and sufficient to sample the dynamics of most changes in cellular calcium. At a predetermined time during this continuing sequence of images, the test array is pushed down the guide pins 17 by the actuating arm 12 and its sprung contacts 14, driven by unit 13. In close apposition to the cells in the detector layer, the test array begins to release the compounds it carries. The compounds dissolve into the the liquid layer, and these fall vertically downwards onto the cells below. Because there is only a thin liquid layer between the membrane of the test array and the cells below, there is insignificant intermixing of adjacent test compounds. If a test compound activates cells below it bearing Gq GPCRs, these cells will respond with an immediate increase in free cytoplasmic calcium, and the fluorescence signal from the Fluo-3 dye they contain will increase. The sequence of images collected during the period of the response (which is typically of 1 to 10 minutes duration) will reveal which cells have so responded, and their position in the area of the detector layer will be correlated with the identity of the compound in the array above. An analysis of the entire area of each image in the sequence, performed on-line by the processing unit 6, yields the following information: the identity of the compound eliciting the response, the profile of the response with time, the intensity of the response, and also the potency of the compound with reference to a chosen standard. The latter information is contained in the radius of the area of cells responding within a particular time, and can be compared directly to a known standard which is included in the array at known points. The use of standard compounds at known points in the array also provides a control for the experiment, and helps to identify coordinates in the detector layer from which other responses can be mapped.

At the end of the screening assay, the sequence of images is stopped, the actuating arm 12 raised, and the test assembly removed. The next assembly is then moved in and the sequence begun afresh. Assembling the test units and exchanging them on the test stage can be automated by appropriate robotic control (not shown in the diagrams).

One of the advantages of the method of the invention is that the method does not require that either the components of the detector layer (e.g. living cells), or the different test compounds, be isolated from one another within discrete chambers or compartments, as is common to all high throughtput screening procedures currently in use or development. The method also removes the need to dispense microvolumes of test compounds during the period of the assay itself. Delivery of test compounds to detector layers is either by direct contact or by simple diffusion across thin liquid films. Delivery and detection becomes a (massively) parallel process.

#### **CLAIMS**

#### What is claimed is:

- 1. A method for screening test compounds for bioactivity, comprising:
- (a) contacting an array of test compounds with a detector layer; and
- (b) detecting a detector layer response, wherein a response is indicative of bioactivity.
- 2. The method of claim 1, wherein the detector layer is comprised of physiologically viable cells.
- 3. The method of claim 2, wherein the detector layer is supported by an optically clear substrate.
- 4. The method of claim 3, wherein the reactive sensing surface is held stationary in the field of view of the optical detector and the sample surface is moved into contact with it during the course of the measurement.
- 4. The method of claim 1, wherein the detection of step (b) is a change in a fluorescence or luminescence property of the cell.
- 5. The method of claim 4, wherein detection is determined with an illumination system capable of exciting the fluorescence of the reactive surface with any of a number of previously selected wavelengths with defined order and of defined time duration.
- 6. The method of claim 2, wherein the physiologically viable cells form a monolayer.
- 7. The method of claim 1, wherein the test compounds are generated on a solid support by combinatorial chemistry.
- 8. The method of claim 1, wherein the test compound array is generated by one- or twodimensional gel electrophoresis.



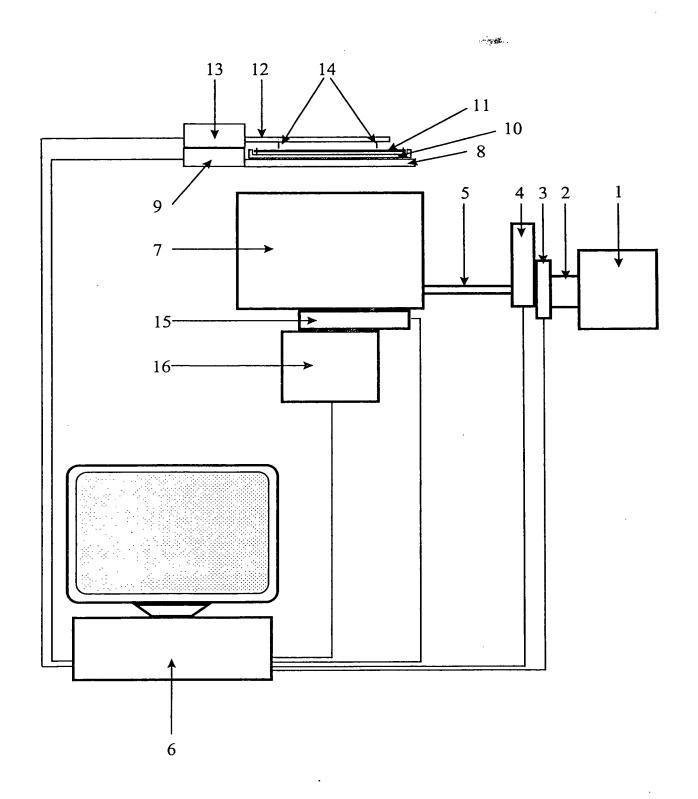
- 9. A method for high throughput screening of test compounds for bioactivity, comprising:
- (a) contacting a solid support comprising an array of multiple test compounds with a cell layer, wherein each test compound comes into contact with a localized liquid which is in contact with a detector layer; and
- (b) detecting a response of the detector layer to the test compound, wherein a response is indicative of a bioactive compound.
- 10. A method for simultaneously exposing an array of test compounds with a reactive sensing surface, comprising the steps of:
- (a) contacting an array of test compounds on a solid matrix with a porous membrane which is in contact with a liquid layer overlaying a reactive sensing surface layer; and
- (b) allowing the test compounds to diffuse through the porous membrane to the liquid layer overlaying the reactive sensing surface.
- 11. An apparatus for screening an array of test compounds for bioactivity, comprising:
  - (a) a solid support comprising an array of test compounds;
  - (b) a porous membrane; and
- (c) a detector layer layer, wherein a liquid layer is between the porous membrane and detector layer layer, and wherein the test compounds contact the detector layer layer by diffusion through the porous membrane.

## METHOD AND APPARATUS FOR HIGH DENSITY FORMAT SCREENING FOR BIOACTIVE MOLECULES

### Abstract

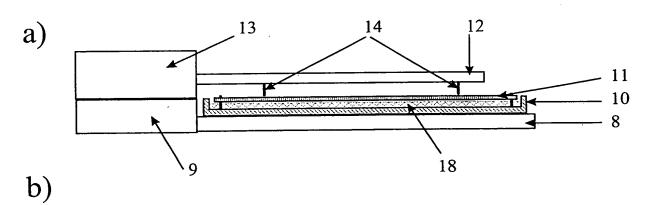
A method and apparatus for screening an array of test compounds for bioactivity by contacting an array of test compounds with a detector layer capable of detecting bioactivity, and detecting a detector layer response. The detector layer is comprised of physiologically viable cells. The method and apparatus allow a large number of test compounds to be simultaneously assayed in parallel.

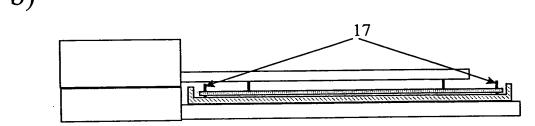
Fig. 1 Schematic view of equipment; not to scale

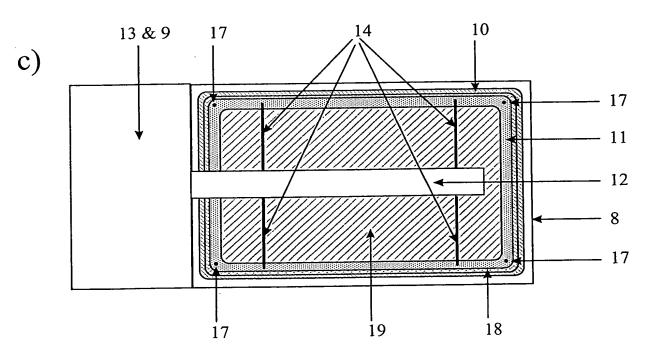


### Fig. 2

### Side views of test stage; not to scale



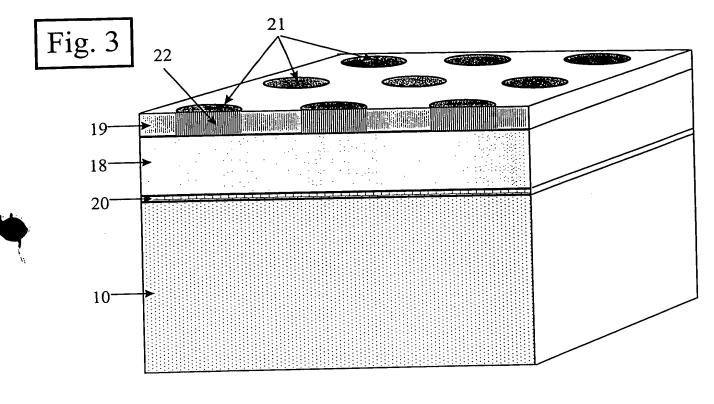


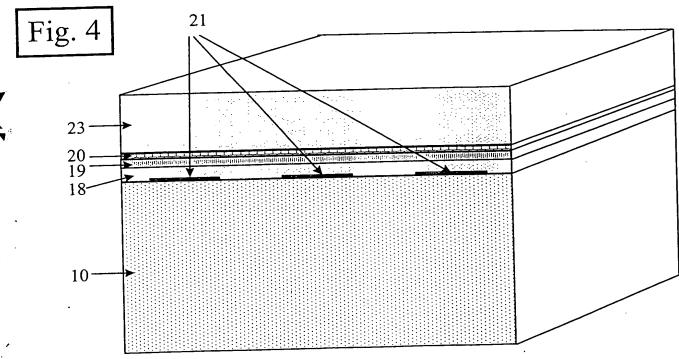


Top view of test-stage; not to scale

22129DK1 Appendix II

# 3-D sectional representations of portions of the test-array/detector layers: not to scale





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